# PARP inhibitors in cancer therapy: magic bullets but moving targets

# Girish M. Shah<sup>1</sup>\*, Mihaela Robu<sup>1</sup>, Nupur K. Purohit<sup>1</sup>, Jyotika Rajawat<sup>1</sup>, Lucio Tentori<sup>2</sup> and Grazia Graziani<sup>2</sup>

<sup>1</sup> Laboratory for Skin Cancer Research, CHU-Q (CHUL) Research Centre, Laval University, Quebec City, QC, Canada

<sup>2</sup> Department of System Medicine, University of Rome "Tor Vergata", Rome, Italy

\*Correspondence: girish.shah@crchul.ulaval.ca

#### Edited by:

Christina Annunziata, National Cancer Institute, USA

#### Reviewed by:

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The pharmacological inhibitors of poly(ADP-ribose) polymerase-1 (PARP-1) have reached the first milestone toward their inclusion in the arsenal of anti-cancer drugs by showing consistent benefits in clinical trials against BRCA-mutant cancers that are deficient in the homologous recombination repair (HRR) of DNA double strand breaks (DSB) (1, 2). PARP inhibitors (PARPi) also potentiate therapeutic efficacy of ionizing radiation and some chemotherapeutic agents (1). These effects of PARPi were initially linked to inhibition of the role of PARP-1 in base excision repair (BER) of DNA damaged by endogenous or exogenous agents, resulting in accumulation of single strand breaks (SSB), which upon conversion to toxic DSB lesions would kill cancer cells deficient in DSB repair (1, 3, 4). However, PARPi lethality in HRR-deficient cancers can also be explained by other mechanisms not involving a direct effect of PARPi on BER [reviewed in Ref. (5, 6)]. In addition, therapeutic benefits of PARPi with agents such as carboplatin in HRR-proficient and -deficient tumors [reviewed in Ref. (1, 7)], simply cannot be explained by BER inhibitory effect of PARPi. Therefore, PARPi are like magic bullets that can kill cancer cells under different circumstances, but to comprehend their global scope and limitations, here we discuss the full range of their targets and the possible impact of broad specificity of current PARPi during prolonged therapy of cancer patients.

#### MECHANISMS OF ACTION OF PARPI IN CANCER THERAPY: MAGIC BULLETS BUT MOVING TARGETS

It is not surprising that the mechanism of action of PARPi in killing cancer cells still remains an open question, because its principal target PARP-1 is a multifunctional protein implicated in various cellular responses to DNA damage ranging from different pathways of DNA repair and cell death to stress signaling, transcription, and genomic stability (8, 9), all of which could be affected by PARPi and thus influence outcome of cancer therapies. Following are various possibly overlapping mechanisms for the anti-cancer effect of PARPi.

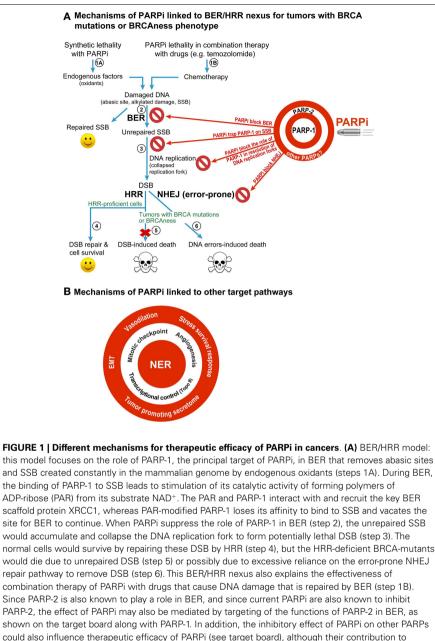
#### BER/HRR NEXUS FOR SYNTHETIC LETHALITY OF PARPI IN BRCA-MUTANT CANCERS

It was first demonstrated by two teams (3, 4) that two individually non-lethal conditions, i.e., PARPi-mediated inhibition of PARP-1 and BRCA mutationinduced HRR deficiency in cancer cell, would become synthetic lethal when combined in a single cell [reviewed in Ref. (1, 5, 10, 11)] (Figure 1A). This model focuses on the role of PARP-1 in BER, the pathway that repairs abasic sites and SSB that are constantly created in the mammalian genome by endogenous oxidants. When PARPi suppress the role of PARP-1 in BER, the unrepaired SSB would accumulate and collapse the DNA replication fork to form potentially lethal DSB. The normal cells would survive by repairing these DSB by HRR, but the HRR-deficient BRCAmutants would die due to unrepaired DSB or possibly due to excessive reliance on the

error-prone non-homologous end-joining (NHEJ) repair pathway to remove DSB (Figure 1A). This model also covers minor variations of the central theme as reviewed recently (1, 10) (Figure 1A). For example, tumors with other conditions that cause HRR deficiency or "BRCAness" phenotype would also be susceptible to PARPi. It permits inclusion of PARP-2 and its role in BER as target of PARPi, because most current PARPi also inhibit PARP-2 (10). It also explains the potentiating effect of PARPi in the combination therapy with radiation or chemicals, such as temozolomide, irinotecan, or topotecan, because DNA damage caused by these agents is also repaired by BER.

#### ALTERNATIVE TARGETS OF PARPI IN BRCA-MUTANT CANCERS

However, the above mechanism is inadequate to explain all the effects of PARPi seen in BRCA-mutant cancers, which could be explained by the effect of PARPi on alternate targets, as reviewed earlier (5, 6, 10) (Figure 1A). In brief, (i) PARPi could be trapping PARP-1 or PARP-2 to SSB with resultant PARP-SSB complex that would be more toxic than unrepaired SSB or even knockdown of PARPs (5, 12). (ii) PARPi could act via upregulation of NHEJ pathway, which would presumably cause genomic instability and eventual lethality (13). (iii) PARPi could suppress the role of PARP-1 in reactivating DNA replication forks (5). Thus, apart from BER/HRR nexus, there could be NHEJ/HRR or DNA replication/HRR nexus to explain PARPi lethality in BRCA-mutant cancers.



BER/HRR mediated therapeutic effect of PARPi is not yet fully assessed. (**B**) Other targets of PARPi that can confer therapeutic benefits of PARPi PARPi could also be effective anticancer agents by targeting the role of PARP-1 in other DNA repair pathways, such as NER; or other cellular pathways, such as control over cell cycle, tumor angiogenesis, transcription, epithelial-mesenchymal transition (EMT), stress survival response, vasodilation, or tumor-promoting secretome.

#### EXPANDING UNIVERSE OF POTENTIAL TARGETS OF PARPi

Therapeutic effectiveness of PARPi seen with some drugs cannot be explained by any of the above models, e.g., the potentiating effects of PARPi on the platinum-based drugs such as carboplatin, cisplatin, or oxaliplatin on HRRdeficient or -proficient tumors [reviewed in Ref. (1, 7)] (**Figure 1B**). These observations were further supported by recent studies showing the potentiating effect of PARPi veliparib on carboplatin treatment of patients with BRCA-mutant breast cancers (14) or carboplatin and phosphoinositide 3-kinase mTOR inhibitor treatment of mouse xenografts of BRCA-competent triple negative breast cancer cells (15). Since platinum compounds cause DNA damage that is largely repaired by the nucleotide excision repair (NER) pathway and not BER, we need to think beyond BER for an explanation. Moreover, BER was shown to mediate toxicity of cisplatin by competing with the repair of cisplatin inter-strand cross-links and DSB caused by these links (16). Therefore, if PARPi effect was mainly via inhibition of BER, we should have observed less and not more toxicity of cisplatin.

One possible explanation is that PARPi could be causing vasodilation (Figure 1B) to improve intra-tumoral delivery of platinum drugs (1), although it needs to be confirmed if this generalized effect could also potentiate other drugs. On the other hand, recently discovered roles of PARP-1 in improving the efficiency of NER-mediated removal of UV-induced DNA damage (17-19) provides a more handy explanation for the PARPi-induced potentiation of platinum compound-based drugs, which also cause DNA damage that is repaired by NER (Figure 1B). This NER targeting effect of PARPi alone can account for death of HRR-proficient tumors, as seen in clinical trials [reviewed in Ref. (1, 7)] and supported by in vitro results showing that PARP-1 depletion (20) or inhibition (19) decreases clonogenic survival of UV-exposed human skin fibroblasts with no reported HRR-deficiencies. Of course, PARPi could have an additional effect in this model due to suppression of the role of PARP-1 in HRR pathway (21). In addition, in the PARPi-treated BRCA-mutant HRR-deficient tumors, the unrepaired DNA damage by platinum drugs could collapse the DNA replication fork to form DSB and cause lethality. Thus, the NER effect alone or NER-HRR nexus could be possible explanations for the lethality of PARPi/platinum compounds in HRR-proficient or -deficient tumors.

The clinical and preclinical studies have also revealed other targets of PARPi in cancer therapies that are linked to various roles of their multifunctional target PARP-1 in following cellular processes (**Figure 1B**). (i) Transcriptional control of drug-target genes: PARPi have been shown to increase toxicity of topoisomerase II-poison doxorubicin *in vitro* (22) or in xenografted tumors in mice (23). This effect could be due to doxorubicin-induced decrease in expression and activity of PARP-1 (24) or PARPi-mediated increase in expression of topoisomerase II, because the transcription activator Sp1 loses its affinity for the topoisomerase II-promoter region upon modification by polymer of ADP-ribose (PAR) created by the activated PARP-1 (22). (ii) Mitotic checkpoint: the beneficial effects of PARPi with microtubule stabilizing mitotic inhibitor paclitaxel in patients with recurrent metastatic gastric cancers with BRCAness phenotype (25) could be linked to suppression of the role of PARP-1 in maintaining the mitotic checkpoint via PARylation of itself or the mitotic checkpoint protein CHFR (26, 27). An abrogation of mitotic checkpoint would kill cancer cells, because they will be forced to divide before resolution of the damage. (iii) Tumor-promoting secretome: PARPimediated suppression of the role of PARP-1 in elaborating tumor-promoting secretome containing cytokines and growth factors has been suggested as a cause for decreasing the resistance to another mitotic inhibitor docetaxel (28). (iv) Angiogenesis: the role of PARP-1 in promoting angiogenesis that fuels the growth of tumors can also be target of PARPi, because PARP-1 depletion or PARPi reduce vessel formation (29) and expression of markers of angiogenesis in melanoma (30) or endothelial cells (31). (v) Epithelial-mesenchymal transition (EMT) and metastasis: PARPi or PARP-1 depletion-induced reduction in aggressiveness and growth of metastatic melanoma in animal studies (30, 31) along with decreased markers for EMT (31, 32) suggest that the increase in progressionfree survival of PARPi-treated patients could be due to reduction in the proliferation rate of the primary tumor and repression of its metastatic potential. (vi) Stress survival response: finally, cancer cells respond to any therapy by elaborating various stress responses to survive; and PARP-1 and its product PAR play key roles in these stress responses (9). Hence the suppression of pro-survival stress responses could explain the effectiveness of PARPi with any anti-cancer drug. An expanding list of potential targets of PARPi provides us with a much larger vision of the future applications of PARPi in cancer therapy.

#### BROAD SPECIFICITY OF PARPi: A KEY ISSUE FOR THE FUTURE OF PARPi THERAPY

There are two basic issues arising from the broad specificity of current PARPi.

- (a) PARPi can inhibit more than one PARP ("they are bazookas not bullets"): many of the current PARPi in clinical trials display strong binding to PARPs 1-4 (33), and inhibit both PARP-1 and 2 at clinically relevant concentrations (10). Most studies assume that the effect of PARPi on both PARP-1 and 2 is important for therapy; however, this may not be the case. In fact, some studies using specific knockdown of PARPs showed that only the knockdown of PARP-1, but not PARP-2, replicates: (i) the synthetic lethal effect of PARPi on BRCA2 mutant cells (3); (ii) potentiation of cisplatin by PARPi in BRCAproficient triple negative breast cancer cells (34); and (iii) sensitization of melanoma cells in vitro to temozolomide (35). On the other hand, the effect of PARPi on gemcitabine in the above breast cancer cells was replicated by PARP-2 knockdown and not PARP-1 knockdown (34). In contrast, the siRNA for PARP-1 could specifically prevent the growth of BRCA-deficient ovarian cancer cell-derived tumors in mice (36). Since the double knockout of PARP-1 and PARP-2 is embryonic lethal (37), we must verify the assumption that gratuitous inhibition of unrelated PARPs has no effect on the end-results.
- (b) Indiscriminate inhibition of all the roles of a given PARP by PARPi ("we are nuking the entire PARPlandscape"): PARP-1, the principal target of PARPi, is a multifunctional protein that is implicated not only in DNA repair but also in various forms of cell death, transcription, epigenetic control of gene expression, and chromatin remodeling (8, 38). Hence even if we were to develop novel PARPi to specifically inhibit only PARP-1, it will still shut down most if not all the functions of PARP-1. Similar arguments can be made for PARPimediated suppression of different roles

of PARP-2. Although adverse genomic consequences of PARPi therapy have not vet been reported, we need to consider that prolonged PARPi therapy may cause genome instability because PARP- $1^{-/-}$  mouse embryonic fibroblasts have a tendency to become tetraploid (39, 40), and the susceptibility of PARP-1<sup>-/-</sup> female mice to develop mammary carcinoma is enhanced if p53 is also mutated, a phenomenon frequently observed in cancers (41). In effect, PARPi are the magic bullets, but instead of doing precision targeting with them for the desired effect, we are simply nuking the entire spectrum of functions of that target PARP, which could result in unintended consequence during maintenance (prolonged) therapy with PARPi including survival of damaged cancer cells, development of secondary tumors as a consequence of genomic instability and resistance to PARPi. Thus, while the current broad specificity PARPi work properly for shortterm cancer therapy, there is a need for development of new and more specific PARPi that are unique not only for a given PARP but also for a given function of that PARP related to its anti-cancer effect.

It is heartening that PARPi have shown some clinical benefit for BRCAmutant cancer patients in clinical trials as monotherapy or as a combination therapy, but we need to do a lot more to understand the therapeutic effect of PARPi to establish them firmly in the arsenal of anti-tumor agents against variety of cancers.

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# REFERENCES

- Curtin NJ, Szabo C. Therapeutic applications of PARP inhibitors: anticancer therapy and beyond. *Mol Aspects Med* (2013) 34(6):1217–56. doi:10. 1016/j.mam.2013.01.006
- Do K, Chen AP. Molecular pathways: targeting PARP in cancer treatment. *Clin Cancer Res* (2013) 19(5):977–84. doi:10.1158/1078-0432. CCR-12-0163
- Bryant HE, Schultz N, Thomas HD, Parker KM, Flower D, Lopez E, et al. Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature* (2005) 434(7035):913–7. doi:10.1038/nature03443
- Farmer H, McCabe N, Lord CJ, Tutt AN, Johnson DA, Richardson TB, et al. Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature* (2005) 434(7035):917–21. doi:10.1038/nature03445
- Helleday T. The underlying mechanism for the PARP and BRCA synthetic lethality: clearing up the misunderstandings. *Mol Oncol* (2011) 5(4):387–93. doi:10.1016/j.molonc.2011. 07.001
- De Lorenzo SB, Patel AG, Hurley RM, Kaufmann SH. The elephant and the blind men: making sense of PARP inhibitors in homologous recombination deficient tumor cells. *Front Oncol* (2013) 3:228. doi:10.3389/fonc.2013.00228
- Tentori L, Muzi A, Dorio AS, Dolci S, Campolo F, Vernole P, et al. MSH3 expression does not influence the sensitivity of colon cancer HCT116 cell line to oxaliplatin and poly(ADP-ribose) polymerase (PARP) inhibitor as monotherapy or in combination. *Cancer Chemother Pharmacol* (2013) **72**(1):117–25. doi:10.1007/s00280-013-2175-0
- Gibson BA, Kraus WL. New insights into the molecular and cellular functions of poly(ADPribose) and PARPs. *Nat Rev Mol Cell Biol* (2012) 13(7):411–24. doi:10.1038/nrm3376
- 9. Luo X, Kraus WL. On PAR with PARP: cellular stress signaling through poly(ADP-ribose) and PARP-1. *Genes Dev* (2012) **26**(5):417–32. doi:10. 1101/gad.183509.111
- Montoni A, Robu M, Pouliot E, Shah GM. Resistance to PARP-inhibitors in cancer therapy. Front Pharmacol (2013) 4:18. doi:10.3389/fphar.2013. 00018
- Lord CJ, Ashworth A. The DNA damage response and cancer therapy. *Nature* (2012) 481(7381):287–94. doi:10.1038/nature10760
- Murai J, Huang SY, Das BB, Renaud A, Zhang Y, Doroshow JH, et al. Trapping of PARP1 and PARP2 by clinical PARP inhibitors. *Cancer Res* (2012) **72**(21):5588–99. doi:10.1158/0008-5472. CAN-12-2753
- Patel AG, Sarkaria JN, Kaufmann SH. Nonhomologous end joining drives poly(ADP-ribose) polymerase (PARP) inhibitor lethality in homologous recombination-deficient cells. *Proc Natl Acad Sci* U S A (2011) 108(8):3406–11. doi:10.1073/pnas. 1013715108
- 14. Somlo G, Frankel PH, Luu TH, Ma C, Arun B, Garcia A, et al. Efficacy of the combination of ABT-888 (veliparib) and carboplatin in patients with BRCAassociated breast cancer. *J Clin Oncol* (2013) **31**(15-Suppl):1024.

- 15. Dey N, Sun Y, Carlson J, Friedman L, De P, Leyland-Jones B. A combination of dual PI3K-mTOR inhibitor, GDC-0980, with PARP inhibitor plus carboplatin blocked tumor growth of BRCA-competent triple-negative breast cancer cells. J Clin Oncol (2013) **31**(15-Suppl): 2613.
- Kothandapani A, Dangeti VS, Brown AR, Banze LA, Wang XH, Sobol RW, et al. Novel role of base excision repair in mediating cisplatin cytotoxicity. *J Biol Chem* (2011) 286(16):14564–74. doi:10.1074/jbc.M111.225375
- Robu M, Shah RG, Petitclerc N, Brind'Amour J, Kandan-Kulangara F, Shah GM. Role of poly(ADPribose) polymerase-1 in the removal of UVinduced DNA lesions by nucleotide excision repair. *Proc Natl Acad Sci U S A* (2013) 110(5):1658–63. doi:10.1073/pnas.1209507110
- Luijsterburg MS, Lindh M, Acs K, Vrouwe MG, Pines A, van Attikum H, et al. DDB2 promotes chromatin decondensation at UV-induced DNA damage. J Cell Biol (2012) 197(2):267–81. doi:10. 1083/jcb.201106074
- Pines A, Vrouwe MG, Marteijn JA, Typas D, Luijsterburg MS, Cansoy M, et al. PARP1 promotes nucleotide excision repair through DDB2 stabilization and recruitment of ALC1. *J Cell Biol* (2012) 199(2):235–49. doi:10.1083/jcb.201112132
- Ghodgaonkar MM, Zacal NJ, Kassam SN, Rainbow AJ, Shah GM. Depletion of poly(ADP-ribose)polymerase-1 reduces host cell reactivation for UV-treated adenovirus in human dermal fibroblasts. DNA Repair (Amst) (2008) 7:617–32. doi:10.1016/j.dnarep.2008.01.001
- Helleday T, Bryant HE, Schultz N. Poly(ADPribose) polymerase (PARP-1) in homologous recombination and as a target for cancer therapy. *Cell Cycle* (2005) 4(9):1176–8. doi:10.4161/cc.4.9. 2031
- Magan N, Isaacs RJ, Stowell KM. Treatment with the PARP-inhibitor PJ34 causes enhanced doxorubicin-mediated cell death in HeLa cells. *Anticancer Drugs* (2012) 23(6):627–37. doi:10. 1097/CAD.0b013e328350900f
- Mason KA, Valdecanas D, Hunter NR, Milas L. INO-1001, a novel inhibitor of poly(ADP-ribose) polymerase, enhances tumor response to doxorubicin. *Invest New Drugs* (2008) 26(1):1–5. doi:10. 1007/s10637-007-9072-5
- 24. Zaremba T, Ketzer P, Cole M, Coulthard S, Plummer ER, Curtin NJ. Poly(ADP-ribose) polymerase-1 polymorphisms, expression and activity in selected human tumour cell lines. *Br J Cancer* (2009) **101**(2):256–62. doi:10.1038/sj.bjc. 6605166
- 25. Bang Y-J, Im S-A, Lee K-W, Cho JY, Song E-K, Lee KH, et al. Olaparib plus paclitaxel in patients with recurrent or metastatic gastric cancer: a randomized, double-blind phase II study. *J Clin Oncol* (2013) **31**(15-Suppl):4013.
- 26. Ahel I, Ahel D, Matsusaka T, Clark AJ, Pines J, Boulton SJ, et al. Poly(ADP-ribose)-binding zinc finger motifs in DNA repair/checkpoint proteins. *Nature* (2008) 451(7174):81–5. doi:10.1038/ nature06420
- 27. Kashima L, Idogawa M, Mita H, Shitashige M, Yamada T, Ogi K, et al. CHFR protein regulates mitotic checkpoint by targeting PARP-1 protein

for ubiquitination and degradation. *J Biol Chem* (2012) **287**(16):12975–84. doi:10.1074/jbc.M111. 321828

- Zhao S, Coleman I, Coleman R, Nelson P. Association of PARP inhibitors and docetaxel resistance through suppressing a tumor microenvironment-associated secretory program. *J Clin Oncol* (2013) 31(15-Suppl):e22212.
- 29. Tentori L, Lacal PM, Muzi A, Dorio AS, Leonetti C, Scarsella M, et al. Poly(ADP-ribose) polymerase (PARP) inhibition or PARP-1 gene deletion reduces angiogenesis. *Eur J Cancer* (2007) 43(14):2124–33. doi:10.1016/j.ejca.2007. 07.010
- Tentori L, Muzi A, Dorio AS, Bultrini S, Mazzon E, Lacal PM, et al. Stable depletion of poly (ADP-ribose) polymerase-1 reduces in vivo melanoma growth and increases chemosensitivity. *Eur J Cancer* (2008) 44(9):1302–14. doi:10.1016/j.ejca.2008. 03.019
- Rodríguez MI, Peralta-Leal A, O'Valle F, Rodriguez-Vargas JM, Gonzalez-Flores A, Majuelos-Melguizo J, et al. PARP-1 regulates metastatic melanoma through modulation of vimentin-induced malignant transformation. *PLoS Genet* (2013) 9(6):e1003531. doi:10.1371/journal.pgen.1003531
- McPhee TR, McDonald PC, Oloumi A, Dedhar S. Integrin-linked kinase regulates E-cadherin expression through PARP-1. *Dev Dyn* (2008) 237(10):2737–47. doi:10.1002/dvdy.21685
- 33. Wahlberg E, Karlberg T, Kouznetsova E, Markova N, Macchiarulo A, Thorsell AG, et al. Family-wide chemical profiling and structural analysis of PARP and tankyrase inhibitors. *Nat Biotechnol* (2012) 30(3):283–8. doi:10.1038/nbt.2121
- 34. Hastak K, Alli E, Ford JM. Synergistic chemosensitivity of triple-negative breast cancer cell lines to poly(ADP-Ribose) polymerase inhibition, gemcitabine, and cisplatin. *Cancer Res* (2010) **70**(20):7970–80. doi:10.1158/0008-5472. CAN-09-4521
- 35. Tentori L, Muzi A, Dorio AS, Scarsella M, Leonetti C, Shah GM, et al. Pharmacological inhibition of poly(ADP-ribose) polymerase (PARP) activity in PARP-1 silenced tumour cells increases chemosensitivity to temozolomide and to a N3-adenine selective methylating agent. *Curr Cancer Drug Targets* (2010) **10**(4):368–83. doi:10.2174/156800910791208571
- Goldberg MS, Xing D, Ren Y, Orsulic S, Bhatia SN, Sharp PA. Nanoparticle-mediated delivery of siRNA targeting Parp1 extends survival of mice bearing tumors derived from Brca1-deficient ovarian cancer cells. *Proc Natl Acad Sci U S A* (2011) 108(2):745–50. doi:10.1073/pnas. 1016538108
- Menissier de Murcia J, Ricoul M, Tartier L, Niedergang C, Huber A, Dantzer F, et al. Functional interaction between PARP-1 and PARP-2 in chromosome stability and embryonic development in mouse. *EMBO J* (2003) 22(9):2255–63. doi:10. 1093/emboj/cdg206
- 38. Zampieri M, Guastafierro T, Calabrese R, Ciccarone F, Bacalini MG, Reale A, et al. ADP-ribose polymers localized on Ctcf-Parp1-Dnmt1 complex prevent methylation of Ctcf target sites. *Biochem J* (2012) 441(2):645–52. doi:10.1042/BJ20111417

- 39. Simbulan-Rosenthal CM, Haddad BR, Rosenthal DS, Weaver Z, Coleman A, Luo R, et al. Chromosomal aberrations in PARP(-/-) mice: genome stabilization in immortalized cells by reintroduction of poly(ADP-ribose) polymerase cDNA. *Proc Natl Acad Sci U S A* (1999) **96**(23):13191–6. doi:10.1073/pnas.96.23. 13191
- Halappanavar S, Shah GM. Defective control of mitotic and post-mitotic checkpoints in Poly(ADP-ribose) polymerase-1(-/-) fibroblasts after mitotic spindle disruption. *Cell Cycle* (2004) 3(3):335–42. doi:10.4161/cc.3.3.670
- 41. Tong WM, Yang YG, Cao WH, Galendo D, Frappart L, Shen Y, et al. Poly(ADP-ribose) polymerase-1 plays a role in suppressing mammary tumourigenesis in mice. *Oncogene* (2006) 26(26):3857–67. doi:10.1038/sj.onc.1210156

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