

Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.4251/wjgo.v8.i1.30 World J Gastrointest Oncol 2016 January 15; 8(1): 30-39 ISSN 1948-5204 (online) © 2016 Baishideng Publishing Group Inc. All rights reserved.

REVIEW

Antitumor effects of the benzophenanthridine alkaloid sanguinarine: Evidence and perspectives

Roberta Gaziano, Gabriella Moroni, Cristina Buè, Martino Tony Miele, Paola Sinibaldi-Vallebona, Francesca Pica

Roberta Gaziano, Gabriella Moroni, Cristina Buè, Martino Tony Miele, Paola Sinibaldi-Vallebona, Francesca Pica, Department of Experimental Medicine and Surgery, University of Rome Tor Vergata, 00133 Rome, Italy

Author contributions: Gaziano R, Moroni G, Buè C, Miele MT, Sinibaldi-Vallebona P and Pica F contributed to this paper.

Conflict-of-interest statement: Authors declare no conflict of interest for this article.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/ licenses/by-nc/4.0/

Correspondence to: Francesca Pica, MD, PhD, Department of Experimental Medicine and Surgery, University of Rome Tor Vergata, Via Montpellier, 1, 00133 Rome, Italy. pica@uniroma2.it Telephone: +39-6-72596462 Fax: +39-6-72596550

Received: June 12, 2015 Peer-review started: June 15, 2015 First decision: August 25, 2015 Revised: October 9, 2015 Accepted: November 3, 2015 Article in press: November 4, 2015 Published online: January 15, 2016

Abstract

Historically, natural products have represented a significant source of anticancer agents, with plant-derived drugs becoming increasingly explored. In particular, sanguinarine is a benzophenanthridine alkaloid obtained from the root of Sanquinaria canadensis, and from other poppy Fumaria species, with recognized anti-microbial, anti-oxidant and anti-inflammatory properties. Recently, increasing evidence that sanguinarine exibits anticancer potential through its capability of inducing apoptosis and/or antiproliferative effects on tumor cells, has been proved. Moreover, its antitumor seems to be due not only to its pro-apoptotic and inhibitory effects on tumor growth, but also to its antiangiogenic and anti-invasive properties. Although the precise mechanisms underlying the antitumor activity of this compound remain not fully understood, in this review we will focus on the most recent findings about the cellular and molecular pathways affected by sanguinarine, together with the rationale of its potential application in clinic. The complex of data currently available suggest the potential application of sanguinarine as an adjuvant in the therapy of cancer, but further pre-clinical studies are needed before such an antitumor strategy can be effectively translated in the clinical practice.

Key words: Sanguinarine; Cancer; Apoptosis; Cell-cycle; Chemotherapy

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Sanguinarine is a benzophenanthridine alkaloid isolated from the root of *Sanguinaria canadensis*, and other poppy *Fumaria* species, which exibits a clearcut anticancer potential by inducing apoptosis and/or antiproliferative effects on tumor cells. Sanguinarine also shows antiangiogenic and anti-invasive properties, as demonstrated *in vitro* and *in vivo*. In consideration of the multiple biological effects of sanguinarine, which suggest its possible use in cancer therapy, further detailed pharmacokinetic and toxicologic studies are required to assess both the efficacy and safety of the compound before proposing a possible translation into the clinic.



Gaziano R, Moroni G, Buè C, Miele MT, Sinibaldi-Vallebona P, Pica F. Antitumor effects of the benzophenanthridine alkaloid sanguinarine: Evidence and perspectives. *World J Gastrointest Oncol* 2016; 8(1): 30-39 Available from: URL: http://www.wjgnet. com/1948-5204/full/v8/i1/30.htm DOI: http://dx.doi.org/10.4251/wjgo.v8.i1.30

INTRODUCTION

Tumor initiation is the result of multiple genetic and epigenetic events. Transformed cells are characterized by indefinite proliferation, apoptosis-resistance and the capability to metastasize and support angiogenesis^[1].

Chemotherapy, irradiation and/or immunotherapy represent the gold standard approach for the treatment of cancer worldwide. The increased frequency of tumor relapse and the toxicity of the anticancer drugs, however, often reduce the therapeutical effectiveness of several antitumor therapy protocols. Therefore, the identification of more effective therapeutic protocols is needed and, in this direction, phytochemicals may represent an attractive alternative because of their low toxicity and low cost^[2]. In this scenario, sanguinarine (Figure 1) and chelerythrine are the principal members of quaternary benzo[c]phenanthridine alkaloids (QBAs)^[3] obtained from Sanguinaria canadensis, Chelidonium majus, and Macleava cordata. Alkaloids include a large group of secondary metabolites (SMs) that differ in relation to structure, function and biodistribution^[4]. In the past, QBAs have attracted the attention of many pharmacologists because of their own low toxicity^[5,6] and their multiple biological activities, such as the antitumor^[7], antimicrobial^[8,9], anti-inflammatory^[10], anti-HIV^[11], anti-platelet^[12], anti-angiogenesis^[13], and antiparasitic activities^[14-16]. The influence of QBAs on the activity of various important biological enzymatic pathways has been also demonstrated^[7]. For long times, sanguinarinecontaining herbs were believed to possess anticancer activity but only recently evidence that sanguinarine possesses a strong anti-neoplastic activity, which is mediated mainly by the induction of tumor cell apoptosis has been proved.

This review summarize the most recent findings on the molecular mechanisms underlying the antitumor activity of sanguinarine both *in vitro*, in a variety of human tumor cells, and *in vivo* in selected experimental tumor models, together with the rationale of its potential application in clinical practice.

SANGUINARINE INDUCES APOPTOSIS IN TUMOR CELLS

Physiologically, the human body controls homeostasis by eliminating damaged and aged cells by means of a genetically programmed process named apoptosis^[17,18]. Tumor cells evade apoptosis and grow indefinitely. Several proteins, among which are caspases, proapoptotic Bax and anti-apoptotic B cell lymphoma (Bcl)-2, cytochrome c, and apoptotic protease activating factor -1, carry out the apoptotic programme either by intrinsic or extrinsic pathways. The first one is dependent on mitochondria, whereas the second one is initiated by the so-called death receptors (DRs). Selected anti-apoptotic proteins, among which Bcl-2, have been found over-expressed in different types of cancers. The down-regulation of anti-apoptotic proteins in cancer cells represents a promising therapeutic strategy of intervention in cancer therapy.

A number of plant-derived agents, have been shown to be capable of hampering disease progression by inducing cell apoptosis in multiple types of human and experimental cancers. Recently QBAs, and particularly sanguinarine, have been indicated as potential anticancer compounds. In detail, it has been reported that micromolar concentrations of sanguinarine are capable of inhibiting tumor cell growth, and this inhibitory effect is associated with cell cycle arrest and induction of apoptosis^[19-22]. The anti-proliferative and/ or pro-apoptotic activities of sanguinarine have been demonstrated in in vitro studies on several cancer cell types including epidermal^[23], keratinocyte^[24,25], prostate^[26-28], cervical^[29], breast^[20,30-33], leukaemia^[34,35], lymphoma^[36], melanoma^[37-39], colon^[40,41], colorectal^[21], gastric^[42], pancreatic^[19], lung^[22], neuroendocrine^[43], osteosarcoma^[44], and human neuroblastoma cells^[45]. By contrast, there are few studies on the in vivo effectiveness of sanguinarine administration per os^[46,47] in animal tumor models^[33,48].

It has been reported that sanguinarine exerts an antiproliferative activity on murine melanoma cells both in vitro and in vivo (B16 melanoma 4AS in the syngeneic host C57BL/mice), as well as in A375 human melanoma xenografts in athymic nude mice^[48]. We also have conducted a study aimed at evaluating the anti-tumor effect of sanguinarine both in vitro and in vivo in a rat colorectal cancer model (DHD/K12/TRb cell line)^[49]. We found that the *in vitro* addition of sanguinarine has a dose-dependent inhibitory effect on the proliferation of DHD/K12/TRb cells and induces tumor cell apoptosis. Sanguinarine also showed a clearcut in vivo anti-tumor activity, leading to an inhibition of tumor growth higher than 70%^[49]. The sanguinarineinduced inhibition of tumor growth was associated with its pro-apoptotic effect on tumor cells, as confirmed by the ex-vivo histopathological examinations performed on experimental tumor sections and by TUNEL assay^[49].

It is known that sanguinarine-induced apoptosis occurs through multiple pathways, including the activation of nuclear factor- κ B (NF- κ B)^[50], the mitochondrial damage resulting in activation of the caspase machinery^[24] and the cell cycle arrest^[27]. In detail, the sanguinarine-induced apoptosis occur either *via* a mithocondrial pathway dependent on caspase-9 or by the DR pathways, with the activation of caspase 8. The activation of caspase 3, which represents a key factor for apoptosis execution in both pathways, and the following



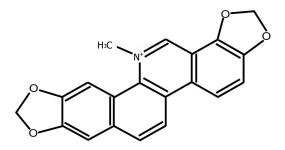


Figure 1 Chemical structure of sanguinarine.

cleavage of PARP together with the down-regulation of Bcl-2 and c-FLIP, may play a very important role in the apoptosis induced by sanguinarine^[26,51,52]. Studies performed in human neuroblastoma cells SH-SY5Y have shown that sanguinarine reduces the expression of anti-apoptotic genes, particularly of NOL3, BCL2, and HRK genes^[45]. A down-regulation of pro-caspase 3, Bcl-2, clAP2, XIAP, and c-FLIPs^[20,52] has been also observed in basal cell-like MDA-MB-231 human breast carcinoma cells treated with sanguinarine. The effect of sanguinarine treatment has been evaluated also on the expression levels of Bax and Bcl-2 proteins in immortalized human keratinocytes (HaCaT)^[24,25], human leukaemia JM1 and K562 cells^[35] and in Hela and SiHa human cervical tumor cells^[53]. These findings indicate that sanguinarine, depending on the dose employed, down-regulates the expression levels of Bcl-2 protein while increasing those of Bax protein, which is a key regulator of mitochondrial damage. Notably, Bax expression has been associated with an increased sensitivity of cancer cells to chemotherapy^[54], whereas an increase of Bcl-2 has been associated with the occurrence of drug-resistance phenomena^[55].

It has been proved that sanguinarine is capable of inducing DNA damage, acting as an intercalating agent^[56,57], and also a very rapid cell apoptosis which does not seem to be mediated by a p53-dependent DNA damage signalling in human colon cancer^[41] and in malignant melanoma cells^[38].

The concentration of sanguinarine plays a key role in the induction of cell death. Consistently, both apoptotic and non-apoptotic cell death pathways have been observed in response to sanguinarine. Thus, a sanguinarine-related and bimodal cell death effect, which consists of two different types of cell death, *i.e.*, by apoptosis (induced by low SA concentration; characterized by caspase 3 and PARP positivity) and oncosis (induced by high SA concentration; characterized by caspase 3 and PARP negativity), has been demonstrated in various cancer cells types^[52].

SANGUINARINE INDUCES ALTERATIONS IN CELL CYCLE

Tumor cells are characterized by deregulated proliferation. Conversely, normal cells proliferation is the results of the action of selected growth signals [cyclins and cyclin-dependent kinases (CDKs)] and anti-growth signals (p21 and p27 proteins). Cyclins and CDKs cooperate in G1 for the initiation of the S phase and in G2 for inducing mitosis, whereas p21 and p27 selectively block the catalytic activity of CDK. Following addition of anti-mitogenic compounds or DNA injury, p21 and p27 bind to cyclin-CDK complex blocking their catalytic activity and consequently the cell cycle progression.

Actually a number of inhibitors and/or regulators of the cell cycle, among which sanguinarine, are suggested as potential antitumor agents. Sanguinarine treatment (0.2-2 mol/L for 24 h) blocks cell cycle by enhancing the expression of CDK inhibitors and by reducing not only cyclin D1, D2 and E, but also CDK2, 4 and 6 in human prostate cancer cells^[27]. This alkaloid also up-regulates p27 and down-regulates cyclin D1, while inhibiting the activation of STAT3, as demonstrated in vitro in basal cell-like MDA-MB-231 human breast cancer cells and in vivo in a murine breast cancer model^[33]. Holy et $al^{(31)}$ studied the effects of sanguinarine (5-10 μ mol/L) on the cell cycle regulatory molecules, by immunecytochemistry, that visualized the cyclin D1 and topoisomerase II in MCF-7 breast cancer cells. They reported that sanguinarine-mediated cellular events induce cell cycle arrest in G0/G1 and inhibit cell proliferation, which is associated with a striking re-localization of cyclin D1 and topoisomerase II from the nucleus to the cytoplasm.

SANGUINARINE-INDUCED APOPTOSIS THROUGH THE GENERATION OF REACTIVE OXYGEN SPECIES

Apoptosis induced by sanguinarine has been associated also with the production of reactive oxygen species (ROS)^[20,36,52,58]. ROS are a group of highly reactive molecules, among which are superoxide anion radical, hydrogen peroxide, singlet oxygen, and hydroxyl radical. ROS are the products of the oxygen metabolism within the cell. ROS are known as key regulators of normal cell proliferation and differentiation, however, high levels of ROS have also been associated with damage of DNA and proteins and thus with the occurrence of apoptosis^[59,60]. Moreover, an overdone oxidative stress has been shown capable of inducing a reduction of the normal mitochondrial membrane potential, which in turn leads to apoptosis^[21,61-63]. It has been shown that ROS generation, is crucial for the apoptosis induced by sanguinarine in human breast cancer^[52], SK-Mel-2 human melanoma^[37], human prostate cancer^[25] and in both HCT-116^[21] and HT-29 human colon cancer cells^[40]. Consistently, pre-treatment of tumor cells with antioxidants such as N-acetylcysteine or glutathione counteracts the apoptosis induced by sanguinarine^[21,32,37,52]. Moreover, the over-expression of cyclooxygenase-2 (COX-2) also rescues prostate cancer cells from sanguinarine-induced apoptosis by



inhibiting the activity of NO synthase, thus suggesting the possibility to use a combination of COX-2 inhibitors and sanguinarine in the treatment of human prostate cancer^[28].

SANGUINARINE-MEDIATED INHIBITION OF NF-KB

The molecular pathways associated with carcinogenesis are linked also with chronic inflammation, which emerges as an important co-factor in tumor development. The NF- κ B controls the inflammatory gene expression and recently it has been suspected to be involved also in the control of tumor development^[64]. Resting NF- κB localizes within the cell cytoplasm in the form of a heterodimer composed by p50, p65, and the inhibitory subunit IkB $\alpha^{[65]}$. Following activation, the IkB α protein is phosphorylated, ubiquitinated and finally degradated. Then, the p50 and p65 reach the nucleus of cell, where they interact with selected DNA sequences localized in the promoter region of various genes, leading to their transcription. Consistently, the NF- κ B signalling pathway has been indicated as a key-target for the development of new chemotherapeutic approaches in cancer.

Sanguinarine has been suggested as a potential actor in the control of NF- κ B-dependent pathological responses by blocking phosphorylation and degradation of IkB α . Studies by Chaturvedi *et al*⁽⁵⁰⁾ showed that in human myeloid ML-1a cells, the treatment with sanguinarine is capable of abrogating, dose- and time-dependently, the activation of NF- κ B induced by tumor necrosis factor.

INHIBITION OF TUMOR ANGIOGENESIS BY SANGUINARINE

Many reports indicate that sanguinarine exerts antitumor activity not only by inhibiting tumor cells migration and/ or invasion, but also by repressing angiogenesis^[22,66]. Since solid tumors require active angiogenesis, the inhibition of endothelial cell proliferation result in the inhibition of tumor growth and progression. The best known angiogenic growth factor is represented by VEGF. Several studies have explored the relationship existing among sanguinarine, angiogenesis and metastatization. In particular, Eun and Koh^[13] showed that sanguinarine inhibits the VEGF-induced endothelial cell migration, sprouting and survival in vitro, and blocks blood vessel formation in vivo in different experimental models. Furthermore, Basini *et al*^[67] showed that sanguinarine is capable of blocking the VEGF-induced blood vessel growth. Depending on the concentration used, sanguinarine also inhibits VEGF secretion in human microvascular endothelial cells HMVEC as well as in A549 lung cancer cells^[68]. This inhibitory effect has been associated with the suppression of the phosphorilation of Akt, p38 and VE-cadherin, which are well known modulators of the VEGF signal transduction pathway^[67,69]. Moreover, sanguinarine enhances apoptosis in human mammary adenocarcinoma MCF-7 through the inhibition of VEGF release, induced by generation of ROS^[32]. Sanguinarine also inhibits angiogenesis in preclinical experimental tumor models, such as mouse melanoma^[48] and rat colorectal cancer, as we reported previously^[49]. In both the experimental studies, the therapeutic efficacy of sanguinarine could not be attributed only to a direct antiproliferative activity but also to the inhibition of tumor angiogenesis induced by this alkaloid.

The rationale of using VEGF-targeted therapies in the treatment of cancer lies in the possibility they offer to counteract the over-expression of VEGF provoked by chemotherapeutic drugs and radiation^[70]. Consistently, dacarbazine, which is used in the therapy of human melanoma, induces increased VEGF-A production^[71], and dacarbazine-resistant melanoma cells show an increased *in vivo* growth together with an increased microvessel density^[72]. These studies suggest the potential application of sanguinarine, alone or in association with other VEGF inhibitors, in the control of both angiogenesis and metastatization of solid tumors.

INHIBITION OF TUMOR CELL INVASION BY SANGUINARINE

In solid tumors, neoplastic cells can penetrate the basement membrane by proteolysis and initiate metastatization, which accounts for the majority of cancer deaths. Metastatization is the result of the cooperation between cancer cells and a sort of "inflammed" microenvironment^[73]. Consistently, inflammatory cells are an important source of proteases capable of causing a degradation of extracellular matrix, which represents a crucial event in the initiation of cancer cell invasion. Matrix metalloproteinases (MMPs) are an example of agents capable to degrade the extracellular matrix^[74,75] and an over-production of these enzymes has been detected in various metastatic cancers^[76-78]. Indeed, there is a strong evidence that increased expression and activation of MMP-2 and MMP-9 is present in tumor tissues but not in normal tissues in patients with breast cancer^[79] and that MMP-2 induces cancer cell migration by means of its interaction with collagen^[80].

Recent findings show that sanguinarine inhibits the tetradecanoylphorbolmyristate acetate (TPA)-induced breast cancer cell migration and invasion while inhibiting the expression of MMP-9, NF- κ B and AP-1 signaling pathways^[81]. Moreover, previous studies by Sun *et al*^[66] have showed that sanguinarine reduces prostate cancer cell growth and invasion by the inhibition of STAT3 activation. STAT3 is constitutively active in human prostate cancer metastases and has a key role in the phenomena of tumor cell migration and invasion in different types of cancer^[82-84]. Since the invasivity and/or metastatic potential of a tumor parallel its maligncy, the above findings indicate that sanguinarine may play a crucial role as a therapeutic agent in anticancer therapy

WJGO | www.wjgnet.com

Gaziano R et al. Antitumor effects of sanguinarine

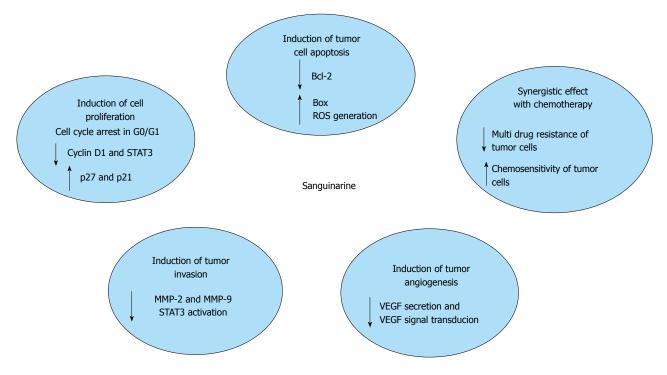


Figure 2 Cellular and molecular mechanisms underlying the antitumor activity of sanguinarine, as assessed by means of *in vitro* and *in vivo* experimental studies. ROS: Reactive oxygen species; Bcl: B cell lymphoma; VEGF: Vascular endothelial growth factor; MMP: Matrix metalloproteinase; STAT: Signal transducer and activator of transcription.

not only for its ability to induce apoptosis but also for its own "anti-invasive" properties.

SYNERGISTIC INTERACTION OF SANGUINARINE WITH CHEMOTHERAPEUTIC AGENTS

Several plant SMs are capable of influencing effectively the multidrug resistance phenomenon in tumor cells and are able also to "chemo-sensitize" them^[85-89]. Some clinical studies have explored the possible advantage of combining natural products with classical chemotherapeutic regimens^[90-92]. Phytotherapy, which employs plants extracts, is still used worldwide for the treatment of various human diseases. However, evidence has been proved that combinations of individual SM in an extract may exert synergistic effects. As an example, a recent study demonstrates that the combined use of non-toxic concentrations of sanguinarine and digitonin with doxorubicin, synergistically sensitizes Caco-2 (human colorectal adenocarcinoma) and CEM/ADR5000 adryamicin-resistant leukemia cells and increases the cytotoxicity of the chemotherapeutic agent doxorubicin^[93]. In this regard, it is worth mentioning that the main advantage of combination therapies is represented by the possibility of reducing the doses and thus the toxicity of chemotherapy, while retaining its own efficacy. Thus, because of its potential synergistic interaction with chemotherapeutic agents, the therapeutic use of sanguinarine as an adjuvant, in association with chemotherapy, might be considered as a theoretical option in cancer therapy.

CONCLUSION

A successful resolution to the design of antitumor drugs relies, at least in part, on the possibility to overcome the intrinsic resistance to undergo apoptosis detected in many transformed cells. Findings from the studies above mentioned show that sanguinarine is capable of inhibiting tumor growth through different molecular pathways (Figure 2). A summary of the results is shown in Table 1. In conclusion, despite sanguinarine has been extensively studied, the precise mechanisms responsible for its antitumor effects still have not been completely elucidated and are strictly dependent on the cell type studied. According to the results obtained so far, it can be said that the anti-tumor action of this alkaloid is the result of a combined effect both on proliferation and invasiveness of tumor cells, that on regulation of the complex phenomena of tumor angiogenesis. In particular, owing to its pro-apoptotic potential, sanquinarine is a good candidate for the development of new anticancer therapies either when used alone or in combination with other chemotherapeutic regimens. More extensive investigation and greater caution are needed, however, to clarify the following important issues. First of all, most of the studies above mentioned have been performed in vitro using cancer cell lines, whereas there are only a few in vivo studies validating the efficacy and safety of sanguinarine administration in animal tumor models. The results of our in vivo studies confirm the effectiveness and safety of using oral sanguinarine administration to control tumor growth in rats^[49]. Similar results had been previously reported in a murine melanoma model^[48]. In that study, and

Table 1 The antitumor activity of sanguinarine

Sanguinarine induces apoptosis in tumor cells through multiple pathways, including the activation of NF-KB, the mitochondrial damage and cell cycle arrest

Sanguinarine-induced apoptosis is associated with the decrease of Bcl-2 and the increase of Bax proteins and the generation of reactive oxygen species Sanguinarine causes cell cycle arrest by increasing the expression of p27 and decreasing cyclin D1, D2 and E, and CDK2, 4 and 6

Sanguinarine inhibits tumor progression associated with chronic inflammation via the inhibition of NF-KB

Sanguinarine inhibits tumor angiogenesis through the inhibition of VEGF secretion and VEGF signal transduction (Akt, p38 and VE-cadherin) Sanguinarine has an inhibitory effect on tumor cell migration by the inhibition of MMP-9 and STAT3 activation

Sanguinarine exerts a synergistic effect with chemotherapeutic agents and enhances the chemosensitivity of Caco 2 and CEM/ADR5000 adryamicinresistant leukemia cells

NF-κB: Nuclear factor-κB; CDK: Cyclin-dependent kinase; Bcl: B cell lymphoma; VEGF: Vascular endothelial growth factor; STAT: Signal transducer and activator of transcription.

in agreement with our findings, the anti-proliferative and anti-angiogenic effects of the oral sanguinarine administration were observed at a dosage, *i.e.*, 5 mg/kg, devoid of apparent toxicity. On the other hand, an increase of serum levels of transaminases and LDH, hepatic vacuolization, lipid accumulation and peroxidation in the liver and a reduction of triglycerides, were observed in mice treated with high-dose sanguinarine (10 mg/kg), suggesting liver injury^[94]. Previous studies showed that sanguinarine can cause physiological dysfunction in skeletal, smooth and cardiac muscles^[95-97]. More recent studies clearly indicate that sanguinarine acts as a pro-apoptotic factor and alters mouse normal embryonic development at a physiological dosage, i.e., 0.5-2 μ mol/L, which are obtained *via* dietary intake^[98]. These experimental results need further confirmation in view of the possible administration of the compound in pregnancy, although at present no teratogenic effects have been reported in humans.

Most of the studies actually known have reported that sanguinarine exerts cytotoxic activity selectively on cancer cells. Consistently, sanguinarine is a negative regulator of human epidermoid carcinoma cells (A431) but not of normal epidermal keratinocytes^[23]. Evidence of this differential activity have been reported recently, showing that mouse lymphocytic leukemic cells are more sensitive to sanguinarine than normal splenocytes^[99].

It is a matter of fact, however, that sanguinarine has been listed as responsible for the toxicity of Argemone mexicana seed oil^[100-102]. Das et al^[103] reported that topical use of argemone oil (0.15-0.3 mL) or sanguinarine (4.5-18 μ mol/L) followed by application of TPA induces tumor development in a murine experimental model. Ansari et al^[104] also reported that intraperitoneal administration of sanguinarine induces DNA damage in Swiss albino mice. Sanguinarine in argemone oil, is suspected to cause glaucoma^[101,102]. Argemone oil increases incidence of bladder cancer in animal models^[103] and of gall bladder cancer in humans^[104]. Furthermore, sanguinarine extract from bloodroot (Sanguinaria canadensis), previously used in oral hygiene products, was discontinued until a link between product administration and occurrence of leukoplakia was established^[105,106]. Hepatic microsomes transform sanguinarine in a mutagenic epoxide and the same sanguinarine is capable of activating polycyclic aromatic hydrocarbon signaling^[107]. However, related to this topic, the results available in literature are not univocal^[3]. So that is still not clear if sanguinarine may act as a carcinogenic without the cooperation of other risk factors or it is capable of acting in concert with various cocarcinogens. In light of the above facts, the possibility of obtaining beneficial effects in humans by using sanguinarine remains largely unpredictable.

Finally, since at present there is increasing interest in nanotechnology application in cancer therapy and in order to prevent the potential toxic and/or side effects induced by sanguinarine administration, in vivo studies might be performed in experimental tumor models by encapsulating the alkaloid in tumor-targeted nanoparticles^[108], which accumulate preferentially in tumors recognizing single cancer cells for diagnosis and treatment. Actually, the administration of sanguinarine (10 mg/kg) per os and encapsulated by lipid nanoparticles (SG-SLNs), has been shown to induce an antiinflammatory effect in an LPS-induced endotoxin shock murine model, and the pharmacokinetic studies have proved that the AUC0-24 and Cmax of SG-SLNs were significantly increased when compared to those of sanguinarine alone^[109].

In conclusion, several studies indicate the potential application of sanguinarine as an adjuvant in the therapy of cancer, but further detailed pharmacokinetic and toxicology studies, which have to be conducted in appropriate experimental tumor models, are absolutely required to assess the efficacy and safety of this compound before such an antitumor strategy can be translated in clinical trials.

REFERENCES

- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; 100: 57-70 [PMID: 10647931 DOI: 10.1016/S0092-8674(00)8168 3-9]
- 2 Reddy L, Odhav B, Bhoola KD. Natural products for cancer prevention: a global perspective. *Pharmacol Ther* 2003; 99: 1-13 [PMID: 12804695 DOI: 10.1016/S0163-7258(03)00042-1]
- 3 Dvorak Z, Kuban V, Kledjus B, Hlavac J, Vicar J, Ulrichova J, Simanek V. Quaternary benzo[c]phenanthridines sanguinarine and chelerythrine: a review of investigations from chemical and biological studies. *Heterocycles* 2006; 68: 2403-2422 [DOI: 10.1002/chin.200709233]
- 4 Dostàl J, Slavik J. Some aspects of the chemistry of quaternary benzo[c]phenanthridines alkaloids. *Stud Nat Prod Chem* 2002; 27:



155-184 [DOI: 10.1016/S1572-5995(02)80036-9]

- 5 Psotova J, Vecera R, Zdarilova A, Anzebacherova E, Kosina P, Svobodova A, Hrbac J, Jirovsky D, Stiborova M, Lichnovsky V. Safety assessment of sanguitrin, alkaloid fraction of Macleaya cordata, in rats. *Vet Med* 2006; **51**: 145-155
- 6 Kosina P, Walterová D, Ulrichová J, Lichnovský V, Stiborová M, Rýdlová H, Vicar J, Krecman V, Brabec MJ, Simánek V. Sanguinarine and chelerythrine: assessment of safety on pigs in ninety days feeding experiment. *Food Chem Toxicol* 2004; **42**: 85-91 [PMID: 14630132 DOI: 10.1007/s11101-013-9290-8]
- 7 Slaninova I, Pencikova K, Urbanova J, Slanina J, Taborska E. Antitumor activities of sanguinarine and related alkaloids. *Phytochem Rev* 2014; 13: 51-68 [DOI: 10.1007/s1101-013-9290-8]
- 8 Miao F, Yang XJ, Zhou L, Hu HJ, Zheng F, Ding XD, Sun DM, Zhou CD, Sun W. Structural modification of sanguinarine and chelerythrine and their antibacterial activity. *Nat Prod Res* 2011; 25: 863-875 [PMID: 21491327 DOI: 10.1080/14786419.2010.482055]
- 9 Yang XJ, Miao F, Yao Y, Cao FJ, Yang R, Ma YN, Qin BF, Zhou L. In vitro antifungal activity of sanguinarine and chelerythrine derivatives against phytopathogenic fungi. *Molecules* 2012; 17: 13026-13035 [PMID: 23124471 DOI: 10.3390/molecules1711130 26]
- 10 Lenfeld J, Kroutil M, Marsálek E, Slavík J, Preininger V, Simánek V. Antiinflammatory activity of quaternary benzophenanthridine alkaloids from Chelidonium majus. *Planta Med* 1981; 43: 161-165 [PMID: 7312984]
- 11 Tan GT, Pezzuto JM, Kinghorn AD, Hughes SH. Evaluation of natural products as inhibitors of human immunodeficiency virus type 1 (HIV-1) reverse transcriptase. *J Nat Prod* 1991; 54: 143-154 [PMID: 1710653]
- 12 Tsai IL, Wun MF, Teng CM, Ishikawa T, Chen IS. Anti-platelet aggregation constituents from Formosan Toddalia asiatica. *Phytochemistry* 1998; 48: 1377-1382 [PMID: 9720317 DOI: 10.1016/S0031-9422(97)00678-X]
- 13 Eun JP, Koh GY. Suppression of angiogenesis by the plant alkaloid, sanguinarine. *Biochem Biophys Res Commun* 2004; 317: 618-624 [PMID: 15063803 DOI: 10.1016/j.bbrc.2004.03.077]
- 14 Miao F, Yang XJ, Ma YN, Zheng F, Song XP, Zhou L. Structural modification of sanguinarine and chelerythrine and their in vitro acaricidal activity against Psoroptes cuniculi. *Chem Pharm Bull* (Tokyo) 2012; 60: 1508-1513 [PMID: 23000953 DOI: 10.1248/cpb. c12-00618]
- 15 Yao JY, Li XL, Shen JY, Pan XY, Hao GJ, Xu Y, Ying WL, Ru HS, Liu XL. Isolation of bioactive components from Chelidonium majus L. with activity against Trichodina sp. *Aquaculture* 2011; 318: 235-238 [DOI: 10.1016/j.aquaculture.2011.04.035]
- 16 Wang GX, Zhou Z, Jiang DX, Han J, Wang JF, Zhao LW, Li J. In vivo anthelmintic activity of five alkaloids from Macleaya microcarpa (Maxim) Fedde against Dactylogyrus intermedius in Carassius auratus. *Vet Parasitol* 2010; **171**: 305-313 [PMID: 20413222 DOI: 10.1016/j.vetpar.2010.03.032]
- Steller H. Mechanisms and genes of cellular suicide. *Science* 1995;
 267: 1445-1449 [PMID: 7878463 DOI: 10.1126/science.7878463]
- Meier P, Finch A, Evan G. Apoptosis in development. *Nature* 2000;
 407: 796-801 [PMID: 11048731 DOI: 10.1038/35037734]
- 19 Ahsan H, Reagan-Shaw S, Breur J, Ahmad N. Sanguinarine induces apoptosis of human pancreatic carcinoma AsPC-1 and BxPC-3 cells via modulations in Bcl-2 family proteins. *Cancer Lett* 2007; 249: 198-208 [PMID: 17005319 DOI: 10.1016/j.canlet.2006.08.018]
- 20 Choi WY, Kim GY, Lee WH, Choi YH. Sanguinarine, a benzophenanthridine alkaloid, induces apoptosis in MDA-MB-231 human breast carcinoma cells through a reactive oxygen speciesmediated mitochondrial pathway. *Chemotherapy* 2008; 54: 279-287 [PMID: 18667818 DOI: 10.1159/000149719]
- 21 Han MH, Kim GY, Yoo YH, Choi YH. Sanguinarine induces apoptosis in human colorectal cancer HCT-116 cells through ROS-mediated Egr-1 activation and mitochondrial dysfunction. *Toxicol Lett* 2013; 220: 157-166 [PMID: 23660334 DOI: 10.1016/ j.toxlet.2013.04.020]
- 22 Jang BC, Park JG, Song DK, Baek WK, Yoo SK, Jung KH, Park

GY, Lee TY, Suh SI. Sanguinarine induces apoptosis in A549 human lung cancer cells primarily via cellular glutathione depletion. *Toxicol In Vitro* 2009; **23**: 281-287 [PMID: 19135517 DOI: 10.1016/j.tiv.2008.12.013]

- 23 Ahmad N, Gupta S, Husain MM, Heiskanen KM, Mukhtar H. Differential antiproliferative and apoptotic response of sanguinarine for cancer cells versus normal cells. *Clin Cancer Res* 2000; 6: 1524-1528 [PMID: 10778985]
- 24 Adhami VM, Aziz MH, Mukhtar H, Ahmad N. Activation of prodeath Bcl-2 family proteins and mitochondrial apoptosis pathway by sanguinarine in immortalized human HaCaT keratinocytes. *Clin Cancer Res* 2003; 9: 3176-3182 [PMID: 12912970]
- 25 Reagan-Shaw S, Breur J, Ahmad N. Enhancement of UVB radiation-mediated apoptosis by sanguinarine in HaCaT human immortalized keratinocytes. *Mol Cancer Ther* 2006; 5: 418-429 [PMID: 16505117 DOI: 10.1158/1535-7163.MCT-05-0250]
- 26 Malíková J, Zdarilová A, Hlobilková A, Ulrichová J. The effect of chelerythrine on cell growth, apoptosis, and cell cycle in human normal and cancer cells in comparison with sanguinarine. *Cell Biol Toxicol* 2006; 22: 439-453 [PMID: 16964588 DOI: 10.1007/ s10565-006-0109-x]
- 27 Adhami VM, Aziz MH, Reagan-Shaw SR, Nihal M, Mukhtar H, Ahmad N. Sanguinarine causes cell cycle blockade and apoptosis of human prostate carcinoma cells via modulation of cyclin kinase inhibitor-cyclin-cyclin-dependent kinase machinery. *Mol Cancer Ther* 2004; **3**: 933-940 [PMID: 15299076]
- 28 Huh J, Liepins A, Zielonka J, Andrekopoulos C, Kalyanaraman B, Sorokin A. Cyclooxygenase 2 rescues LNCaP prostate cancer cells from sanguinarine-induced apoptosis by a mechanism involving inhibition of nitric oxide synthase activity. *Cancer Res* 2006; 66: 3726-3736 [PMID: 16585199 DOI: 10.1158/0008-5472. CAN-05-4033]
- 29 Ding Z, Tang SC, Weerasinghe P, Yang X, Pater A, Liepins A. The alkaloid sanguinarine is effective against multidrug resistance in human cervical cells via bimodal cell death. *Biochem Pharmacol* 2002; 63: 1415-1421 [PMID: 11996882 DOI: 10.1016/S0006-2952 (02)00902-4]
- 30 Debiton E, Madelmont JC, Legault J, Barthomeuf C. Sanguinarineinduced apoptosis is associated with an early and severe cellular glutathione depletion. *Cancer Chemother Pharmacol* 2003; 51: 474-482 [PMID: 12700925]
- 31 Holy J, Lamont G, Perkins E. Disruption of nucleocytoplasmic trafficking of cyclin D1 and topoisomerase II by sanguinarine. *BMC Cell Biol* 2006; 7: 13 [PMID: 16512916 DOI: 10.1186/147-2121-7-13]
- 32 Dong XZ, Zhang M, Wang K, Liu P, Guo DH, Zheng XL, Ge XY. Sanguinarine inhibits vascular endothelial growth factor release by generation of reactive oxygen species in MCF-7 human mammary adenocarcinoma cells. *Biomed Res Int* 2013; 2013: 517698 [PMID: 23762849 DOI: 10.1155/2013/517698]
- 33 Kalogris C, Garulli C, Pietrella L, Gambini V, Pucciarelli S, Lucci C, Tilio M, Zabaleta ME, Bartolacci C, Andreani C, Giangrossi M, Iezzi M, Belletti B, Marchini C, Amici A. Sanguinarine suppresses basal-like breast cancer growth through dihydrofolate reductase inhibition. *Biochem Pharmacol* 2014; **90**: 226-234 [PMID: 24875448 DOI: 10.1016/j.bcp.2014.05.014]
- 34 Han MH, Yoo YH, Choi YH. Sanguinarine-induced apoptosis in human leukemia U937 cells via Bcl-2 downregulation and caspase-3 activation. *Chemotherapy* 2008; 54: 157-165 [PMID: 18560221 DOI: 10.1159/000140359]
- 35 Weerasinghe P, Hallock S, Tang SC, Liepins A. Role of Bcl-2 family proteins and caspase-3 in sanguinarine-induced bimodal cell death. *Cell Biol Toxicol* 2001; 17: 371-381 [PMID: 11787859]
- 36 Hussain AR, Al-Jomah NA, Siraj AK, Manogaran P, Al-Hussein K, Abubaker J, Platanias LC, Al-Kuraya KS, Uddin S. Sanguinarinedependent induction of apoptosis in primary effusion lymphoma cells. *Cancer Res* 2007; 67: 3888-3897 [PMID: 17440103 DOI: 10.1158/0008-5472.CAN-06-3764]
- 37 **Burgeiro A**, Bento AC, Gajate C, Oliveira PJ, Mollinedo F. Rapid human melanoma cell death induced by sanguinarine through



oxidative stress. *Eur J Pharmacol* 2013; **705**: 109-118 [PMID: 23499690 DOI: 10.1016/j.ejphar.2013.02.035]

- 38 Hammerová J, Uldrijan S, Táborská E, Slaninová I. Benzo[c] phenanthridine alkaloids exhibit strong anti-proliferative activity in malignant melanoma cells regardless of their p53 status. J Dermatol Sci 2011; 62: 22-35 [PMID: 21324654 DOI: 10.1016/j. jdermsci.2011.01.006]
- 39 Serafim TL, Matos JA, Sardão VA, Pereira GC, Branco AF, Pereira SL, Parke D, Perkins EL, Moreno AJ, Holy J, Oliveira PJ. Sanguinarine cytotoxicity on mouse melanoma K1735-M2 cellsnuclear vs. mitochondrial effects. *Biochem Pharmacol* 2008; **76**: 1459-1475 [PMID: 18692024 DOI: 10.1016/j.bcp.2008.07.013]
- 40 Lee JS, Jung WK, Jeong MH, Yoon TR, Kim HK. Sanguinarine induces apoptosis of HT-29 human colon cancer cells via the regulation of Bax/Bcl-2 ratio and caspase-9-dependent pathway. *Int J Toxicol* 2012; **31**: 70-77 [PMID: 22215411 DOI: 10.1177/109158 1811423845]
- 41 Matkar SS, Wrischnik LA, Hellmann-Blumberg U. Sanguinarine causes DNA damage and p53-independent cell death in human colon cancer cell lines. *Chem Biol Interact* 2008; 172: 63-71 [PMID: 18243168 DOI: 10.1016/j.cbi.2007.12.006]
- 42 Choi WY, Jin CY, Han MH, Kim GY, Kim ND, Lee WH, Kim SK, Choi YH. Sanguinarine sensitizes human gastric adenocarcinoma AGS cells to TRAIL-mediated apoptosis via down-regulation of AKT and activation of caspase-3. *Anticancer Res* 2009; 29: 4457-4465 [PMID: 20032392]
- 43 Larsson DE, Wickström M, Hassan S, Oberg K, Granberg D. The cytotoxic agents NSC-95397, brefeldin A, bortezomib and sanguinarine induce apoptosis in neuroendocrine tumors in vitro. *Anticancer Res* 2010; **30**: 149-156 [PMID: 20150630]
- 44 Park H, Bergeron E, Senta H, Guillemette K, Beauvais S, Blouin R, Sirois J, Faucheux N. Sanguinarine induces apoptosis of human osteosarcoma cells through the extrinsic and intrinsic pathways. *Biochem Biophys Res Commun* 2010; **399**: 446-451 [PMID: 20678472 DOI: 10.1016/j.bbrc.2010.07.114]
- 45 Cecen E, Altun Z, Ercetin P, Aktas S, Olgun N. Promoting effects of sanguinarine on apoptotic gene expression in human neuroblastoma cells. *Asian Pac J Cancer Prev* 2014; 15: 9445-9451 [PMID: 25422239 DOI: 10.7314/APJCP.2014.15.21.9445]
- 46 Deroussent A, Ré M, Hoellinger H, Cresteil T. Metabolism of sanguinarine in human and in rat: characterization of oxidative metabolites produced by human CYP1A1 and CYP1A2 and rat liver microsomes using liquid chromatography-tandem mass spectrometry. *J Pharm Biomed Anal* 2010; **52**: 391-397 [PMID: 19804952 DOI: 10.1016/j.jpba.2009.09.014]
- 47 Lopez Lozano MJ, Rios Santos V, Bullon Fernandez P. [Effectiveness of chemical products as antiplaque agents]. *Rev Eur Odontoestomatol* 1991; 3: 115-122 [PMID: 1867730]
- 48 De Stefano I, Raspaglio G, Zannoni GF, Travaglia D, Prisco MG, Mosca M, Ferlini C, Scambia G, Gallo D. Antiproliferative and antiangiogenic effects of the benzophenanthridine alkaloid sanguinarine in melanoma. *Biochem Pharmacol* 2009; 78: 1374-1381 [PMID: 19643088 DOI: 10.1016/j.bcp.2009.07.011]
- 49 Pica F, Balestrieri E, Serafino A, Sorrentino R, Gaziano R, Moroni G, Moroni N, Palmieri G, Mattei M, Garaci E, Sinibaldi-Vallebona P. Antitumor effects of the benzophenanthridine alkaloid sanguinarine in a rat syngeneic model of colorectal cancer. *Anticancer Drugs* 2012; 23: 32-42 [PMID: 21849887 DOI: 10.1097/ CAD.0b013e32834a0c8e]
- 50 Chaturvedi MM, Kumar A, Darnay BG, Chainy GB, Agarwal S, Aggarwal BB. Sanguinarine (pseudochelerythrine) is a potent inhibitor of NF-kappaB activation, IkappaBalpha phosphorylation, and degradation. *J Biol Chem* 1997; 272: 30129-30134 [PMID: 9374492 DOI: 10.1074/jbc.272.48.30129]
- 51 Malikova J, Zdarilova A, Hlobilkova A. Effects of sanguinarine and chelerythrine on the cell cycle and apoptosis. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 2006; 150: 5-12 [PMID: 16936897]
- 52 **Kim S**, Lee TJ, Leem J, Choi KS, Park JW, Kwon TK. Sanguinarine-induced apoptosis: generation of ROS, down-regulation

of Bcl-2, c-FLIP, and synergy with TRAIL. *J Cell Biochem* 2008; **104**: 895-907 [PMID: 18189268 DOI: 10.1002/jcb.21672]

- 53 Xu JY, Meng QH, Chong Y, Jiao Y, Zhao L, Rosen EM, Fan S. Sanguinarine inhibits growth of human cervical cancer cells through the induction of apoptosis. *Oncol Rep* 2012; 28: 2264-2270 [PMID: 22965493 DOI: 10.3892/or.2012.2024]
- 54 Xu ZW, Friess H, Büchler MW, Solioz M. Overexpression of Bax sensitizes human pancreatic cancer cells to apoptosis induced by chemotherapeutic agents. *Cancer Chemother Pharmacol* 2002; 49: 504-510 [PMID: 12107556 DOI: 10.1007/s00280-002-0435-5]
- 55 Reed JC. Bcl-2: prevention of apoptosis as a mechanism of drug resistance. *Hematol Oncol Clin North Am* 1995; 9: 451-473 [PMID: 7642473]
- 56 Slaninová I, Slanina J, Táborská E. Quaternary benzo[c]phenanthridine alkaloids--novel cell permeant and red fluorescing DNA probes. *Cytometry A* 2007; 71: 700-708 [PMID: 17549765 DOI: 10.1002/cyto.a.20423]
- 57 Saran A, Srivastava S, Coutinho E, Maiti M. 1H NMR investigation of the interaction of berberine and sanguinarine with DNA. *Indian J Biochem Biophys* 1995; 32: 74-77 [PMID: 7642203]
- 58 Chang MC, Chan CP, Wang YJ, Lee PH, Chen LI, Tsai YL, Lin BR, Wang YL, Jeng JH. Induction of necrosis and apoptosis to KB cancer cells by sanguinarine is associated with reactive oxygen species production and mitochondrial membrane depolarization. *Toxicol Appl Pharmacol* 2007; **218**: 143-151 [PMID: 17196629 DOI: 10.1016/j.taap.2006.10.025]
- 59 Carmody RJ, Cotter TG. Signalling apoptosis: a radical approach. *Redox Rep* 2001; 6: 77-90 [PMID: 11450987 DOI: 10.1179/135100 001101536085]
- 60 Pelicano H, Carney D, Huang P. ROS stress in cancer cells and therapeutic implications. *Drug Resist Updat* 2004; 7: 97-110 [PMID: 15158766 DOI: 10.1016/j.drup.2004.01.004]
- 61 Simon L, Szilágyi G, Bori Z, Telek G, Magyar K, Nagy Z. Low dose (-)deprenyl is cytoprotective: it maintains mitochondrial membrane potential and eliminates oxygen radicals. *Life Sci* 2005; 78: 225-231 [PMID: 16242156 DOI: 10.1016/j.lfs.2005.04.078]
- 62 Skulachev VP. Bioenergetic aspects of apoptosis, necrosis and mitoptosis. *Apoptosis* 2006; 11: 473-485 [PMID: 16532373 DOI: 10.1007/s10495-006-5881-9]
- 63 Kim IH, Kim SW, Kim SH, Lee SO, Lee ST, Kim DG, Lee MJ, Park WH. Parthenolide-induced apoptosis of hepatic stellate cells and anti-fibrotic effects in an in vivo rat model. *Exp Mol Med* 2012; 44: 448-456 [PMID: 22581380 DOI: 10.3858/emm.2012.44.7.051]
- 64 Karin M. NF-kappaB as a critical link between inflammation and cancer. *Cold Spring Harb Perspect Biol* 2009; 1: a000141 [PMID: 20066113 DOI: 10.1101/cshperspect.a000141]
- 65 Ghosh S, Karin M. Missing pieces in the NF-kappaB puzzle. *Cell* 2002; 109 Suppl: S81-S96 [PMID: 11983155 DOI: 10.1016/ S0092-8674(02)00703-1]
- 66 Sun M, Liu C, Nadiminty N, Lou W, Zhu Y, Yang J, Evans CP, Zhou Q, Gao AC. Inhibition of Stat3 activation by sanguinarine suppresses prostate cancer cell growth and invasion. *Prostate* 2012; 72: 82-89 [PMID: 21538419 DOI: 10.1002/pros.21409]
- Basini G, Bussolati S, Santini SE, Grasselli F. Sanguinarine inhibits VEGF-induced angiogenesis in a fibrin gel matrix. *Biofactors* 2007; 29: 11-18 [PMID: 17611290]
- 68 Xu JY, Meng QH, Chong Y, Jiao Y, Zhao L, Rosen EM, Fan S. Sanguinarine is a novel VEGF inhibitor involved in the suppression of angiogenesis and cell migration. *Mol Clin Oncol* 2013; 1: 331-336 [PMID: 24649171 DOI: 10.3892/mco.2012.41]
- 69 Basini G, Santini SE, Bussolati S, Grasselli F. The plant alkaloid sanguinarine is a potential inhibitor of follicular angiogenesis. J Reprod Dev 2007; 53: 573-579 [PMID: 17310078]
- 70 Ellis LM, Hicklin DJ. VEGF-targeted therapy: mechanisms of anti-tumour activity. *Nat Rev Cancer* 2008; 8: 579-591 [PMID: 18596824 DOI: 10.1038/nrc2403]
- 71 Lev DC, Ruiz M, Mills L, McGary EC, Price JE, Bar-Eli M. Dacarbazine causes transcriptional up-regulation of interleukin 8 and vascular endothelial growth factor in melanoma cells: a possible escape mechanism from chemotherapy. *Mol Cancer Ther* 2003; 2:

753-763 [PMID: 12939465]

- 72 Lev DC, Onn A, Melinkova VO, Miller C, Stone V, Ruiz M, McGary EC, Ananthaswamy HN, Price JE, Bar-Eli M. Exposure of melanoma cells to dacarbazine results in enhanced tumor growth and metastasis in vivo. *J Clin Oncol* 2004; 22: 2092-2100 [PMID: 15123733 DOI: 10.1200/JCO.2004.11.070]
- 73 Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010; 140: 883-899 [PMID: 20303878 DOI: 10.1016/ j.cell.2010.01.025]
- 74 Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. *Annu Rev Cell Dev Biol* 2001; 17: 463-516 [PMID: 11687497 DOI: 10.1146/annurev.cellbio.17.1.463]
- 75 Jiang MC, Liao CF, Lee PH. Aspirin inhibits matrix metalloproteinase-2 activity, increases E-cadherin production, and inhibits in vitro invasion of tumor cells. *Biochem Biophys Res Commun* 2001; 282: 671-677 [PMID: 11401513 DOI: 10.1006/bbrc.2001.4637]
- 76 Cox TR, Erler JT. Remodeling and homeostasis of the extracellular matrix: implications for fibrotic diseases and cancer. *Dis Model Mech* 2011; 4: 165-178 [PMID: 21324931 DOI: 10.1242/dmm.004 077]
- 77 Deryugina EI, Quigley JP. Matrix metalloproteinases and tumor metastasis. *Cancer Metastasis Rev* 2006; 25: 9-34 [PMID: 16680569 DOI: 10.1007/s10555-006-7886-9]
- 78 Jinga DC, Blidaru A, Condrea I, Ardeleanu C, Dragomir C, Szegli G, Stefanescu M, Matache C. MMP-9 and MMP-2 gelatinases and TIMP-1 and TIMP-2 inhibitors in breast cancer: correlations with prognostic factors. *J Cell Mol Med* 2006; 10: 499-510 [PMID: 16796815 DOI: 10.1111/j.1582-4934.2006.tb00415.x]
- 79 Scorilas A, Karameris A, Arnogiannaki N, Ardavanis A, Bassilopoulos P, Trangas T, Talieri M. Overexpression of matrixmetalloproteinase-9 in human breast cancer: a potential favourable indicator in node-negative patients. *Br J Cancer* 2001; 84: 1488-1496 [PMID: 11384099 DOI: 10.1054/bjoc.2001.1810]
- 80 Xu X, Wang Y, Chen Z, Sternlicht MD, Hidalgo M, Steffensen B. Matrix metalloproteinase-2 contributes to cancer cell migration on collagen. *Cancer Res* 2005; 65: 130-136 [PMID: 15665288]
- 81 Park SY, Jin ML, Kim YH, Lee SJ, Park G. Sanguinarine inhibits invasiveness and the MMP-9 and COX-2 expression in TPA-induced breast cancer cells by inducing HO-1 expression. *Oncol Rep* 2014; 31: 497-504 [PMID: 24220687 DOI: 10.3892/ or.2013.2843]
- 82 Abdulghani J, Gu L, Dagvadorj A, Lutz J, Leiby B, Bonuccelli G, Lisanti MP, Zellweger T, Alanen K, Mirtti T, Visakorpi T, Bubendorf L, Nevalainen MT. Stat3 promotes metastatic progression of prostate cancer. *Am J Pathol* 2008; **172**: 1717-1728 [PMID: 18483213 DOI: 10.2353/ajpath.2008.071054]
- 83 Itoh M, Murata T, Suzuki T, Shindoh M, Nakajima K, Imai K, Yoshida K. Requirement of STAT3 activation for maximal collagenase-1 (MMP-1) induction by epidermal growth factor and malignant characteristics in T24 bladder cancer cells. *Oncogene* 2006; 25: 1195-1204 [PMID: 16205632 DOI: 10.1038/ sj.onc.1209149]
- 84 Silver DL, Naora H, Liu J, Cheng W, Montell DJ. Activated signal transducer and activator of transcription (STAT) 3: localization in focal adhesions and function in ovarian cancer cell motility. *Cancer Res* 2004; 64: 3550-3558 [PMID: 15150111 DOI: 10.1158/0008-54 72.CAN-03-3959]
- 85 Möller M, Weiss J, Wink M. Reduction of cytotoxicity of the alkaloid emetine through P-glycoprotein (MDR1/ABCB1) in human Caco-2 cells and leukemia cell lines. *Planta Med* 2006; **72**: 1121-1126 [PMID: 16783693 DOI: 10.1055/s-2006-941546]
- 86 Ma Y, Wink M. Lobeline, a piperidine alkaloid from Lobelia can reverse P-gp dependent multidrug resistance in tumor cells. *Phytomedicine* 2008; 15: 754-758 [PMID: 18222670 DOI: 10.1016/ j.phymed.2007.11.028]
- 87 El-Readi MZ, Hamdan D, Farrag N, El-Shazly A, Wink M. Inhibition of P-glycoprotein activity by limonin and other secondary metabolites from Citrus species in human colon and leukaemia cell lines. *Eur J Pharmacol* 2010; 626: 139-145 [PMID: 19782062 DOI: 10.1016/j.ejphar.2009.09.040]

- 88 Li S, Lei Y, Jia Y, Li N, Wink M, Ma Y. Piperine, a piperidine alkaloid from Piper nigrum re-sensitizes P-gp, MRP1 and BCRP dependent multidrug resistant cancer cells. *Phytomedicine* 2011; 19: 83-87 [PMID: 21802927 DOI: 10.1016/j.phymed.2011.06.031]
- 89 Wink M, Ashour ML, El-Readi MZ. Secondary Metabolites from Plants Inhibiting ABC Transporters and Reversing Resistance of Cancer Cells and Microbes to Cytotoxic and Antimicrobial Agents. *Front Microbiol* 2012; **3**: 130 [PMID: 22536197 DOI: 10.3389/ fmicb.2012.00130]
- 90 Li JH. [A study on treatment of lung cancer by combined therapy of traditional Chinese medicine and chemotherapy]. *Zhongguo Zhong Xi Yi Jie He Za Zhi* 1996; **16**: 136-138 [PMID: 9208532]
- 91 Cai HB, Dai FG, Min QF, Shi M, Miao JX, Luo RC. [Clinical study of the effects of radiotherapy in combination with traditional Chinese medicine on non-small cell lung cancer]. Di Yi Jun Yi Da Xue Xue Bao 2002; 22: 1112-1113 [PMID: 12480588]
- 92 Efferth T, Davey M, Olbrich A, Rücker G, Gebhart E, Davey R. Activity of drugs from traditional Chinese medicine toward sensitive and MDR1- or MRP1-overexpressing multidrug-resistant human CCRF-CEM leukemia cells. *Blood Cells Mol Dis* 2002; 28: 160-168 [PMID: 12064912 DOI: 10.1006/bcmd.2002.0492]
- 93 Eid SY, El-Readi MZ, Wink M. Synergism of three-drug combinations of sanguinarine and other plant secondary metabolites with digitonin and doxorubicin in multi-drug resistant cancer cells. *Phytomedicine* 2012; 19: 1288-1297 [PMID: 23146422 DOI: 10.1016/j.phymed.2012.08.010]
- 94 Choy CS, Cheah KP, Chiou HY, Li JS, Liu YH, Yong SF, Chiu WT, Liao JW, Hu CM. Induction of hepatotoxicity by sanguinarine is associated with oxidation of protein thiols and disturbance of mitochondrial respiration. *J Appl Toxicol* 2008; 28: 945-956 [PMID: 18548746 DOI: 10.1002/jat.1360]
- 95 Hu CM, Cheng HW, Cheng YW, Kang JJ. Induction of skeletal muscle contracture and calcium release from isolated sarcoplasmic reticulum vesicles by sanguinarine. *Br J Pharmacol* 2000; 130: 299-306 [PMID: 10807666 DOI: 10.1038/sj.bjp.0703279]
- 96 Hu CM, Cheng HW, Cheng YW, Kan JJ. Mechanisms underlying the induction of vasorelaxation in rat thoracic aorta by sanguinarine. *Jpn J Pharmacol* 2001; 85: 47-53 [PMID: 11243574]
- 97 Hu CM, Cheng YW, Liao JW, Cheng HW, Kang JJ. Induction of contracture and extracellular Ca2+ influx in cardiac muscle by sanguinarine: a study on cardiotoxicity of sanguinarine. *J Biomed Sci* 2005; 12: 399-407 [PMID: 15920678 DOI: 10.1007/s11373-005-3007-y]
- 98 Chan WH. Embryonic toxicity of sanguinarine through apoptotic processes in mouse blastocysts. *Toxicol Lett* 2011; 205: 285-292 [PMID: 21722720 DOI: 10.1016/j.toxlet.2011.06.018]
- 99 Kaminskyy V, Lin KW, Filyak Y, Stoika R. Differential effect of sanguinarine, chelerythrine and chelidonine on DNA damage and cell viability in primary mouse spleen cells and mouse leukemic cells. *Cell Biol Int* 2008; **32**: 271-277 [PMID: 18029203 DOI: 10.1016/j.cellbi.2007.09.004]
- Das M, Khanna SK. Clinicoepidemiological, toxicological, and safety evaluation studies on argemone oil. *Crit Rev Toxicol* 1997; 27: 273-297 [PMID: 9189656 DOI: 10.3109/10408449709089896]
- 101 Hakim SA. Argemone oil, sanguinarine, and epidemic-dropsy glaucoma. Br J Ophthalmol 1954; 38: 193-216 [PMID: 13149763 DOI: 10.1136/bjo.38.4.193]
- 102 Hakim SA. Argemone oil, sanguinarine, dropsy, glaucoma and cancer. *Indian Pract* 1967; 20: 129-141 [PMID: 6037511]
- 103 Das M, Ansari KM, Dhawan A, Shukla Y, Khanna SK. Correlation of DNA damage in epidemic dropsy patients to carcinogenic potential of argemone oil and isolated sanguinarine alkaloid in mice. *Int J Cancer* 2005; 117: 709-717 [PMID: 15981203 DOI: 10.1002/ijc.21234]
- 104 Ansari KM, Dhawan A, Khanna SK, Das M. In vivo DNA damaging potential of sanguinarine alkaloid, isolated from argemone oil, using alkaline Comet assay in mice. *Food Chem Toxicol* 2005; 43: 147-153 [PMID: 15582207 DOI: 10.1016/j.fct.2004.09.005]
- 105 Anderson KM, Stoner GD, Fields HW, Chacon GE, Dohar AL, Gregg BR, Mallery SR. Immunohistochemical assessment of

Gaziano R et al. Antitumor effects of sanguinarine

Viadent-associated leukoplakia. *Oral Oncol* 2005; **41**: 200-207 [PMID: 15695122 DOI: 10.1016/j.oraloncology.2004.08.008]

- 106 Damm DD, Curran A, White DK, Drummond JF. Leukoplakia of the maxillary vestibule--an association with Viadent? Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1999; 87: 61-66 [PMID: 9927082 DOI: 10.1016/S1079-2104(99)70296-9]
- 107 Karp JM, Rodrigo KA, Pei P, Pavlick MD, Andersen JD, McTigue DJ, Fields HW, Mallery SR. Sanguinarine activates polycyclic aromatic hydrocarbon associated metabolic pathways in human oral

keratinocytes and tissues. *Toxicol Lett* 2005; **158**: 50-60 [PMID: 15993743 DOI: 10.1016/j.toxlet.2005.02.007]

- 108 Wang M, Thanou M. Targeting nanoparticles to cancer. *Pharmacol Res* 2010; 62: 90-99 [PMID: 20380880 DOI: 10.1016/j.phrs.2010.0 3.005]
- 109 Li W, Li H, Yao H, Mu Q, Zhao G, Li Y, Hu H, Niu X. Pharmacokinetic and anti-inflammatory effects of sanguinarine solid lipid nanoparticles. *Inflammation* 2014; **37**: 632-638 [PMID: 24272172 DOI: 10.1007/s10753-013-9779-8]

P- Reviewer: Batistoni R, Mohammad RM S- Editor: Ji FF L- Editor: A E- Editor: Jiao XK







Published by Baishideng Publishing Group Inc

8226 Regency Drive, Pleasanton, CA 94588, USA Telephone: +1-925-223-8242 Fax: +1-925-223-8243 E-mail: bpgoffice@wjgnet.com Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com

