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Inside the different types of carbon black as nanomodifiers for screen-printed electrodes



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ABSTRACT

The need to deliver high performant and miniaturised electrochemical sensors has boosted the use of carbon nanomaterials as smart modifiers of working electrode surface. Among the carbon nanomaterials. the common and cost-effective carbon black has recently attracted the attention from the scientific community for its outstanding features as electrode nanomodifier for analyte detection. Herein, we report the structural and morphological characterisation of several types of carbon blacks, namely HP 160, HS20, MTN 990, N115, N220, N375, N660, PL6, Super P, and XE2B, by means of Raman spectroscopy, transmission electron microscopy (TEM) and scanning electron microscopy (SEM) analyses. Furthermore, the electrochemical characterisation of screen-printed electrodes modified with these carbon blacks was carried out by cyclic voltammetry and electrochemical impedance spectroscopy with ferro/ferricyanide as redox probe, highlighting the advantage to use carbon black as nanomodifier in respect to the bare electrode. Among several tested carbon black types, the lowest peak-to-peak separation and resistance to electron transfer values were achieved using screen-printed electrodes modified with CB N115, N375, HP 160 and PL6. The electrodes modified with these types of CB were successively tested in cyclic voltammetry towards epinephrine, benzoquinone, ascorbic acid, cysteine, catechol, and caffeic acid, observing a remarkable improvement of electrochemical performances in respect to the bare electrode, even when the amperometric mode was used. The results obtained demonstrated that several types of CB can remarkably improve the electrochemical performances of the sensors in terms of the decrease of applied potential or peak-to-peak separation, the improvement of the peak intensity, and the decrease of the resistance of the electron transfer due to several key features, including nanodimensions, the onionlike carbon structure, and the high number of defect sites.

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1. Introduction

Recent advances of nanotechnology have boosted the use of nanomaterials as smart components in the design of electrochemical sensors thanks to their countless features, including high surface area, large number of defect sites, easiness of functionalisation, and fast electron transfer. Among carbon-based materials, the discovery of carbon nanotubes in 1991 by Ijima [1] laid out one of the main pillar in nanomaterial field, demonstrating how the carbon materials at nanoscale largely affect several characteristics of electrochemical sensors such as sensitivity, selectivity, tailorability, and robustness.

In 2004, Konstantin S. Novoselov and Andre K. Geim highlighted the features of a thick monocrystalline graphitic film, namely graphene, obtained from pyrolytic graphite by an easy exfoliation technique [2]. This thick monocrystalline graphitic film turned out in one of most promising nanomaterial in several fields including

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the electrochemical sensor one [3-5]. As highlighted in our recent review focused on graphene-based screen-printed electrodes (SPEs), graphene could be seen as a *kaleidoscope* because it confers different features to the sensing tools, being used to tailor the electrochemical properties of printed electrodes as well as to act as a label and loading agent for biomolecules and inorganic nanomaterials [6]. With the aim of employing sustainable and costeffective nanomaterials, since 2010 we have started to use carbon black as nanomodifier for printed sensors, taking into account that few applications were reported in literature until 2010 for analyte detection in solution [7,8]. Carbon black (CB) is an amorphous carbon belonging to pure soot and can be produced by incomplete combustion of carbon e.g. pyrolysis of carbonaceous industrial material. The most relevant production method is the oil furnace process, involving the partial combustion of aromatic liquid hydrocarbons in a furnace, delivering fine black particles with diameters in the range of nm, which form fused aggregates at μ m level. After 2010, some works have demonstrated the fascinating properties of CB as nanomaterial to provide high performing electrochemical sensors. Our group reported the attractive electrochemical properties of CB N220 to be used both in the ink/paste as well as in modifying electrodes by drop-casting for the quantification of several analytes such as phenolic compounds, NADH, thiols, ascorbic acid [9–11]. Its interesting features as nanomodifier for working electrode surface have been successfully demonstrated when compared with the most famous multiwalled carbon nanotubes [12,13] or graphene [13,14]. The enhancement in terms of electron transfer, sensitivity to the detected analyte, resistance to fouling, and decrease of applied potential for analyte quantification has been observed by using different types of CB, such as N220, M1100 [12], 5358 R [14], VXC72R [15], with several electrodes i.e. glassy carbon, carbon paste and screen-printed electrodes [16–20], demonstrating the robustness of this nanomaterial. Furthermore, additional interesting features of this nanomaterial rely on its use without any further chemical treatment (avoiding the use of strong oxidant as in the case of carbon nanotubes), the cost-effectiveness (around $1 \in$ for 1 Kg), and the capability to obtain a stable dispersion useful to modify the electrode by an easily scalable dropcasting method. Although the outstanding electroanalytical properties have been mentioned and demonstrated by several research groups for a wide range of analytes e.g. amino acids [21], organic acid and DNA [22], dopamine and catechol [23], there is still a lack in the understanding the key features for the electrochemical behaviour. To address this issue, herein we report an electrochemical study of several types of carbon black namely HP 160, HS20, MTN 990, N115, N375, N660, PL6, Super P, and XE2B by testing several species i.e. epinephrine, benzoquinone, ascorbic acid, cysteine, catechol, and caffeic acid in cyclic voltammetry. Moreover, a morphological and structural study has been combined with, to investigate the possible correlation between the CB structure and electrochemical behaviour of CB-based printed electrodes.

2. Experimental section

2.1. Materials

Potassium ferrocyanide (K_4 Fe(CN)₆), potassium ferricyanide (K_3 Fe(CN)₆), hexaammineruthenium(III) chloride, sodium phosphate dibasic (Na_2 HPO₄), epinephrine, benzoquinone, ascorbic acid, cysteine, catechol, caffeic acid, and N,N-dimethylformamide were purchased from Sigma-Aldrich (St. Louise, USA). CB N220, N115, N375, and N660 were obtained from Cabot Corporation (Ravenna, Italy). HP 160 and HS20 were purchased by Orion Engineered Carbons (Kingwood, USA). MTN 990, PL6, Super P, and XE2B were

kindly supplied by Prof. Franco Cataldo.

2.2. Screen-printed electrode fabrication

SPEs were produced using a 245 DEK (Weymouth, UK) screenprinting machine. Flexible polyester film (Autostat HT5), purchased from Autotype Italia (Milan, Italy), was used as substrate to print the electrodes. Graphite-based ink (Electrodag 423 SS) from Acheson (Milan, Italy) was used to print both the working and auxiliary electrodes, while silver/silver chloride ink (Electrodag 6038 SS) to print the pseudo-reference electrode, and Grey Dielectric Paste D2070423P5 from Gwent Electronic Materials (Pontypool, UK) for the insulating layer to define the working electrode surface area. The resultant diameter of the working electrode was 0.3 cm with a geometric area equal to 0.07 cm².

2.3. Procedure of CB dispersion

The powder of each CB kind was dispersed in a mixture of dimethylformamide/water in ratio 1:1 v/v for a final concentration of 1 mg/mL. In detail, 10 mg of each CB were first dipped in 5 mL of dimethylformamide and then 5 mL of water were added. The dispersion was sonicated for 60 min at 59 kHz.

2.4. Procedure for SPE modification

SPEs were modified using drop-casting method by adding on the surface of working electrode $6 \,\mu$ L of the dispersion via three successive steps of 2 μ L. The modified SPEs were tested after the evaporation of the solvent used for the dispersion.

2.5. Electrochemical study

Cyclic voltammetry (CV), amperometry and Electrochemical Impedance Spectroscopy (EIS) were performed by using the portable potentiostat PalmSens⁴ (Palm Instrument, The Netherlands) connected to a laptop and controlled by PSTrace software. CV and EIS measurements were carried out using a solution of 5 mM ferro/ferricyanide (1:1 v/v) in KCl 0.1 M. In the case of EIS measurements, a frequency range between 1 Hz and 10 KHz, 10 frequencies per decade, at an open circuit potential, with a voltage amplitude of 0.01 V, was used while data fitting was provided by Z-View software (Scribner Associates Inc.). Cyclic voltammetry was also used as technique to test epinephrine, benzoquinone, ascorbic acid, cysteine, catechol, and caffeic acid at concentration of 2 mM in phosphate buffer 0.05 M + KCl 0.1 M, pH 7.4.

2.6. Microstructural characterisation

Raman analyses were performed to investigate the structure and defectiveness of the CB samples. The Raman spectra were acquired by a Renishaw inVia Reflex Raman microscope (Renishaw plc, Wotton-under-Edge, UK) equipped with a 785 nm solid state laser source. The radiation was focused through a $20 \times$ objective and collected in backscattering configuration, using a laser power of 10 mW and an acquisition time of 50 s. To further reduce the power density, a defocusing level of 70% was selected. Each CB sample was analysed at three different positions and the obtained spectra were fitted and deconvoluted according to the 5 curves method proposed by Sadezky et al. [24] by means of the Raman software WiRE 5.1. The I_D/I_G ratio was calculated starting from the intensities of the D and G curves obtained by the deconvolution procedure.

X-ray photoelectron spectroscopy (XPS) studies were carried out with a PHI 5000 Versaprobe Scanning X-ray photoelectron spectrometer (monochromatic Al K-alpha X-ray source with 1486.6 eV energy) in order to investigate surface chemical composition. A spot size of 100 μ m was used to collect the photoelectron signal for both the survey and the high-resolution (HR) spectra. All samples were analysed using a combined electron and argon ion gun neutralizer system to reduce the charging effect during the measurements. Spectra were analysed using Multipak 9.0 dedicated software. All core-level peak energies were referenced to C1s peak at 284.5 eV and the background contribution in HR scans has been subtracted by means of a Shirley function [25].

The nanoparticles size and shape were investigated with transmission electron microscopy. The powders were dispersed in ethanol and a drop of dispersion was put on a standard holey carbon copper grid. The measurements were carried out with a FEI TECNAI F20ST operating at 200 KV.

The deposition and distribution of the used CB dispersion on the surface of the SPE working electrode were analysed at field emission scanning electron microscopy (FEG-SEM, Leo Supra 35, UK).

3. Results and discussion

3.1. Structural and morphological characterisation

TEM analyses were conducted on the different CBs with the aim to evaluate their morphological and structural properties. In Fig. 1 two magnifications for each type of CB are reported. All the samples exhibit an onion-like carbon structure [26,27]. In details, XE2B powder is composed of hollow polyhedral carbon onions with few (4-8) layers (Fig. 1 L), observing that the layer distance fits that of graphene lavers and particles size is in the 10–20 nm range. PL6 and Super P (Fig. 1 H and 1 I) powders show the same structural characteristics: they consist of multi-layered carbon onion (10-30 layers), as the predominant component, with an amorphous core and the particle size in the range of 20-50 nm. The not regular distance of the layers can be attributed to a reduced degree of graphitization [28]. Sample HP 160 (Fig. 1 A) presents of carbon onions with size in the range of 10-25 nm, composed of tenths of graphitic-like layers, with a very small inner amorphous core and with very irregular interplanar distances. Similar features can be found in samples N115 (Fig. 1 D), N220 (Fig. 1 E) and N375 (Fig. 1 F), and with carbon onions size in the range of 20-40 nm. Samples HS20 (Fig. 1 B), MNT 990 (Fig. 1 C) and N660 (Fig. 1 G) show the biggest particle size, with almost round carbon onion in the 50 nm range for HS20 and N660, and very big round carbon onion in the range of 200-300 nm for MNT 990. They consist of by tenths of graphitic-like layers with irregular interplanar distances. Except for the sample XE2B, all the samples show a remarked content of defects given by the not complete graphitization of the carbon onions. The correlation of these observations with Raman spectroscopy is of paramount importance to confirm and to also establish the role of these defects in the different types of CB.

Indeed, in carbon-based electrodes, the catalytic activity and electron transfer principally happens at the surface defect sites [29], and Raman spectroscopy is an useful technique to evaluate the disorder and imperfection of the carbon materials e.g. basal plane defects. The Raman spectra of the CB samples were therefore collected and are reported in Fig. 2.

All the spectra are characterised by the presence of two main vibrational features around 1580 and 1350 cm^{-1} . The first band corresponds to the E_{2g} mode of graphite (G mode), due to the inplane vibration of the carbon-carbon bonds of the graphitic layers, while the second one (D mode), assigned to the breathing of the 6-membered rings, only arises in the presence of defects which disrupt the symmetry of the sp² systems [30,31]. Three additional disorder-induced bands are identified: the D' band around 1615 cm⁻¹, the D3 band around 1500 cm⁻¹, associated to the



Fig. 1. TEM images of CB HP 160 (A), HS20 (B), MTN 990 (C), N115 (D), N220 (E), N375 (F) N660 (G), PL 6 (H), Super P (I), XE2B (L). (scale: 20 or 50 nm, 5 nm or 10 nm for the inset).

presence of amorphous carbon, and the usually weak D4 band at about 1200 cm⁻¹. Their presence is pointed out by the deconvolution procedure of the first order Raman spectra of the CB samples according to the procedure optimized by Sadezky et al. [24]. Three broad and often unresolved bands appear instead in the high Raman shift range and correspond to the second order modes of the D (2D, at 2675 cm^{-1}) and D' (2D', at 3180 cm^{-1}) bands and to a combination mode of the D and G bands $(D+G, at 2915 \text{ cm}^{-1})$ [32]. Despite the common vibrational pattern of the CB samples, some differences in the relative intensity ratios and band widths are noticed. In particular, the I_D/I_G ratio is a useful parameter to evaluate the level of order/disorder in the graphitic carbon and its values calculated for the CB samples, are listed in Table 1. As a general trend, the nanometric CB particles display higher I_D/I_G ratios with respect to the bigger ones, probably because of the combination of a greater number of defects and an increased distortion of the graphitic-like domains at the surface of the small nanoparticles. Indeed, quite similar ratios, i.e. around 1.3 were detected for most of the samples, except for the N660 and MTN 990 powders, whose particle size is the largest, as evidenced from the related TEM images (Fig. 1C and 1 F), resulting in the lowest I_D/I_G ratios (around 1.2). Instead, the highest I_D/I_G was measured for the



Fig. 2. Normalized average Raman spectra of the CB powders.

Table 1

Average D and G band positions and I_D/I_C ratios obtained by the deconvolution of the first order Raman spectra of the CB powders.

Sample	G (cm ⁻¹)	D (cm ⁻¹)	I _D /I _G
HP 160	1577.0 ± 0.1	1346 ± 2	1.28 ± 0.20
PL6	1575 ± 5	1342 ± 3	1.41 ± 0.04
N220	1580.0 ± 0.8	1349 ± 1	1.28 ± 0.06
N115	1578 ± 2	1348.0 ± 0.5	1.33 ± 0.02
N375	1577.0 ± 0.6	1348 ± 1	1.27 ± 0.01
XE2B	1584 ± 2	1342 ± 1	1.57 ± 0.06
N660	1577.0 ± 0.2	1355 ± 3	1.22 ± 0.10
Super P	1577.0 ± 0.7	1347.0 ± 0.5	1.41 ± 0.03
HS20	1582.0 ± 0.9	1358.0±0.2	1.32 ± 0.02
MTN 990	1587 ± 2	1357.0 ± 0.9	1.19 ± 0.08

XE2B, followed by the Super P and the PL6 samples. The increase of the relative intensity of the defect-induced D band for these particles is not only related to their small dimension, but also to their different structural features, as highlighted by a spectral profile characterised by narrow D and G bands. Indeed, it is well known that an increase of the I_D/I_G ratio can be correlated to a decrease of the size of the graphitic-like domains [33]. The sharpness of the D and G bands witnesses, however, a globally increased degree of crystalline order for such samples, consistently with a drop of the spectral intensity between the D and G bands. Such a decrease is due to a reduced contribution in this region of the D3 disorderinduced mode whose intensity was proposed to be indicative of the amorphous carbon content by previous studies on carbonaceous materials [32]. The greater graphitic fraction of these samples is also confirmed by an analogous narrowing of the band widths, observed in the second order region.

To establish the effect of the elemental composition of different CB samples, XPS analysis was carried out. Undeniably, XPS measurements seek to obtain information regarding the chemical state of each of the CB samples involved in this study, since electrochemical performances are deeply related with materials surface chemistry. From survey spectra (Fig. S1) we have obtained

Table 2	
XPS relative atomic concentration (at.%), obtained from HR spec	tra.

Sample	C1s	O1s
HP 160	90.1	9.9
PL6	97.3	2.7
N220	98.3	1.7
N115	98.4	1.6
N375	96.7	3.3
XE2B	97.8	2.2
N660	98.1	1.9
Super P	99.3	0.7
HS20	97.0	3.0
MTN 990	97.6	2.4

information about elemental composition and relative atomic concentration of each present species (Table 2). The O concentration varies from 9.9 at.% for sample HP160 to 0.7 for SUPER P sample: in between the average O concentration is almost 2 at.%. Then, we focused our attention on C1s photoelectron peaks, to evaluate the chemical shifts due to the C-O bonds. After a deconvolution procedure applied on all the C1s HR spectra (by using a pseudo-Voigt function for each peak), we obtained six components for all the samples apart from HP160, who needed an extra component (C–C sp¹ at 283.5 eV) [34] to reproduce the experimental curve. The chemical shift found are: $C-C \text{ sp}^2$ (284.5 eV), C-C sp³ (285.2 eV), C-O-C/H (286.0 eV), O-C-O (287.4 eV), -O-C=O- (289.2 eV) and $\pi - \pi^*$ shake up satellite (>291 eV) [35]. All the chemical shift percentages are reported in Table 3.

Finally, in order to investigate the deposition and the distribution of the used CB types on the surface of the working electrode, SEM analysis was carried out on all SPEs. From the comparison among the SEM micrographs reported in Fig. 3, it is evident that, in all cases, a continuous and uniform layer of CB particles was obtained, which completely covered the underneath randomly orientated micrometric graphite flakes, evidenced in the SEM micrograph of bare SPE (Fig. 3 A, B). Indeed, the bare SPE presented graphite flakes (Fig. 3 A), characterised by numerous small particles consisting in the cross-linking agents present in the original ink (Fig. 3 B) [36]. Thus, these experimental evidences confirmed the obtainment of a good dispersion and the efficacy of the used dropcasting technique. However, in all cases it is possible to observe the presence of CB agglomerates, as expected: due to its very high specific surface area, CB primary particles spontaneously tend to generate nanostructured aggregates which can form agglomerates, leading to distinct and characteristic porous structures based on nanometric spherical particles [37]. From the reported SEM micrographs, it is evident that the samples HP 160, N115, N220, N375, PL6, Super P and XE2B appear comparable, due to the similarity of the used CB powders, composed of nanometric particles (with an average diameter size of 20-40 nm) which tend to aggregate due to their very high specific surface area, in very good agreement with TEM results (Fig. 1). On the other hand, the samples HS20, MTN 990 and N660 were characterised by bigger, more heterogeneous and more rounded particles, as evident from the higher magnifications micrographs (insets of Fig. 3 D, E and I, respectively), particularly in the case of MTN 990 modified SPE, being MTN 990 powder composed of heterogeneous particles with different diameter sizes (ranging between 100 nm and 300 nm). In these latter cases, the bigger dimension of the used particles led to the formation of a less uniform layer, characterised by diffuse voids (Fig. 3 D, E and I, insets). Accordingly, the CB layers resulted more compact and uniform in the case of smaller CB particles modified SPEs (Fig. 3C, F, H, L, M, N), as expected.

Table 3		
XPS C1s chemical shifts assignments, obtained from deconvolution procedures (not rep	ported)

Sample	C1s components							
	283.5 eV C—C sp ¹	284.5 eV C—C sp ²	285.2 eV C—C sp ³	286.0 eV C-O-C/H	287.4 eV 0-C-0	289.2 eV -0-C=0-	291.0 eV $\pi-\pi^*$ shake up	
HP 160	19.99%	52.34%	9.07%	3.64%	9.26%	2.19%	3.50%	
HS20	_	67.65%	12.95%	7.99%	4.22%	2.95%	4.24%	
MTN 990	_	67.90%	16.01%	6.11%	3.58%	1.83%	4.58%	
N115	_	63.73%	12.47%	8.74%	4.64%	3.99%	6.43%	
N220	-	65.51%	8.87%	9.12%	5.53%	4.07%	6.91%	
N375	_	62.59%	14.49%	8.80%	5.55%	2.94%	5.63%	
N660	_	67.25%	11.84%	8.17%	4.27%	2.88%	5.59%	
PL6	_	64.00%	11.52%	9.33%	5.23%	3.69%	6.23%	
Super P	_	67.97%	13.71%	5.78%	4.41%	2.71%	6.41%	
XE2B	-	59.36%	19.23%	5.44%	5.14%	3.54%	7.30%	



Fig. 3. SEM micrographs of the bare SPE (A–B) and of the SPEs modified with HP 160 (C), HS20 (D), MTN 990 (E), N115 (F), N220 (G), N375 (H), N660 (I), PL6 (L), Super P (M), XE2B (N) (insets: higher magnification micrographs).

3.2. Electrochemical characterisation using ferro/ferricyanide as redox probe

In order to evaluate the electrochemical performances of CB as nanomaterial for electrode fabrication, the most common approach relies on the modification of well-known characterised electrode materials by drop casting. To explore the response of CB-modified electrode surface in function of coverage effect, which can affect the electrochemical performances [7], the response of ferro/ferricyanide was evaluated after having modified the working electrode surface with 2, 4, 6 and 8 µL of different CB dispersions at concentration of 1 mg/mL. As depicted by Fig. 4, generally at increasing of CB amount a decrease of peak to peak separation was observed, in combination with the increase of peak intensity, due to the higher amount of CB cast on the working electrode surface. Taking into account the electrochemical performances as well as the time of modified electrode preparation, 6 µL (means 6 µg of CB) was selected as optimal volume of dispersion at 1 mg/mL to cast on the working electrode surface, in agreement with our previous work using only CB N220 [11]. After having selected the amount of CB, cyclic voltammetric measurements were carried out to obtain deeper information about the electrochemical properties of the different types of CB used as electrode modifier, except for CB N220 being previously largely characterised in our previous studies [9-11].

Fig. 5 shows the voltammograms obtained in ferro/ferricyanide 5 mM + KCl 0.1 mM, by using SPEs modified with the selected CBs. Different behaviours were observed depending on the CB used to modify SPEs by drop-casting method. SPE modified with MTN 990 gave the largest peak to peak separation (i.e. 470 mV) even when compared with bare SPE, and this behaviour is ascribed to its structure, which is not at the nanodimension level. A peak-to-peak separation comprised between 160 and 180 mV was observed with SPEs modified with CB N660, N375, Super P. The smallest peak to peak separations were observed in the case of SPEs modified with N115 and HP 160, being equal to 120 ± 7 and 129 ± 6 mV, respectively.

To quantify the effect of CB types on the electron transfer, the heterogeneous rate constant (k°) for the ferro/ferricyanide redox probe was calculated based on Nicholsons theory [38], using $D_0 = D_R = 5.9 \times 10^{-5} \text{ cm}^2/\text{s}$ and an electrochemical transfer coefficient $\alpha = 0.5$, taking into account that the I_{ap}/I_{cp} value is very close to the unity for bare and CB modified SPEs [39]. The heterogeneous rate constants, reported in Table 4, highlighted the fastest electron transfer in the case of SPEs modified with N115 and HP 160, giving a value of $(6.3 \pm 0.6) \times 10^{-3}$ and $(5.4 \pm 0.9) \times 10^{-3}$ cm/s, respectively, which are comparable to the one obtained using CB N220 as modifier (4.5×10^{-3} cm/s) using the same solvent for the dispersion and the same batch of CB [13]. In contrast, the lowest value of



Fig. 4. Value for anodic peak current (black dots) and peak to peak separation (red dots) for SPE modified with 2 μ L, 4 μ L, 6 μ L and 8 μ L of CB dispersion HP 160 (A), HS20 (B), MTN 990 (C), N115 (D) N220 (E), N375 (F), N660 (G), PL6 (H), Super P (I), XE2B (L). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

heterogeneous rate constant was obtained using SPEs modified with HS20 $(1.2\pm0.3)\times10^{-3}\,cm/s.$

Analysing the intensity of the anodic and cathodic peaks, the lowest intensities were observed using SPEs modified with CB Super P, N660, N375, HS20, MTN 990 with the anodic peak current equal to 73 ± 1 , 75 ± 6 , 73 ± 3 , 73 ± 1 , and $42 \pm 7 \mu$ A, respectively. In contrast, higher peak intensities were found using SPEs modified with HP 160, N115, PL6 and XE2B (i.e. 91 ± 6 , 86 ± 1 , 89 ± 6 , and $88 \pm 2 \mu$ A).

The enhancement of the intensity of the anodic and cathodic peaks in the case of SPEs modified with these latter CBs is ascribed to the higher electroactive surface area calculated using Randles-Sevcik equation with the diffusion coefficient equal to $D = 5.9 \times 10^{-5} \text{ cm}^2/\text{s}$ [40]. Indeed, these CB-SPEs were characterised by electroactive surface areas equal to 3.8 ± 0.2 , 3.6 ± 0.2 , 3.7 ± 0.3 , and $3.7 \pm 0.2 \text{ mm}^2$, respectively, higher values when compared with the other ones reported in Table 4 for the remaining CB-SPEs. In addition, as expected, the electroactive areas resulted higher than the one reported for the bare electrode, $1.6 \pm 0.2 \text{ mm}^2$ calculated in our previous work [40].

Furthermore, the voltammetric peak intensity for each CB-SPE was evaluated as a function of the scan rate and plotted versus the square root of the scan rate ($v^{0.5}$). Fig. S2 showed linear



Fig. 5. Cyclic voltammetry measurements performed in $[Fe(CN)_6]^{3-/4-}$ 5 mM + KCl 0.1 M, obtained with bare SPEs (dashed black line) and SPEs modified with HP 160 (green line), HS20 (dashed green line), MTN 990 (dashed red line), N115 (grey line), N375 (black line), N660 (red line), PL6 (cyan line), Super P (dashed blue line) and XE2B (blue line). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

behaviour for bare SPE and for each modified CB-SPE, indicating that the process is fully controlled by the diffusion of the electroactive species. The following slopes $(mA/(mVs^{-1})^{0.5})$ were obtained: 3.6 ± 0.2 ($R^2 = 0.983$), 13.6 ± 0.4 ($R^2 = 0.992$), 10.3 ± 0.4 ($R^2 = 0.998$), 3.6 ± 0.2 ($R^2 = 0.937$), 12.7 ± 0.3 ($R^2 = 0.996$), 14.9 ± 0.3 ($R^2 = 0.993$), 11.0 ± 0.4 ($R^2 = 0.990$), 12.8 ± 0.4 ($R^2 = 0.981$), 9.0 ± 0.4 ($R^2 = 0.987$), 10.6 ± 0.6 ($R^2 = 0.976$), respectively for bare SPE, HP 160, HS20, MTN 990, N115, N375, N660, PL6, Super P, and XE2B. The highest slope was observed using HP 160, confirming its better electrochemical properties.

Additionally, we have analysed the variation of log *Ip* versus log *v* using ferro/ferricyanide as redox probe, obtaining the following gradients 0.52 ± 0.03 , 0.51 ± 0.03 , 0.52 ± 0.02 , 0.52 ± 0.03 , 0.50 ± 0.02 , 0.52 ± 0.03 , 0.54 ± 0.03 , 0.50 ± 0.03 , 0.48 ± 0.03 , respectively for HP 160, HS20, N115, N220, N375, N660, PL6, Super P, and XE2B (Fig. S3). The values obtained, close to 0.5, suggested no thin-layer effects, ensuring the absence of the redox probe entrapped within the network of the film. Therefore, a purely diffusional (semi-infinite) electrode response was observed. While in the case of bare SPE and SPE modified with MTN 990, a different gradient equal 0.37 ± 0.02 and 0.37 ± 0.03 was found, respectively, demonstrating a dissimilar behaviour in respect to nanomodified electrode.

To deeply investigate the electrochemistry of the different CBs, Electrochemical Impedance Spectroscopy (EIS) was used, obtaining information on the impedance changes of the electrode surface by varying the CB modifier (Fig. 6). Indeed, R_{ct} value gives the difficulty of electron transfer of ferro/ferricyanide redox probe between the solution and the electrode, and its measurement can highlight the capability of CB to improve the electron transfer to the redox probe. Using this technique, the higher R_{ct} values were observed in the case of SPEs modified with MTN 990, Super P and HS20 with R_{ct} equal to $4 \pm 2 M\Omega$, $1.3 \pm 0.5 M\Omega$, and $1.2 \pm 0.2 M\Omega$, respectively, while the lowest R_{ct} using SPEs modified with HP 160, N115 and PL6 (51 \pm 8, 51 \pm 6 and 55 \pm 7 Ω), confirming the behaviour of cyclic voltammetric experiments.

3.3. Reproducibility and storage stability of CB dispersion

One of the main advantageous key feature of CB is the facile preparation of a stable dispersion assisted by just one-step of sonication (60 min at 59 kHz) without any further chemical and

N220 $1.59 \times 10^{-2} [10], 4.5 \times 10^{-3} [13], \\ 1.4 \times 10^{-2} [40]^{6}, 1.85 \times 10^{-2} [41], \\ 3.6 \times 10^{-3} [42]^{c}$ $124 [45,43], \\ 120 [10]$ $3.2 [40], 9.3 [44], \\ 23 [41], 24 [42], \\ 3.1 [this work]$ $17.95-32.5 [10], 19-29 [43]^{3}, \\ 19-25 [44], 19-29 [45]^{3}, 25 [16]$ $0.96 [10], \\ 0.97 [13], \\ 1.28 [this work]$ CAT [43], NADH [9,10,13,42], BQ [10], Hg [50], BPA [54], Glucose [44], HQ [44], Hg [50], BPA [54], Glucose [44], HQ [44], Hg [54], Glucose [46], Hg [56], Glucose [46]	Carbon Black	k ⁰ (cm/s)	Surface Area (BET) (m/g)	Electroactive Area (mm ²)	Particle diameter (nm)	I_D/I_G	Analytes
VXC72R $9.26 \times 10^{-2} [16]^d, 9.77 \times 10^{-3} [48]^e$ $52 [17], 20-25 [56], 32.1 [16]$ $2.64 [16,17], 1.01 [56], 0.99 [58]$ DA [16,17,58,59], APAP [16,17,47,59,60], AA [17], UA [17], LEV [47], Mesotrione [48] Estricl [56], AM [57], NI [57], EP [58], IP [60], PP [60], CAF [60], CAT [61]N375 $3 \times 10^{-3} [this work]$ $90 [10], 14 [63], 3.1 \pm 0.1$ $36 [62,63]$ $1.27 [this work]$ $NADH [10], Sulfonamides [62], Nitrite [63], BQ [63], HQ [63]N77230 [10,62,63]8 [63]124 [62,63]1.24 [62,63]NADH, Sulfonamides [62], Nitrite [63], BQ [63], HQ [63]N1156.3 \times 10^{-3} [this work]19 [62], 3.6 \pm 0.2 [this work]1.33 [this work]DA [7], AA [7,8], Pb [7], Cd [7], Cu [8] [M 430Printex XE 28.1 \times 10^{-3} [8]95030 [7] [25-37 [20]DA [7], AA [7,8], Pb [7], Cd [7], Cu [8] [FM [20], APAP [20], Phenolic compound [64] [Start]$	N220	$\begin{array}{l} 1.59\times10^{-2}\ [10],\ 4.5\times10^{-3}\ [13],\\ 1.4\times10^{-2}\ [40]^b,\ 1.85\times10^{-2}\ [41],\\ 3.6\times10^{-3}\ [42]^c \end{array}$	124 [45,43], 120 [10]	3.2 [40], 9.3 [44], 23 [41], 24 [42], 3.1 [this work]	17.95–32.5 [10],19–29 [43] ^a , 19–25 [44], 19–29 [45] ^a , , 25 [16]	0.96 [10], 0.97 [13], 1.28 [this work]	CAT [43], NADH [9,10,13,42], BQ [10], EP [10], H ₂ O ₂ [10,44,49], Heme protein [40], Hg [50], BPA [51,52], As [53], AA [44], Cys [13], Phosphate [54], Glucose [44],HQ [44], Free Chlorine [55]
N375 3×10^{-3} [this work] 90 [10], 14 [63], 3.1 \pm 0.1 105 [62,63] 36 [62,63] 1.27 [this work] NADH [10], Sulfonamides [62], Nitrite [63], BQ [63], HQ [63] N772 30 [10,62,63] 8 [63] 124 [62,63] NADH, Sulfonamides [62], Nitrite [63], BQ [63], HQ [63] N115 6.3×10^{-3} [this work] 19 [62], 3.6 ± 0.2 [this work] 1.33 [this work] DA [7], AA [7,8], Pb [7], Cd [7], Cu [8] Printex XE 2 8.1×10^{-3} [8] 950 30 [7] DA [7], AA [7,8], Pb [7], Cd [7], Cu [8] M 430 25-37 [20] 57.7 [20] DA [7], AA [7,8], Pb [7], Cd [7], Cu [8]	VXC72R	$9.26\times 10^{-2}~[16]^d,9.77\times 10^{-3}~[48]^e$			52 [17], 20–25 [56], 32.1 [16]	2.64 [16,17], 1.01 [56], 0.99 [58]	DA [16,17,58,59], APAP [16,17,47,59,60], AA [17], UA [17], LEV [47], Mesotrione [48], Estriol [56], AM [57], NI [57], EP [58], IP [60], PP [60], CAF [60], CAT [61]
N772 30 [10,62,63] 8 [63] 124 [62,63] NADH, Sulfonamides [62], Nitrite [63], BQ [63], HQ [63] N115 6.3 × 10 ⁻³ [this work] 19 [62], 3.6 ± 0.2 [this work] 1.33 [this work] Printex XE 2 8.1 × 10 ⁻³ [8] 950 30 [7] [25-37 [20] DA [7], AA [7,8], Pb [7], Cd [7], Cu [8] [52-37 [20]	N375	3×10^{-3} [this work]	90 [10], 105 [62.63]	$14 [63], 3.1 \pm 0.1$ [this work]	36 [62,63]	1.27 [this work]	NADH [10], Sulfonamides [62], Nitrite [63], BO [63], HO [63]
N115 6.3 × 10 ⁻³ [this work] 19 [62], 3.6 ± 0.2 [this work] 1.33 [this work] Printex XE 2 8.1 × 10 ⁻³ [8] 950 30 [7] 25-37 [20] DA [7], AA [7,8], Pb [7], Cd [7], Cu [8] M 430 25-37 [20] FM [20], APAP [20], Phenolic compound [64]	N772		30 [10,62,63]	8 [63]	124 [62,63]		NADH, Sulfonamides [62], Nitrite [63], BO [63], HO [63]
Printex XE 2 8.1 × 10 ⁻³ [8] 950 30 [7] DA [7], AA [7,8], Pb [7], Cd [7], Cu [8] M 430 25-37 [20] FM [20], APAP [20], Phenolic compound [64]	N115	6.3×10^{-3} [this work]		19 [62], 3.6 ± 0.2 [this work]		1.33 [this work]	
	Printex XE 2	8.1×10^{-3} [8]	950	. ,	30 [7]		DA [7], AA [7,8], Pb [7], Cd [7], Cu [8]
ECP-600JD 1270 [65] 34 [65] Paraoxon [65] M 1100 14 [12] Nicotine [12]	ECP-600JD M 1100		1270 [65]		34 [65] 14 [12]		Paraoxon [65] Nicotine [12]
5358R 1.7×10^{-3} [14] ^f 29 [14] 0.91 [14]	5358R	$1.7 imes 10^{-3} \ [14]^{ m f}$			29 [14]	0.91 [14]	
BP4750 5.74×10^{-3} [16] ^d 26 [16] 2.36 [16] DA [16], APAP [16]	BP4750	$5.74 imes 10^{-3}$ [16] ^d			26 [16]	2.36 [16]	DA [16], APAP [16]
E2000 $6.23 \times 10^{-4} [16]^{d}$ 27 [16] 3.65 [16] DA [16], APAP [16]	E2000	$6.23 imes 10^{-4} \ [16]^{d}$			27 [16]	3.65 [16]	DA [16], APAP [16]
Super P Li 50.84 [66]	Super P Li	2		50.84 [66] ^r			
HP 160 5.4×10^{-3} [this work] 3.8 ± 0.2 [this work] 1.28 [this work] AA, EP, CYS, BQ, CA, CAT [This work]	HP 160	5.4×10^{-3} [this work]		3.8 ± 0.2 [this work]		1.28 [this work]	AA, EP, CYS, BQ, CA, CAT [This work]
HS20 1.2×10^{-5} [this work] 3.0 ± 0.2 [this work] 1.32 [this work]	HS20	1.2×10^{-3} [this work]		3.0 ± 0.2 [this work]		1.32 [this work]	
$\frac{1}{2} \int \frac{1}{2} \int \frac{1}$	MIN 990	$\frac{1}{2}$ (10^{-3}) [this work]		1.7 ± 0.1 [this work]		1.19 [this work]	
NOOU 2.0×10^{-1} [IIIS WOIK] 3.2 ± 0.2 [IIIIS WOIK] 1.22 [IIIIS WOIK]	NOOU DLC	2.6×10^{-5} [this work]		3.2 ± 0.2 [LIIIS WORK]		1.22 [UIIS WORK]	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	FLO Super D	4.0×10^{-3} [this work]		3.7 ± 0.5 [UIIS WOFK] 3.1 ± 0.1 [this work]		1.41 [UIIS WOIK]	
Superior 2.2×10^{-1} [mis work] 3.1 ± 0.1 [mis work] 1.41 [mis work] 1.42×10^{-3} [this work] 3.1 ± 0.2 [this work] 1.57 [this work]	XF2R	4.2×10^{-3} [this work]		3.1 ± 0.1 [this work] 3.7 ± 0.2 [this work]		1.57 [this work]	

Table 4 Electrochemical and structural features of several CB types.

 a Given by the company. b Using $D_0 = D_R = 5.9 \times 10^{-6} \mbox{ cm}^b/s.$ c Using $D_0 = D_R = 7.6 \times 10^{-6} \mbox{ cm}^b/s.$

^d Using ruteniumexaammine.

^e Calculated by impedance measurements.

^f Calculated by chronocoulometry. AA = Ascorbic Acid, AM = Amoxicillin, APAP = acetaminophen, BQ = Benzoquinone, BPA = Bisphenol A, CA = Caffeic acid, CAF = Caffeine, CAT = Catechol, CYS = Cysteine, DA = Dopamine, FA = Folic Acid, EP = Epinephrine, FM = Ferrocenemethanol, IP = Isoproterenol, LEV = Levoflaxacina, NI = Nimesulide, PP = Propranolol, TRP = Tryptophan, TYR = Tyrosine.



Fig. 6. Nyquist plot of EIS measurement obtained with bare SPEs (dashed black line) and modified SPEs with HS20 (dashed green line), MTN 990 (dashed red line), Super P (dashed blue line), and N660 (red line). Inset: Nyquist plot at high frequencies for EIS measurement obtained with SPEs modified with HP 160 (green line), N115 (grey line), N375 (black line), PL6 (cyan line), XE2B (blue line). Measurements performed in $[Fe(CN)_6]^{2-/4-5}$ m M + KCl 0.1 M at open circuit potential. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

physical treatment. The easy way to prepare a stable dispersion allows for a reproducible dispersion as well as reproducible CBmodified electrodes. The reproducibility of CB dispersion was investigated by modifying SPE with three different dispersions of each CB samples and by testing them with ferro/ferricyanide as redox probe. The results reported in Fig. S4 demonstrated similar electrochemical response towards the redox probe investigated within the experimental error, highlighting the reproducibility of the different CBs dispersion, even when diverse CB samples were used. The good reproducibility proved the robustness of the dispersion procedure preparation as well as the process of CBmodified electrode production, establishing a well-known route for CB-modified electrode fabrication.

Almost all CB samples gave a homogeneous dispersion by using just one-step of sonication, except XE2B and PL6, which required further sonication time. The homogeneity of the dispersions using different CB samples was evaluated in the same day of preparation and after 1 week, demonstrating good storage stability for almost all CB samples tested, except in the case of XE2B and PL6, for which a poor uniformity of the layer cast on the filter paper was observed (Fig. S5). The stability of CB dispersions has been also evaluated by drop-casting the dispersion with two-week as life-time, maintained at room temperature without any additional sonication step. In Fig. S4, the response of CB-modified SPEs prepared using a dispersion with 1 h (continuous black line) or two weeks as life-time (dashed black line) sheds light a similar behaviour, demonstrating the good storage stability of almost all CB dispersions.

3.4. Electrochemical behaviour of epinephrine, benzoquinone, ascorbic acid, cysteine, catechol and caffeic acid

To get a further insight into CB types that gave the best performances using ferro/ferricyanide probe (i.e. HP 160, N115, N375 and PL6), the modified SPEs were challenged towards some interesting compounds for biological and environmental applications namely epinephrine, benzoquinone, ascorbic acid, cysteine, catechol and caffeic acid. Fig. 7 sheds light the improvement of all modified SPEs when compared with bare SPE (black dashed line), demonstrating the electrocatalytic properties of the different CBs tested.

For epinephrine, caffeic acid, benzoquinone, and catechol a decrease of peak to peak separation was observed together with an increase of the peak intensity. In the case of the epinephrine and



Fig. 7. Cyclic voltammograms obtained in phosphate buffer 0.05 M + KCl 0.1 M, pH 7.4, in presence of 2 mM of epinephrine (A) benzoquinone (B), ascorbic acid (C), cysteine (D), catechol (E) and caffeic acid (F). All recorded at a scan rate of 50 mV/s using bare SPEs (dashed line) and SPEs modified with HP 160 (green line), N115 (yellow line), N375 (black line), and PL6 (cyan line). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

caffeic acid a smaller peak to peak separation combined with higher anodic and cathodic peak intensity was observed for SPEs modified with HP 160 and N115. For instance, the cyclic voltammetries of epinephrine using HP 160 and N115 gave a peak to peak separation equal to 422 ± 6 and 419 ± 1 mV and the anodic peak intensity equal to 111 ± 4 and $101 \pm 7 \mu$ A, respectively. In the case of benzoquinone and catechol the best electrochemical performances were observed using HP 160 and N375 as nanomodifier in terms of peak to peak separation (i.e. 109 ± 1 and 220 ± 20 in case of benzoquinone) and peak intensities (i.e. 36 ± 2 and $28 \pm 2 \mu$ A in case of benzoquinone), respectively. For the irreversible redox reaction, as the case of ascorbic acid and cysteine, the presence of CB to modify working electrode surface largely decreased the applied potential allowing for the detection of the analyte at a lower applied potential as well as an improvement of sensitivity. In the case of ascorbic acid, the presence of the HP 160 and N115 gave the best resolution of the peaks and the lowest applied potential, while in the case of cysteine we observed a similar behaviour for all CBs tested. The achieved results demonstrated that CB positively affects the detection of the analytes decreasing the peak to peak separation in the case of (semi)reversible reaction or decreasing the applied potential, demonstrating the electrocatalytic properties of CB. In addition, in all cases we observed an increase of the sensitivity due to its high surface.

3.5. Electroanalytical characterisation

SPEs modified with the most performant CBs, namely HP 160 and N115, were tested in amperometric mode toward ascorbic acid and cysteine, analytes chosen for their importance in biological field as well as for their features in electrochemistry (e.g. fouling problem, high applied potential). In addition, to compare the behaviour with the well-known N220, the amperometric analyses were performed by using also SPEs modified with this latter CB. For the amperometric detection, applied potentials equal to +50 mV and +400 mV were chosen for ascorbic acid and cysteine detection, respectively, taking into account the previous cyclic voltammetries. As depicted in Fig. 8, for both ascorbic acid and cysteine detection, the presence of CB largely improves the sensitivity of the measurements in respect to the bare SPE (black line), overcoming the fouling problem, indeed successive additions of the analyte gave almost the same signal intensity. In the case of ascorbic acid detection (Fig. 8 A), higher sensitivity was obtained by using SPEs modified with N220 (0.021 \pm 0.001 μ A/ μ M) when compared to the ones obtained using SPEs modified with HP 160 and N115 (i.e. $0.016 \pm 0.002 \,\mu$ A/ μ M and $0.0196 \pm 0.0003 \,\mu$ A/ μ M). Limit of detection (calculated as S/N = 3) was found equal to 0.1 μ M, 0.4 μ M and 0.3 µM for the SPEs modified respectively with N220, HP 160 and N115.

As for ascorbic acid detection, in the case of cysteine measurement, SPEs modified with CB N220 resulted in higher sensitivity, i.e. $(0.0217 \pm 0.0009) \ \mu A/\mu M$, compared with sensitivities obtained by using HP 160 and N115 modified SPEs, respectively equal to $(0.017 \pm 0.001) \ \mu A/\mu M$ and $(0.0161 \pm 0.0008) \ \mu A/\mu M$. The calculated LOD was 0.3 μ M, 1 μ M and 0.6 μ M for the SPEs modified respectively with N220, HP 160 and N115, confirming the better performances of N220.

3.6. Discussion

The morphological and structural characterisation combined with electrochemical studies with ferro/ferricyanide as redox probe, highlighted that the nanodimension of CB positively affects the electrochemical behaviour of the sensor when it is used as working electrode modifier. Undeniably, CB characterised by large particles gave only a smaller improvement of the electrochemical performances of electrodes: in the case of micromaterial MTN 990, modified electrode with CB MTN 990 gave similar performances in respect to the bare electrode, confirming the advantage of the nanodimension over the microdimension. Additionally, the case of HS20 and N660, with almost round carbon onion in the 50 nm range, a higher peak to peak separation and resistance to electron transfer (R_{CT}) with lower peak intensity were observed, in respect to electrodes modified with CBs characterised by smaller nanoparticles. The results achieved demonstrated that the structure affects the electrochemical response: among the different CB samples, the ones with carbon onions size in the range of 10–25 nm, composed of tenths of graphitic-like layers with a very small inner amorphous core and very irregular interplanar distances, namely HP 160, N220 and N375, provided the best electrochemical performances. As reported in literature [29], the defect



Fig. 8. Amperograms obtained in phosphate buffer 0.05 M + KCl 0.1 M, pH 7.4, after successive addiction (25 μ M) of ascorbic acid (A) and cysteine (B), for bare SPE (black line) and modified with N220 (green line), N115 (blue line) and HP 160 (red line). Applied potentials equal to +50 mV and +400 mV to detect ascorbic acid and cysteine, respectively. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

sites play a crucial role in electrocatalysis. CB samples are characterised by high number of defect sites with I_D/I_G equal to 1.19 or higher, but the defect sites are not the only key features: XE2B has the highest I_D/I_G with smaller particles size in the 10–20 nm range, with hollow polyhedral carbon onions with few (4-8) layers, but it is not the most performant CB among the CB tested. Another relevant issue regards the presence of surface functional groups e.g. C=0, O=C-0, which highly affect the electrochemical response as demonstrated using inner-sphere probe i.e. ferro/ferricyanide. Otherwise, using no-sensitive surface electrochemical redox system i.e. hexaammineruthenium(III) chloride (outsphere probe) [67], the peak to peak separation is similar among the several CB types, having similar electronic structure. However, the higher peak to peak separation was observed in case of MTN 990, which has microstructure, while low values were observed with CB having smaller diameter of the nanoparticles (e.g. CB N115, HP 160, N220) (Table S1). Additionally, XPS analysis gave information related to the effect of C/O group on the electrochemical behaviour. HP 160, which is one the most promising CB, is characterised by the higher content in O atoms (9.9 at.%), while in the remnant samples the average O content is around 2.2 at.%. Moreover it is the only one which possesses a chemical shift in the C1s photoelectron peak, due to sp¹ hybridization (19.99%), together with sp² (52.34%) and $sp^{3}(9.07\%)$, which are also present in the other samples. The more intense C-O bond is the one due to O-C-O chemical shift at 1287.4 eV (9.26%), while an average value for the other samples is 4.7%. Among the C/O groups, the CB with higher content of -O-C=O (i.e. N115 3.99% and N220 4.07%) is characterised by better electrochemical performances, demonstrating that O content and, in detail the -O-C=O amount, can give an improvement because of particular electrochemical electrocatalytic interaction with these specific surface oxygenated groups/species [16].

4. Conclusions

Herein, we reported a comparison among SPEs modified with several types of CB to investigate the influence of the morphological and structural characteristics of the investigated CBs on the electrochemical behaviour. The nanodimension of CB allows for the development of highly sensitive sensor when compared with the bare one, with the advantage to be a cost-effective nanomaterial, to do not require any treatment before the use, and to easily furnish a stable dispersion needed for a reproducible and easy scalable dropcasting procedure. Several CBs were identified as the best candidates for the modification of the electrode surface such as HP 160, N115, N220, N375 and PL6, demonstrating low peak to peak separation in the case of (semi)reversible species and low applied potential in the case of irreversible compounds, always associated to an increase of sensitivity. Because the well-established electrochemical properties combined with advantageous features reported above, several CBs deserve to be used as nanomodifier of electrochemical sensors. The reasons for these outstanding electrochemical properties are ascribed to nanodimensions onion-like carbon structure (carbon onions size in the range of 10-25 nm), high number of defect sites $(I_D/I_G$ ratios equal to 1.3 or higher), higher content in O atoms (especially –O-C=O groups). However, these characteristics are not the only ones able to furnish the high performant electrochemical behaviour of CB-modified electrodes, opening further questions about still undiscovered reasons.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.electacta.2019.05.117.

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