



Management of andrological disorders from childhood and adolescence to transition age: guidelines from the Italian Society of Andrology and Sexual Medicine (SIAMS) in collaboration with the Italian Society for Pediatric Endocrinology and Diabetology (SIEDP) – part-2

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Abstract

Purpose Andrological pathologies in adulthood often originate from conditions arising during childhood, adolescence, or even as early as the gestational and neonatal periods. Despite their clinical relevance, pediatric andrological disorders remain underrepresented in the literature and no shared position statements or comprehensive guidelines are currently available. The present paper complements a previous publication by the same societies, entitled “*Management of andrological disorders from infancy to adolescence and transitional age: Part-1 – The SIAMS–SIEDP position statement*”, by further expanding on these topics.

Methods SIAMS, in collaboration with SIEDP, convened a multidisciplinary expert task force to develop updated guidelines on the diagnosis and management of andrological disorders from childhood through adolescence and into the transitional age. The resulting recommendations were formulated using the Grading of Recommendations, Assessment, Development, and Evaluation (GRADE) system.

Results In this *Part-2* of the guidelines, a comprehensive literature review was conducted, focusing on English-language articles related to “cryptorchidism,” “micropenis,” “hypospadias,” “testicular tumors,” “epididymitis,” “orchitis,” “differences/disorders of sexual development,” and “testicular torsion”. For each condition, three key aspects were analyzed: diagnosis, clinical management, and treatment. Based on this analysis, specific recommendations and suggestions were developed for each disorder.

Conclusions The multidisciplinary guidelines presented in this paper complement those published in the previous Part-1 document. Developed through collaboration among leading medical societies in the field, this joint effort led to a consensus on a set of practical recommendations and suggestions aimed at supporting healthcare professionals in optimizing both andrological and overall health during the transitional age.

Keywords Cryptorchidism · Micropenis · DSD · Testicular Tumors · Hypospadias · Testicular torsion · Orchiepididymitis

Introduction

Many of the andrological conditions encountered in adult patients—ranging from genital, hormonal, reproductive, and oncological issues to sexual and psychological aspects—originate during childhood, adolescence, or even

as early as the gestational and neonatal periods. For this reason, andrology should not be regarded solely as a discipline of adulthood, but rather as a medical specialty concerned with male health and reproductive function across the entire lifespan, starting from birth—or even from fetal life—and extending into aging.

It is well established that the male gonads, along with hypothalamic gonadotropin releasing hormone (GnRH)-secreting neurons and pituitary gonadotroph cells, are highly sensitive to a wide range of influences—including genetic, environmental, chemical, and physical factors—that may disrupt their proper development and function during critical windows such as gestation, the neonatal period, puberty, and the transitional age [1–3]. In this context, andrological evaluation by a pediatric endocrinologist or an andrologist is recommended throughout the prepubertal, pubertal, and postpubertal stages, particularly when indicated by a neonatologist or primary care pediatrician during routine screening visits. Careful assessment in neonates, children, and adolescents enables early diagnosis of a wide spectrum of andrological disorders, including disorders of sexual development, structural anomalies of the genital tract, normal and abnormal pubertal development, undescended testes, genital tumors, and disturbances of gonadal function, with implications for growth, virilization, and fertility. When not promptly diagnosed and treated, these conditions may result in long-term adverse effects on male reproductive health.

Despite their clinical relevance, the current literature on pediatric and adolescent andrological disorders remains limited and fragmented, often focusing on isolated conditions and overlooking the critical transitional period [4–12]. Moreover, effective collaboration between pediatric endocrinologists and andrologists remains essential to ensure appropriate continuity of care during the transition age. This period, following the completion of pubertal development up to young adulthood, involves the transfer of patients from pediatric to adult healthcare services and requires careful attention to provide continuous care and appropriate management of age-specific health issues. The involvement of andrologists and adult endocrinologist may occur at different ages (even early during childhood) according to organizational setting, which may vary across Countries.

While *Part-1* of these guidelines focused on outlining the general framework and highlighted key issues related to andrological care during development [13], the present *Part-2* expands this work by providing evidence-based recommendations on other specific andrological conditions. This joint effort by the Italian Society of Andrology and Sexual Medicine (SIAMS) and the Italian Society of Pediatric Endocrinology and Diabetology (SIEDP) aims to offer practical guidance for the diagnosis, management, and treatment of a wide range of disorders affecting the male population from childhood through the transitional age. Pediatric care systems at both the primary and secondary levels vary widely across Europe [14, 15]. Therefore, these guidelines, developed by physicians practicing in Italy, are primarily tailored to the Italian context, although they include general

recommendations that may also be applicable to other countries and can be adapted to specific local circumstances.

Materials and methods

A literature search of articles in English for the term “cryptorchidism”, “vanishing testis”, testicular agenesis”, “micropenis”, “hypospadias”, “testicular tumors”, “epididymitis”, “orchitis”, “differences/disorders of sexual development” and “testicular torsion” has been performed. In this guideline, we will provide recommendations regarding the evaluation and management of different andrological disorders during childhood and transitional age based on the GRADE (Grading of Recommendations, Assessment, Development, and Evaluation) system for grading both the quality of evidence and the strength of recommendations [16]. According to this system, the strength of recommendation will be divided into ‘strong’, indicated by the number 1 and ‘we recommend’, and ‘weak’ indicated by the number 2 and ‘we suggest’. The grading of the quality of evidence is denoted as follows: $\oplus\circ\circ\circ$ for very low-quality evidence; $\oplus\oplus\circ\circ$ for low quality; $\oplus\oplus\oplus\circ$ for moderate quality; and $\oplus\oplus\oplus\oplus$ for high quality [16].

Minipuberty

This paragraph aims to recall some key concepts related to the critical phase of minipuberty, which are further addressed in the subsequent subsections of these guidelines. Minipuberty is a physiological phase that occurs in the first few months of life, during which the hypothalamic-pituitary-gonadal (HPG) axis, active during fetal development but suppressed at birth, reactivates for a brief period. This leads to measurable increases in gonadotropins, such as luteinizing hormone (LH) and follicle-stimulating hormone (FSH), as well as gonadal steroids like testosterone (T) in male infants and estradiol (E2) in female infants [17–19]. Minipuberty typically begins in the first few weeks of life, peaking between 4 and 12 weeks of age. In males, the T surge gradually diminishes by 4 to 6 months, whereas in females, E2 circulating levels may persist for a longer duration. During this period in males, LH stimulates Leydig cells in the testes to produce T peaking in the 2nd–3rd month of life when T values may reach those of fertile adult men. T is crucial for penile and scrotal growth, testicular descent, and early germ cell maturation. FSH stimulates Sertoli cells, increasing inhibin B and promoting spermatogonial development and testicular growth.

In females, FSH levels are higher than in males during minipuberty, stimulating early ovarian follicular activity. Ovarian granulosa cell products inhibin B and anti

Mullerian hormone (AMH) transiently increase at the time of the postnatal gonadotrophin surge. Unlike T levels in boys that show a clear peak at 1–3 months of age and then decrease steadily towards the age of 6 months, E2 levels in girls fluctuate, probably reflecting cyclic maturation and atrophy of ovarian follicles. E2 levels decrease during the 2nd year of life. Early postnatal E2 secretion contributes to the development of the uterus and breast tissue, although its clinical manifestations remain subtle.

Minipuberty is an essential diagnostic window for evaluating the functional integrity of the HPG axis in conditions like Congenital Hypogonadotropic Hypogonadism (CHH) and Disorders of Sex Development (DSDs) [18, 20, 21]. Hormonal testing during this phase, such as measuring serum LH, FSH, T, Inhibin B, and AMH, provides valuable insights into gonadal function without the need for stimulation tests. In male infants with cryptorchidism or micropenis, evaluating minipuberty can help in the detection of underlying HPG axis abnormalities and guide early interventions. In cases of CHH, the absence of the gonadotropin and sex steroid surge is a hallmark feature [18]. Disorders of androgen receptor sensitivity or enzymatic defects in T synthesis may also manifest during this phase (normal/elevated testosterone levels associated with relatively elevated gonadotropins in androgen receptor defects, reduced T levels with variable gonadotropins in enzymatic defects of gonadal steroidogenesis).

In cases where androgen deficiency is detected during minipuberty, early treatment with low-dose T may be considered to support normal genital development [17–19]. There are also more limited reports suggesting the use of gonadotropin therapy to replicate the missed minipuberty [20, 22]. Nonetheless, it should be emphasized that both mentioned therapeutic approaches are currently considered off label. For female infants, hormonal abnormalities during this time could signal potential reproductive challenges, warranting close follow-up [18].

Cryptorchidism

Definition and epidemiology

Cryptorchidism (or undescended testis) is defined as failure of one or both testicles to complete their descent into the lower part of the scrotum (Table 1) [23]; it is the most common genital disorder identified at birth [24, 25].

Prevalence of cryptorchidism at birth has been estimated between 1.8 and 8.4% among at term and appropriate for gestational age (AGA) babies [23]. Given the relatively late migration of testes through the inguinal canal into the scrotum, prevalence in preterm and/or small for gestational age (SGA) newborns may reach 45%, as inversely related to gestational age [24]. Prevalence at the age of 3 months and 1 year drops to 0.9–1.6% and 1.0–1.5%, respectively, due to the spontaneous testicular descent in around 40% of infants, usually within the first 3 months of age [23–26]. Congenital cryptorchidism is generally diagnosed at birth, but a few children with cryptorchidism may be referred late, either because of missed diagnosis or because they have an ascending testicle (acquired cryptorchidism) [24]. Acquired cryptorchidism is the condition in which one or both testicles cannot be returned into the scrotum after previously having been localized in a stable scrotal position (Table 1) [27]. Its prevalence is 0.6%–7% between 18 and 36 months of age and 1.1%–2.2% between 6 and 13 years, with a peak around 8 years of age [24, 28, 29].

Cryptorchidism should be differentiated from congenital anorchia or vanishing testis syndrome (Table 1), defined as the absence of testicular tissue in 46,XY phenotypic males [30]. The condition is unilateral in 97% of cases and accounts for about 10% of cases in which the testis is absent from the scrotum or inguinal canal [31]. Müllerian structures are absent and Wolffian structures are normal, but the ipsilateral vas deferens is often rudimentary and the epididymis is absent [32].

The frequency of bilateral anorchia is reported to be 1 in 20,000 phenotypic males [33]. Since testicular function is essential to stimulate the development of the external genitalia, testes must have been normal in the first trimester of

Table 1 Cryptorchidism: definitions

Cryptorchidism (undescended testis)	Failure of one or both testicles to complete their descent into the lower part of the scrotum; the testicles are located along the normal testicular descent pathway (intra-abdominal, inguinal, supra-scrotal or high scrotal position)
Congenital anorchia (vanishing testis syndrome)	The absence of testicular tissue in 46,XY phenotypic males
Acquired cryptorchidism (ascending testis)	One or both testicles cannot be returned into the scrotum after previously having been localized in a stable scrotal position
Ectopic testis	Testicle outside its normal route, in a perineal, femoral, or pubo-penile position, or at a crossed scrotal position (unilateral location of both testes).
Retractile testis (hypermobile testes)	Testis located in the upper scrotum or lower inguinal canal that can be moved by manual reduction into the scrotum, where it remains until stimulation by the cremasteric reflex occurs

gestation. Antenatal or perinatal vascular thrombosis or testicular torsion occurring during later testicular descent into the scrotum have been suggested [34, 35].

Boys with bilateral anorchia are distinguished from those with bilateral abdominal cryptorchidism by imaging (ultrasound and/or magnetic resonance), laparoscopy or surgical exploration, low serum levels of AMH and inhibin B and undetectable T levels after human chorionic gonadotropic (hCG) administration.

Rarely, the testis may be found outside its normal route in a perineal, femoral, or pubo-penile position (ectopic testis), or at a crossed scrotal position (unilateral location of both testes) [36].

Undescended testis should be always distinguished from retractile one, defined as a testicle located in the upper scrotum or lower inguinal canal that can be moved by manual reduction into the scrotum, where it remains until stimulation by the cremasteric reflex occurs (Table 1) [27].

Pathophysiology

Cryptorchidism is usually an isolated condition of unknown cause, though it can also occur in genetic syndromes (syndromic cryptorchidism). The presence of micropenis and/or hypospadias may indicate impaired secretion or action of hormones critical for genital development and testicular descent [37, 38]. Testicular descent occurs in two phases. The first, transabdominal phase takes place between 8 and 15 weeks of gestation [39]. During this phase, the testis is anchored in the internal inguinal ring by enlargement of the gubernaculum, preventing upward movement as the embryo grows [40]. Regression of the cranial suspensory ligament, which is androgen-dependent, also contributes to gonadal positioning [41].

Animal studies suggest that gubernaculum development is driven by INSL3 and its receptor [41, 42]. However, mutations in these genes are rarely observed in humans [43], reflecting the infrequent disruption of the transabdominal phase [44–46].

The second, inguinoscrotal phase involves the testis migrating from the internal inguinal area to the scrotum [39]. Widening of the inguinal canal, along with gubernaculum shrinkage and intra-abdominal pressure during the seventh month of gestation, guides the testis into the scrotum [35, 47].

This phase is androgen-dependent and may be impaired in androgen receptor defects [48–50] or inborn error of steroidogenesis [23], although causative mutations are rarely identified in isolated cases [51–53].

Although familial clustering has been reported, cryptorchidism is likely polygenic, with no single causative gene identified [54, 55].

Other mechanisms of under-masculinization may also affect testicular descent. In persistent Müllerian duct syndrome, defects of AMH or its receptor can result in abdominal or inguinal testes, confirming AMH's role in the transabdominal phase [56].

Congenital hypogonadotropic hypogonadism is another common association [57, 58]. Here, hCG can partially compensate for missing LH in utero, explaining why not all affected boys are cryptorchid at birth, while postnatal pituitary gonadotropins may influence testicular position, as seen in secondary (acquired) cryptorchidism [59].

Clinical picture

A testicle is defined as undescended if it cannot be manipulated, tension-free, into the scrotum at any age. Palpable undescended testes lie below the internal inguinal ring and may be detected along the normal descent pathway in inguinal, supra-scrotal, or high scrotal positions. By contrast, a retractile testis can be manually brought into the scrotum without tension, though it may return to its previous position afterward [60].

About 70% of undescended testes are palpable with experienced examination [61, 62]. The remaining 30% are non-palpable and may reside in inguinal, abdominal, or ectopic locations, while a small fraction cannot be located even after laparoscopic exploration, representing vanishing testes [60].

Diagnostic evaluation

Recommendations and Suggestions

R1.1 We recommend always assessing testicular position at birth in all newborns and at 3 and 6 months of age in those with congenital cryptorchidism, for the known possibility of spontaneous descent within the first 6 months of life (1, ⊕⊕⊕⊕).

R1.2 We recommend that primary care pediatricians and/or general practitioner should palpate testes for position at each recommended well-child visit (1, ⊕⊕⊕⊕).

R1.3 We recommend a prompt referral to a tertiary care center for all male newborns with bilateral non-palpable testes, especially when associated with other hallmarks of disorders of sexual development (i.e. hypospadias, microphallus, bifid scrotum) (1, ⊕⊕⊕⊕).

Evidence

The diagnostic work-up of isolated monolateral cryptorchidism is still predominantly clinical. A thorough physical

exam should be sufficient to diagnose undescended testes [63].

In the hands of an experienced clinician, more than 70% of cryptorchid testes are palpable by physical examination. In the remaining 30% of cases the gonads are non-palpable and the challenge is to confirm absence or presence of the gonads and their characteristics [64].

Acquired cryptorchidism is easily distinguished from congenital cryptorchidism if scrotal testicular position has been documented since birth. Retractable testes have been described as more at risk for secondary ascent [65, 66]. Acquired cryptorchidism is more common in boys with a history of proximal hypospadias suggesting a common mechanism related to aberrant androgen signaling [65]. Acquired cryptorchidism may also represent a complication of inguinal hernia surgical correction [67].

Ultrasound shows limited sensitivity and specificity in localizing non-palpable testes, with 45% sensitivity and 78% specificity [68]. Intra-abdominal testes have been localized at surgical exploration in 49% of boys with a non-palpable testicle and a negative ultrasound [68]. The use of computed tomography is limited due to cost and radiation exposure. Magnetic resonance imaging demonstrates better sensitivity and specificity, but its use is discouraged due to the high cost, limited availability, and the requirement for anesthesia [69, 70].

Unilateral or bilateral cryptorchidism associated with micropenis, suggest congenital hypogonadotropic hypogonadism, isolated or in the context of a multiple pituitary hormone defect [71].

Unilateral or bilateral cryptorchidism associated with hypospadias and/or bifid scrotum are suggestive of DSD [72]. A newborn with male phallus or hypospadias, atypical external genitalia and bilateral non-palpable gonads is potentially a genetic female (46,XX) with congenital adrenal hyperplasia due to 21-hydroxylase deficiency or another rare defect of adrenal and gonadal steroidogenesis (11 beta-hydroxylase deficiency, steroidogenic acute regulatory protein (StAR) or cholesterol side-chain cleavage enzyme defect). These newborns are at risk for life threatening salt wasting crises during the first weeks of life and need urgent hormonal and genetic assessment (at least karyotype analysis), followed by further genetic analyses to define the specific steroidogenesis defect and its treatment [73].

The assessment of karyotype and gonadal function is strongly recommended in the definition of complex cases, especially when cryptorchidism is bilateral and/or part of a complex DSD [74, 75].

Infant boys with 46,XY karyotype, bilateral non-palpable testes, normal penile development or micropenis without hypospadias need extensive evaluation to differentiate

vanishing testis syndrome (bilateral congenital anorchia) from bilateral abdominal testes [76].

Remarks

Clinical examination should be always performed with the child placed in a supine and frog-legged position on the examination bed, or on the parent's lap [61, 62]. The clinician should examine the baby in a warm room with warm hands, standing to the patient's right side. The left hand is initially placed lateral to the deep inguinal ring and moves along the inguinal canal up to the pubic tubercle, exerting a moderate pressure to push the testicle down. This maneuver overcomes the cremasteric reflex, which naturally pulls the testicle away from the scrotum. When the left hand has reached the pubic tubercle, the right hand can locate the testis and gently pull it down to the base of the scrotum. The same process can be repeated to locate the testis on the opposite side [60].

Imaging studies do not significantly help diagnosis and rarely assist in decision-making. Diagnostic laparoscopy (or open surgical exploration) should be performed on all non-palpable unilateral and most bilateral cryptorchid patients [77].

Corrected age should be always used to determine optimal surgical timing in infants born preterm (<32 gestational weeks) (*corrected age = chronological age (weeks) minus the number of weeks the infant was born before the due date*).

A basal blood sampling (for LH, FSH, testosterone, AMH, inhibin B) performed between 4 and 16 weeks of life (minipuberty) can assess gonadal function and help the diagnosis of hypogonadism or DSD, with greater accuracy than stimulation tests performed later in childhood. During minipuberty, infants with anorchia obviously do not have any postnatal surge of T, AMH and inhibin B are undetectable, while FSH and LH are elevated [18, 71]. On the contrary, isolated congenital hypogonadotropic hypogonadism is suggested in cryptorchid infants with micropenis and low or undetectable levels of gonadotrophins and T [18, 71, 78, 79].

After minipuberty, the hypothalamic-pituitary-gonadal axis remains quiescent until true puberty [18, 71, 78, 79]. In the past decades, stimulation tests with hCG were recommended to assess Leydig cell function in children with bilateral non-palpable testes outside minipuberty [80]. Nevertheless, the failure of T to increase after hCG stimulation is not diagnostic of anorchia, because undescended dysgenetic testes may not respond to hCG stimulation [81]. On the contrary, undetectable levels of inhibin B and AMH associated with elevated gonadotropin levels are suggestive

of isolated anorchia in a phenotypic 46,XY male with bilateral non-palpable testes [74, 81].

Therapeutic management

Recommendations and suggestions

R1.4 Children with persistently undescended testis after 6 months of age should be referred to surgical evaluation, the best surgical timing being between 6 and 12 months of age to improve potential for fertility and reduce the risk for testicular cancer (1, ⊕⊕⊕○).

R1.5 We recommend annual follow-up until puberty to monitor for a possible secondary ascent in boys with retractile testes, avoiding any medical treatment (1, ⊕⊕⊕○).

R1.6 We suggest considering hormonal therapy only in infants with congenital hypogonadotropic hypogonadism to induce minipuberty, improve surgical outcome, and sometimes induce testicular descent (2, ⊕○○○).

R1.7 Individuals with a history of cryptorchidism and delayed surgical correction (>12–18 months) should be informed by clinicians about potential long-term risk of infertility (in bilateral cryptorchidism) and testicular cancer to ensure long-term andrological follow-up (clinical principle). (Expert opinion).

Evidence

Orchidopexy is the only successful therapy to relocate the testis into the scrotum [77, 82].

Spontaneous descent of the testes into the scrotum during the first months of life highlights the need to repeat clinical examination at 3 and 6 months of age before considering surgical correction [23–26, 83].

Patient age at orchiopexy is directly related to the severity of germ cell and Leydig cell depletion [84, 85]. Non-palpable testes carry the highest risk of germ cell depletion [86]. Adverse histologic features (e.g. loss of germ cells) are similar in congenital cryptorchidism and acquired cryptorchidism [87]. Early surgery improves testicular volume, but the benefits on future fertility are still uncertain [37]. Late complications of surgery include testicular atrophy (up to 15% for intra-abdominal testis) and testicular re-ascent requiring revision surgery (10% for intra-abdominal testis) [88].

Early data suggested a significantly higher risk of carcinoma in abdominal testes [89]. The relative risk of developing testicular cancer in individuals with isolated cryptorchidism has been estimated to be 2.90, even after early surgical correction [90]. Nonetheless, it is also known that this risk is generally lower in congenital forms of hypogonadism (such as congenital hypogonadotropic hypogonadism) [91].

Hormonal therapy with hCG or LHRH in cryptorchidism is controversial and evidence is lacking on any clear benefit in improving fertility [77, 92, 93]. Available data on the efficacy of hormonal therapy to induce testicular descent (FSH+hCG/LH, GnRH) have demonstrated an inadequate efficacy and a high risk of recurrence [37, 94, 95]. Around 60% of men with congenital central hypogonadism (lacking minipuberty) achieve adult testicular volume and semen quality with gonadotropin replacement therapy [96, 97]. A proposed approach to enhance the fertility potential of these patients is to mimic the hormonal milieu of minipuberty, with the aim of promoting Sertoli and germ cell proliferation and initial differentiation [18, 71, 96].

Short-term combined gonadotrophin therapy stimulates normal levels of T, inhibin B and AMH in infants with congenital hypogonadotropic hypogonadism [71, 98, 99]. In these children, gonadotrophin treatment might also facilitate the surgical approach to undescended testes and contribute to reduce surgical complications by the increase of testicular volume [18, 99].

Remarks

Hormonal therapy of cryptorchidism should be considered only in cryptorchid infants with congenital hypogonadotropic hypogonadism, with the aim of mimicking minipuberty, even if there is not yet consensus to use this approach as a routine therapy.

Short-term gonadotropin therapy in these infants aims to improve future testicular development and fertility potential, but it also facilitates surgical approach and outcome. A practical approach with daily injections of recombinant LH and rFSH, has been proposed. Treatment should be started before age 6 to 9 months, at an initial dose of 37.5 IU rLH and 75 IU rFSH per day. This dose can be increased 1 or 2 weeks later, if serum LH concentrations do not reach the physiological range [18]. Data on efficacy and safety of with hCG area also available [100, 101].

Successful scrotal repositioning of the testis may reduce, but not completely prevent, long-term issues of infertility and testicular cancer [102].

A long-term andrological follow-up addressing fertility issues and long-term risk of testicular cancer is appropriate for patients with a history of cryptorchidism, especially if surgical correction was performed late and/or cryptorchidism was bilateral. Diagnostic work-up should include periodical clinical evaluation, hormonal assessment, and imaging studies [103–105].

Disorders of sex development (DSD)

The aim of this chapter is to provide a general approach to a child with a DSD. Due to the high heterogeneity of etiologies of DSDs together with the rapid evolution of knowledge, management and even terminology in this field, we suggest referring also to available international guidelines or position statements for more insights into diagnostic workup and treatment of these conditions [106–115].

Epidemiology

The term “DSD” defines a large and heterogeneous group of congenital conditions in which development of chromosomal, gonadal, or anatomic sex is atypical.

This nomenclature and the subsequent classification of distinct DSDs was introduced in 2005, following the Chicago consensus conference [116]. Nomenclature remains controversial and now other terms are also used such as “differences” or “variations of sex development”; these terms may help to introduce the concept of the range of variation that may occur in sex development, as underlined by some advocacy groups and some patients [106, 107]. The current medical classification is largely based on the genetic status of the patient (Table 2) [106, 116].

The birth prevalence of genital atypicity may be as high as 1 in 300 births, but the birth prevalence of a condition

that may lead to difficulties in gender attribution and require expert examination may be as low as 1 in 5000 births [117]. The age at clinical presentation of a patient with a DSD is mostly at birth, but can be diagnosed also in infancy, late adolescence or even adulthood [110, 118]. Sometimes a DSD can be already suspected in the prenatal age [108].

Pathophysiology

The term “sex development” refers to the biological processes leading to the morphological and functional acquisition of sexual characteristics. It is driven by complex, dose- and time-dependent genetic networks that diverge between the sexes from the 7th week of embryonic development and culminate at puberty with reproductive maturity [119, 120].

Dimorphic sexual development involves two sequential processes [121]:

1. “sexual determination” - the differentiation of the undifferentiated gonad into a testis or ovary according to chromosomal sex.
2. “sexual differentiation” - the development of internal and external genitalia and the sex-specific changes occurring at puberty, guided by gonadal hormones.

Table 2 Current classification of disorders of sex development (DSD) (from [116])

Sex chromosomal DSD	46,XY DSD	46,XX DSD
45,X Turner syndrome and variants	<i>Disorders of gonadal Development</i> • Complete or partial gonadal dysgenesis, monogenic forms (for example, SRY, NR5A1 and WT1) • Testes regression • Ovotesticular DSD • Syndromic forms	<i>Disorders of gonadal Development</i> • (Ovo)testicular DSD • Monogenic forms of primary ovarian insufficiency (mutations in genes involved in gonadal (ovarian) development; for example NR5A1 and WT1) • Syndromic forms
47,XXY Klinefelter syndrome and variants	<i>Disorders of androgen synthesis</i> • Associated solely with androgen biosynthesis defects (mutations or deficiencies in HSD17B3 and SRD5A2, for example) • Associated with congenital adrenal hyperplasia and early androgen biosynthesis defects (mutations and/or deficiencies in STAR, CYP11A1, HSD3B2, POR and CYP17A1) • Associated with placental insufficiency or endocrine disruption • Syndromic forms (for example, Smith–Lemli–Opitz)	<i>Disorders of androgen excess</i> • Aromatase (CYP19A1) deficiency • Congenital adrenal hyperplasia (mutations and/or deficiencies in CYP21A2, HSD3B2, CYP11B1 and POR) • Luteoma • Latrogenic
45,X/46,XY Mixed gonadal dysgenesis	<i>Disorders of androgen Action</i> • Complete and partial androgen insensitivity	<i>Unclassified disorders</i> • MRKH type I and II syndrome • Complex syndromic disorders
46,XX/46,XY Chimerism	<i>Persistent Müllerian duct syndrome</i> • Due to mutations or deficiencies in <i>AMH</i> and <i>AMHR2</i> <i>Unclassified disorders</i> • Hypospadias of unknown origin • Epispadias • Complex syndromic disorders	

Gonads originate from the genital ridges, which appear undifferentiated in both sexes around the 4th week of gestation. Sex determination is orchestrated by gene networks, including SRY on the Y chromosome, which activates SOX9 to promote testicular development [122]. Altered timing or dosage of these genes can impair testis or ovary formation [123]. Internal genitalia arise from two parallel ducts: the Wolff (mesonephric) and Müller (paramesonephric) ducts. In the developing testis, testosterone production begins around the 7th–8th week and acts via androgen receptors to form the epididymis, vas deferens, and seminal vesicles. Sertoli cells produce anti-Müllerian hormone (AMH) from the 7th week onward, inducing regression of Müllerian ducts [124]. In the absence of testosterone, Wolff ducts regress; without AMH, Müllerian ducts persist, forming female internal genitalia. External genitalia develop from the cloaca. In males, the genital tubercle forms the penis, the urogenital folds fuse to create the penile urethra, and the genital swellings merge into the scrotum. Testicular descent, partially driven by Leydig cell–derived INSL3, occurs in the last trimester. Dihydrotestosterone (DHT), produced from testosterone by 5 α -reductase, directs development of the penis, scrotum, and prostate [125]. While largely androgen-dependent, external genital development also involves other hormones and hormone-independent mechanisms; in the absence of DHT, female-typical external genitalia develop.

Clinical picture

Several signs characterize the clinical picture of DSDs and are all included among the following criteria used for raising the clinical suspicion of these conditions according to patient's age (Table 3).

Diagnostic evaluation

Recommendations and suggestions

R2.1 We recommend a careful and thorough evaluation of the genitals in all newborns at birth aiming to identify atypical genitalia. (1, ⊕⊕⊕⊕).

R2.2 We recommend that a newborn/child/adolescent with a suspected DSD is referred quickly to a Pediatric Endocrinology tertiary center with a multidisciplinary team experienced in DSDs for a complete care and a comprehensive diagnostic work up (1, ⊕⊕⊕⊕).

R2.3 We recommend performing genetic testing in all DSD patients (1, ⊕⊕⊕⊕).

R2.4 We recommend that the family and/or the patient should be promptly offered psychosocial support by a professional with experience in DSD. (1, ⊕⊕⊕⊕).

R2.5 We recommend that the diagnosis as well as pros and cons of the various intervention options should be communicated to the parents and the patient in a plain and structured communication strategy by the multidisciplinary team. (1, ⊕⊕⊕⊕).

R2.6 We suggest the early involvement of support groups. (2, ⊕⊕⊕⊕).

Evidence

When DSD is suspected at birth or at any age diagnostic procedures should be performed by a multidisciplinary team at a designated excellence center with efficient communication between all its professional areas. An excellence center should have members of all involved areas of expertise inside (pediatric/adult endocrinologists, psychologists, ethicists, pediatric surgeons/urologists, gynecologists, andrologist, geneticists, pathologists, radiologists, laboratorist, sex therapists, and social workers) or alternatively foster a close connection to those disciplines not available at the center itself [106, 110, 112]. Representatives of the different disciplines and specialties should be experienced in the field of DSDs. The center should ensure DSD patient care

Table 3 Age-specific clinical features suggestive of Disorders of Sex Development (DSD)

Newborn	Child and young adult
1. Overt atypical genitalia	1. Previously unrecognized atypical genitalia
2. Apparent female genitalia with an enlarged clitoris, posterior labial fusion, or an inguinal/labial mass	2. Inguinal hernia in a female
3. Apparent male genitalia with bilateral undescended testes	3. Delayed or incomplete puberty
4. Micropenis	4. Virilization in a female
5. Isolated perineal hypospadias	5. Primary amenorrhea, with or without breast development
6. Mild hypospadias with undescended testis	6. Breast development in a male
7. A family history of DSD	7. Gross and occasionally cyclic hematuria in a male
8. A discordance between genital appearance and a prenatal karyotype	8. Infertility

throughout his (or her) life and ensure the effective transition of care from childhood through adolescence to adulthood [126, 127].

A comprehensive diagnostic evaluation is required to determine the clinical outcome regardless of an immediate indication for treatment [106, 110]. History should be aimed at investigating if there are other family members with atypical genital development or if any unexplained deaths have occurred in the family. Probing for consanguinity may be helpful especially when an autosomal recessive disorder is being considered in the differential diagnosis. A detailed family history regarding fertility, as well as the use of androgenic or estrogenic substances during pregnancy, maternal virilization during pregnancy, and information about procreation may be helpful.

A detailed physical exam is mandatory. In newborn and infants, description of external genitalia is facilitated using specifically designed, quantitative scoring systems. A non-binary version applicable in both boys and girls, has been validated in a European multicentre study (External Genital Score, EGS; Table 4) [128]. The anogenital distance (AGD) correlates with prenatal androgen exposure [129].

Hormonal diagnostics are essential [130]. Basic values to be determined include adrenal (17-hydroxyprogesterone, cortisol, androstenedione, serum electrolytes) and gonadal function tests (E2, T, DHT, gonadotropins, AMH). The analysis of these hormones should be performed at a specialized laboratory. Currently, chromatographic, mass spectrometric methods are recommended for exact steroid hormone measurements [106, 130, 131]. However, these methods are not yet widely available. Measurement of AMH plays a pivotal role in determining the type and quality of gonadal tissue [132]. In undervirilized 46,XY individuals, low AMH levels suggest gonadal dysgenesis, while normal or high levels indicate androgen insensitivity or synthesis defects. In 46,XX individuals, elevated AMH levels may point to the presence of testicular tissue, as seen in ovotesticular DSD.

Gonadal tests may be repeated during minipuberty in the suspicion of hypogonadism [133]. Additional investigations include hCG and adrenocorticotropin (ACTH) stimulation tests [110]. A complete examination by ultrasound of the

pelvic structures as well as the perineal region and of the entire urinary tract is recommended. Magnetic resonance imaging should be reserved for cases when ultrasonography is not informative, when there are abnormalities of the urinary tract, in case of intra-abdominal gonads. For identification of intra-abdominal gonads, laparoscopy may refine the diagnosis. Advantages and risks should be considered before performing a laparoscopy for identification of the gonads and inner genitalia. X-ray based fluoroscopic genitography can be unnecessary. An invasive procedure such as urethrocystoscopy or an inspection of the vagina under anesthesia might be necessary to identify details of the anatomy [134, 135].

Chromosome analysis (including karyotype with fluorescence in-situ hybridization, FISH and array-comparative genomic hybridization, aCGH) is the first step of genetic analyses [136, 137]. The next step includes the use of next-generation sequencing (NGS) assays, designed to sequence multiple DSD genes on a targeted panel in one analysis or whole-exome sequencing (WES) with predetermined filters that target DSD genes [138, 139]. Written consent is mandatory before conducting any procedure. A genetic consultation should outline the possibilities, limits, and problems of such tests.

The psychosocial counseling for parents and patients should be provided for the whole life since it is crucial to promote a positive adaptation to the clinical condition [140–142]. Family counselling to parents and/or patient is a key step in this process and should be integrated at any medical visit [143]. Any aspect of the management should be communicated in a balanced and respectful way by providing risks and benefits of any future choice and avoiding time pressure. Counseling and attendance of children/adolescents with DSD is based on the principle of allowing them to accept their bodies and achieve the best quality of life possible. The child should be informed gradually and repeatedly while it is growing up. Communication must be documented, and a copy of the information should be handed out to the parents. The inclusion of trained peer support in the multidisciplinary DSD team is advisable to the supportive management of patients with DSDs [144, 145].

Table 4 External Genitalia Score (EGS) [128]

EGS	Labioscrotal fusion	Genital tubercle length (mm)	Urethral meatus	Right gonad	Left gonad
3	Fused	> 31	Top of the GT		
2.5		26–30	Coronal glandular		
2			Along the GT		
1.5	Posterior fusion	21–25	At the GT base	Labioscrotal	Labioscrotal
1		10–20	Labioscrotal	Inguino-Labioscrotal	Inguino-Labioscrotal
0.5				Inguinal	Inguinal
0	Unfused	< 10	Perineal	Impalpable	Impalpable

This inclusion will strengthen the acceptance of DSD and facilitate the sharing of experiences, thereby reducing the stress and isolation felt by patients and their families.

Remarks

The management of a newborn, child, or adolescent with a DSD can be complex and often presents a clinical challenge [106, 146, 147]. Initial interactions and communications can have a lasting impact on the patient's medical journey. Therefore, it is crucial that all healthcare professionals who may encounter such cases possess a thorough understanding of DSDs. Equally important is the management of these cases in specialized reference centers staffed by experienced and competent personnel [106]. When immediate referral is not feasible—for instance, in premature infants requiring neonatal intensive care—it is essential for the attending neonatologist or healthcare provider to promptly consult with specialists at a reference center to coordinate care.

A primary concern in the initial evaluation is the identification or exclusion of adrenal insufficiency, as it represents the only clinical scenario necessitating immediate, life-saving hormone replacement therapy [147]. Congenital adrenal hyperplasia (CAH) due to 21-hydroxylase deficiency (21-OHD) is the most prevalent cause of 46, XX DSD. Diagnosis involves measuring serum 17-hydroxyprogesterone levels, performing karyotyping, and conducting ultrasound examinations by experienced operators. Prompt diagnosis allows for the initiation of critical hormone replacement therapy.

However, 21-OHD CAH is not the sole cause of adrenal insufficiency associated with DSD. Other aetiologies include 11 β -hydroxylase deficiency, 3 β -hydroxysteroid dehydrogenase deficiency, and P450 oxidoreductase deficiency, which can present in both 46, XX and 46, XY individuals. In 46, XY patients, conditions such as StAR defects, P450scc, and P450c17 deficiencies can lead to DSD presentations [148].

Assessing gonadal function at birth is important, though commonly used laboratory techniques like immunometric assays have limitations, especially in neonates. The adoption of chromatography coupled with mass spectrometry is expected to improve diagnostic accuracy. Re-evaluation during the “minipuberty” period is recommended.

Genetic testing has advanced significantly and is increasingly becoming a first-line diagnostic tool in certain clinical scenarios. Early and accurate genetic diagnosis facilitates targeted management strategies.

In Italy, birth registration, including the assignment of name and sex, must occur within 10 days. In cases of neonates with atypical genitalia, this timeframe may be insufficient to gather comprehensive clinical information for accurate sex assignment. While many aspects of long-term outcomes in

individuals with DSD remain under investigation, current data support specific recommendations for sex assignment. For example, in individuals with a 46,XY karyotype and 5 α -reductase deficiency or 17 β -hydroxysteroid dehydrogenase deficiency diagnosed at birth, current guidelines support assigning sex in accordance with chromosomal and gonadal sex, as is also recommended for individuals with 46,XX CAH. A medically justified and documented delay in birth registration is generally accepted.

Psychological support is an integral component of comprehensive DSD management and should be provided by professionals with expertise in the field. Studies highlight the importance of accessible mental health services throughout the lifespan of individuals with DSD. Multidisciplinary teams should include mental health professionals to address the psychosocial needs of patients and their families.

Therapeutic management

Recommendations and suggestions

R2.7 We recommend that DSD counseling/therapy should be an interdisciplinary task, performed in centers of excellence aiming to achieve the best quality of life and acceptance of individual anatomy. (1, ⊕⊕⊕⊕).

R2.8 We recommend starting immediately replacement therapy when adrenal insufficiency is diagnosed (1, ⊕⊕⊕⊕).

R2.9 We recommend starting sex hormone replacement therapy in case of documented deficiency at pubertal age or later at the time of diagnosis independently from the underlying cause. (1, ⊕⊕⊕⊕).

R2.10 We suggest considering GnRH-analogue to block the HPG axis when the subjects start to enter puberty and develops a phenotype that is discordant with the existing gender role, to be started only after in-depth psychological assessment and support. (2, ⊕○○○).

R2.11 We suggest that surgical interventions at early age is indicated only when there is a documented risk for the patient's health and well-being in case of no intervention. (2, ⊕⊕○○).

R2.12 We recommend that, if surgical procedures are planned, they should be performed by qualified surgeons with proven experience in this field. (1, ⊕⊕⊕○).

R2.13 We suggest that a long-term follow up should be offered to people who have undergone surgery. (2, ⊕⊕○○).

R2.14 We suggest a continuous follow-up and screening of both function and oncological risk of the gonads (2, ⊕⊕○○).

R2.15 We suggest that transitional care should be provided by a multidisciplinary team which should ideally

include both pediatric and adult professionals with experience in DSD management. (2, ⊕○○○).

Evidence

The therapeutic management of a patient with DSD should be carried out by multidisciplinary teams in centers of excellence. The aim is of achieving the best quality of life and acceptance of individual anatomy and the optimal long-term well-being [106, 107, 110, 118]. However, the therapeutic management is extremely challenging, due to the diagnostic and ethical issues presented by many patients and the difficulty in providing a precise prognosis for any individual patient. These issues must be kept in mind in any decision-making process and in communication with the patients and families. Families and patients, when possible, fully informed, must be directly involved in decision-making process [149].

Adrenal insufficiency is a possible and potentially lethal cause of atypical genitalia. Congenital adrenal hyperplasia (CAH) is the most common cause of 46,XX DSD. In all situations of adrenal insufficiency, hormone replacement therapy is mandatory and must be promptly started [150].

Sexual hormone treatment during infancy and childhood must be deemed to be critical, as hormone therapy induces irreversible effects. In patients with micropenis, T therapy may be proposed (see the micropene chapter). In patients with 5-alpha reductase deficiency, local DHT therapy has been shown to be effective in increasing the size of the penile shaft.

If the patient's gonads do not produce enough or indeed any hormones or after a gonadectomy has been performed, sexual hormone treatment become necessary [151] at the appropriate time, as already detailed in the Part 1 of these guidelines [13]. Hormonal induction of puberty is usually performed according to the current practices used for adolescents with pituitary or gonadal deficiency [13, 152]. Clinical studies of hormonal induction of puberty in patients with DSDs have not been conducted so far. The kind of hormone treatment provided should respect the patient's request. The choice of timing and hormone treatment should be made individually (e.g. in case of uncertainty regarding gender identity). (Patho)physiological aspects must be considered when choosing a hormonal substitution. Refusal of hormone substitution by a patient needs to be discussed, as the procedure is regarded as critical in preventing adverse events. An increase in hormone production during puberty can lead to a change in the child's phenotype. This might differ from the gender role in which the child/adolescent has lived up to this point. To gain time to think about their continuing gender identity, further development can be stopped by offering GnRH-analogue. In these cases, reference can

be made to the guidelines of gender incongruent persons [153–157].

The role of surgery in the management of patients with DSDs is changing, with the advocacy that any surgical intervention in neonates and infants that leads to irreversible changes should be done with the utmost caution. Ideally, surgical interventions should construct cosmetically appealing and well-functioning genitalia and preserve fertility in the best possible way. Although great progress has been made in this direction, often this goal is still difficult to achieve. The few publications on long-term outcomes are all retrospective reports and deal with small numbers of patients in whom surgery was often performed more than 20 years ago [158–160]. Surgical procedures have become much more precise and aim at preserving important anatomical and functional structures, although this does not guarantee better results because long-term data of modern procedures will only be available in 10–15 years. Surgical efforts to 'cure' patients with DSDs have been detrimental to many individuals in the past. Many individuals with a DSD have complained forcefully about the surgical procedures used. A moratorium on genital surgery in infancy has been proposed by advocacy groups. Currently, the prevailing thought (and legislative recommendations in some countries) claims that specific surgical procedures should be postponed until the individuals can make their own decision about the type of surgical intervention. Only in cases of vital harm and a strictly somatic functional or vital medical indication surgical interventions may be performed on minors [161, 162]. However, some parents of children with CAH, as well as several patients with CAH, do not agree with this rigorous recommendation. Studies confirm that most CAH patients identify themselves as female and do not regret, if they underwent feminization surgery during early childhood [163, 164]. If and when the surgery is decided, it should be performed by an experienced surgeon [165–167]. The surgeon has a particular responsibility to provide realistic and honest information about the possible outcomes of the surgery and the available options (not only surgical). The parents and/or the patient should be informed about post-operative care and possible complications and should be offered regular follow-up. The various forms of XY and X/XY DSD have a variable risk of developing a gonadal germ cell cancer. The risk is much higher in men and women with gonadal dysgenesis, specially forms that arise from early gonadal differentiation defects, such as mutations in SRY and WT1, than in individuals with ovotesticular DSD or with hormone synthesis or action disorders (for example, androgen insensitivity) [168–170]. Overall, the risk of tumorigenesis is also increased by the ectopic position of the gonads which may make the detection of malignancies of abdominal gonads more challenging

during early childhood by ultrasound or magnetic resonance imaging. In the absence of reliable tumor markers or imaging technologies for early detection of precursor lesions, it seems prudent to consider gonadal biopsies to exclude the presence of germ cell neoplasia in situ or gonadoblastoma in most individuals with 46,XY or 45,X/46,XY DSD with retained gonads. Given that the age of distribution for testicular germ cell cancers is well established, such biopsies are best performed in late adolescence [171–173]. Because a patient's own gonads might have a residual function, and because the removal of the gonads is an irreversible intervention affecting the patient's personal development, it must be considered very carefully, if and when this must be done. If individuals with high tumor risk retain their gonads into adulthood, regular morphological and functional screening must be guaranteed. If gonadectomy is recommended, the possibility of cryoconservation, if vital sperms are found, should be discussed with the patient [174, 175]. In the last few decades, there has been an increasing awareness of the transition to adult-oriented health care in adolescents and young adults with a chronic illness. Few studies have been conducted in adolescents with DSDs during transition, and there is a need for guidelines to manage the transition of these patients. Patients frequently experience difficulties in accessing specialized medical care in adulthood, resulting in loss to follow-up affecting the patients' physical and psychological health as well as quality of life. Clinical features and long-term outcomes are highly variable in most DSD conditions. Challenges in the care of DSD patients in adulthood are optimization of fertility potential, hormone replacement therapy and sexuality. A good transition process requires close interaction of pediatric with adult endocrinologists with the involvement of urologists, andrologists or gynaecologists, and psychologists [126, 127, 176].

Remarks

The foremost objective in managing a newborn with DSD is to identify or exclude adrenal insufficiency, as it necessitates immediate, life-saving hormone replacement therapy. Caregivers, and eventually the patient, must be thoroughly educated on managing the condition and preventing or treating potential adrenal crises. In recent decades, approaches to the medical and surgical management of individuals with atypical genitalia have evolved significantly. In some countries, such interventions have been restricted or deferred until the individual can provide informed consent. This topic remains the subject of ongoing debate, with differing perspectives even among patients and advocacy groups. In confirmed cases of gonadal insufficiency, hormone replacement therapy aligned with the individual's gender identity should be initiated in accordance with established protocols

for hypogonadal patients. Special consideration must be given to gonads at increased risk for neoplastic transformation. In such cases, prophylactic gonadectomy is indicated, with the potential option of preserving unaffected gonadal tissue when feasible.

Micropenis

Epidemiology

Isolated micropenis is defined as a stretched penile length (SPL) 2.5 standard deviations below the mean for age group (Table 5) without the presence of any other penile anomalies such as hypospadias [177, 178]. Minor ethnic differences in SPL have been reported [179, 180], though these are less evident in infants and young boys, and the growing immigrant populations in some Western countries should be considered [181]. Micropenis is a different condition respect to buried, webbed, trapped or diminutive penis, although all of them are included into the term “inconspicuous” [182, 183]. The incidence of micropenis is known to be 1.5 of 10,000 male newborns in United States [184]. According to the definition, approximately 0.8% of the male general population can carry this condition, although the isolated idiopathic or nonspecific 46,XY DSD form, which we will discuss in this section, constitutes about 25% of these forms [185].

Pathophysiology

The human penis develops from the genital tubercle (GT) visible by 5–6 weeks of gestation [186, 187]. From 8 weeks, maternal hCG stimulates testosterone production from fetal Leydig cells. Testosterone, converted to dihydrotestosterone (DHT), directs penile differentiation, a process largely complete by 12 weeks, when the GT forms the glans, genital folds form the penile shaft, and genital swellings merge to create the scrotum. During the second and third trimesters, penile growth is driven by fetal androgens, regulated by fetal pituitary gonadotropins, with the penis increasing almost 2 cm between weeks 16 and 38 [188]. Therefore, true micropenis reflects a hormonal defect occurring after 12 weeks of gestation.

Congenital micropenis may indicate a range of endocrine or syndromic conditions (Table 6) that require evaluation. After excluding hormonal deficiencies, a minority of patients remain undiagnosed and may be classified as carriers of idiopathic or nonspecific 46,XY DSD [189, 190].

Table 5 Stretched penile length [SPL cm; mean $\pm$ SD (mean-2.5 SD)] in normal males according to age for samples of North American, European and Asian Populations

Age	Asian population [377]	European population [378]	American population [379]
Newborns 30 wks*	-	-	2.5 $\pm$ 0.4 (1.5)*
Newborns 34 wks*	-	-	3.0 $\pm$ 0.4 (2.0)*
0–5 mos.	-	-	3.9 $\pm$ 0.8 (1.9)
6–12 mos.	-	-	4.3 $\pm$ 0.8 (2.3)
0–1	4.1 $\pm$ 0.4 (3.1)	3.5 $\pm$ 0.5 (2.4)	
1–2	4.2 $\pm$ 0.4 (3.2)	3.7 $\pm$ 0.5 (2.3)	4.7 $\pm$ 0.8 (2.6)
2–3	4.3 $\pm$ 0.4 (3.3)	3.9 $\pm$ 0.5 (2.9)	5.1 $\pm$ 0.9 (2.9)
3–4	4.4 $\pm$ 0.4 (3.4)	4.0 $\pm$ 0.6 (2.5)	5.5 $\pm$ 0.9 (3.3)
4–5	4.5 $\pm$ 0.5 (3.2)	4.3 $\pm$ 0.7 (2.5)	5.7 $\pm$ 0.9 (3.5)
5–6	4.6 $\pm$ 0.5 (3.3)	4.4 $\pm$ 0.6 (2.9)	6.0 $\pm$ 0.9 (3.8)
6–7	4.7 $\pm$ 0.6 (3.2)	4.5 $\pm$ 0.6 (3.0)	6.1 $\pm$ 0.9 (3.9)
7–8	4.8 $\pm$ 0.6 (3.3)	4.7 $\pm$ 0.7 (2.9)	6.2 $\pm$ 1.0 (3.7)
8–9	4.9 $\pm$ 0.7 (3.1)	4.7 $\pm$ 0.7 (2.9)	6.3 $\pm$ 1.0 (3.8)
9–10	5.1 $\pm$ 0.8 (3.1)	4.7 $\pm$ 0.7 (2.9)	6.3 $\pm$ 1.0 (3.8)
10–11	5.4 $\pm$ 0.8 (3.4)	4.8 $\pm$ 0.7 (3.0)	6.4 $\pm$ 0.1 (3.7)
11–12	5.7 $\pm$ 1.0 (3.2)	5.1 $\pm$ 0.9 (2.9)	-
12–13	6.2 $\pm$ 1.2 (3.2)	5.9 $\pm$ 1.4 (2.4)	-
13–14	6.8 $\pm$ 1.4 (3.8)	7.1 $\pm$ 1.6 (3.1)	-
14–15	7.6 $\pm$ 1.4 (4.1)	8.0 $\pm$ 1.3 (4.7)	-
15–16	8.2 $\pm$ 1.5 (4.4)	8.7 $\pm$ 1.2 (5.7)	-
16–17	8.8 $\pm$ 1.6 (4.8)	9.0 $\pm$ 1.1 (6.2)	-
17–18	9.5 $\pm$ 1.6 (5.5)	9.1 $\pm$ 1.1 (6.3)	-
18–19	10.2 $\pm$ 1.7 (5.9)	9.1 $\pm$ 1.1 (6.3)	-
19–20		9.5 $\pm$ 1.1 (6.7)	-
Adults			13.3 $\pm$ 1.6 (9.3)

* [380]

Table 6 Conditions associated with micropenis**Hypothalamo pituitary disorders (Hypogonadotropic hypogonadism):**

-Isolated

-Combined with other pituitary hormone deficiency

-Syndromic conditions: Prader-Willi syndrome; Bardet-Biedl syndrome; Laurence-Moon syndrome; Charge syndrome; Silver Russel syndrome; Rud syndrome

-Isolated Growth Hormone or IGF-1 deficiency

Gonadal disorders (Hypergonadotropic hypogonadism):

-Anorchia/Vanishing testes syndrome

-Disorders of gonadal development: Sex chromosome mosaicism; Partial gonadal dysgenesis

-Disorders of androgen synthesis: 3-beta-hydroxysteroid dehydrogenase deficiency; 17-beta-hydroxysteroid dehydrogenase deficiency; 17,20-lyase deficiency isolated or combined with 17-hydroxylase deficiency; 5-alfa-reductase deficiency

-Disorders of androgen action: Partial androgen insensitivity syndrome

-Chromosomal/genetic syndromes: Klinefelter (47XXY) and multiple X syndromes (48XXXY), Downs syndrome, Noonan syndrome, Robinow syndrome, CHARGE syndrome

-Penile agenesis (aphallia)

Miscellaneous:

-Maternal use of antifungals

-Environmental endocrine disruptors

-Nonspecific 46 XY DSD or idiopathic micropenis (i.e., cases with no evidence of endocrine or genetic abnormalities)

Clinical picture

Penile length is measured along the dorsum with the penis stretched, from the pubic symphysis to the glans tip, using a rigid ruler [178, 180, 189]. Penile girth should be recorded at both the base and coronal level [191]. Corpora cavernosa can usually be palpated, even if hypoplastic; the scrotum

is often small, and testes may be small or incompletely descended. Stancampiano et al. [189] reported the median/mean SPL values across childhood and adolescence for different ethnic backgrounds, and Table 5 shows mean-2.5 SD values for country-specific norms. According to international guidelines, micropenis is generally defined as <2 cm at birth and <4 cm after 5 years of age [190].

Diagnostic evaluation

Recommendations and suggestions

R3.1 We recommend that infants with congenital micropenis need to be considered for a complete clinical and endocrine evaluation by a specialized multidisciplinary team to exclude all hormonal or syndromic causes of the condition (1, ⊕⊕⊕⊕).

R3.2 We recommend that newborns with isolated micropenis, regardless of subsequent diagnostic investigations, carry out a hormonal work-up during the first 2 days of life or during the minipuberty window (3 to 6 months of life), assessing the basal pituitary-gonadal function (Testosterone, LH, FSH, Inhibin B and AMH serum levels) (1, ⊕⊕⊕⊕).

R3.3 We recommend that children/adolescents with isolated idiopathic micropenis have an accurate measurement of the stretched penile length, palpation of the corporeal bodies with measurement of girth/diameter, and evaluation for cryptorchidism (1, ⊕⊕⊕⊕).

R3.4 We recommend reviewing the patient with isolated idiopathic micropenis at least once in the prepubertal period and regularly from the time of expected puberty on to evaluate the development of sexual characteristics and possible supportive hormonal therapy (1, ⊕⊕⊕⊕).

R3.5 We suggest that in case of peri-pubertal presentation, the patient be examined in collaboration between pediatric and adult endocrinologists/andrologists, to familiarize the patient to the transition phase and long-term follow-up (2, ⊕⊕○○).

R3.6 We suggest that the family and/or the patient with micropenis should already be offered competent psychosocial attendance by a professional familiar with problems of the genito-urinary tract (2, ⊕⊕○○).

Evidence

Since isolated micropenis can be the marker of a wide range of endocrine alterations that include the hypothalamic-pituitary-gonadal axis or DSD [189], the evaluation of an expert multidisciplinary team is essential for diagnostic and therapeutic orientation (see the chapters on “Pubertal Delay” and “DSD”).

A sample collected during the first 2 days of life or during the minipuberty window (3 to 6 months of life), as above mentioned, is the unique temporal opportunity to highlight any physiological rise in gonadotropins and testosterone to evaluate the functioning of the pituitary-gonadal axis [71, 192, 193].

The penile length must be measured on a stretched penis by a trained expert since this is the measure that is strongly related to erected penis length [191]. The measurement of

penile circumference and diameter should be performed by a flexible centimeter and a caliper, respectively.

The transition process of boys with isolated idiopathic micropenis allows for the establishment of a long-term follow-up useful for evaluating the long-term outcomes of those who have or have not undergone therapies. It is unclear whether the presence of isolated idiopathic micropenis itself leads to long-term consequences apart from psychosocial distress in parents and young adults. In fact, studies investigating aspects of self-esteem, sexual function, and fertility in young adults with this condition are scarce and dated [194]. Psychosocial support should always be offered when available and an external consultant should be identified when it is not [195].

Remarks

When SPL is measured in the newborn during the first 12 h of life the length may be reduced by approximately 10% and should be reassessed [196]. The SPL inter- and intra-observer variation for trained personnel has been reported as 1 SD of 0.34 cm and 0.18 cm, respectively [197]. A standard karyotype does not allow to highlight possible hidden mosaicisms thus a higher number of metaphases (e.g., $n=100$) is always needed to improve accuracy [198]; karyotype may be preceded whenever possible by a fluorescence in situ hybridization (FISH) for the sex-determining region on the Y chromosome (SRY) for a faster exclusion of a DSD condition [199].

In case of micropenis belonging to a patient with a documented diagnosis of delayed puberty, an appropriate management should be settled, according to Part-1 of these guidelines [13].

Therapeutic management

Recommendations and suggestions

R3.7 We recommend that isolated idiopathic micropenis discovered early in life is treated by a pediatric endocrinology center (1, ⊕⊕○○).

R3.8 We suggest that patients with idiopathic micropenis and established androgen sensitivity, may be treated with testosterone/DHT therapy during the first 6 months of life or later in childhood/adolescence to stimulate the growth of the penis (2, ⊕⊕○○).

R3.9 We recommend against surgical therapy before adulthood, postponing this option in adult life (1, ⊕⊕○○).

Evidence

There is no consensus on the age, dosage, method of administration, or duration of T or DHT therapy in boys with isolated micropenis. A regimen of intramuscular depot-testosterone enanthate 25 mg administered monthly for 3 months has been reported to show an increase in penile length of over 100% in prepubertal boys with isolated micropenis. However, the study does not provide information on the long-term durability of the results achieved [200]. The same results with Te enanthate at the same dosage are reported also during minipuberty [22]. Arisaka et al. also documented a significant increase in SPL in 50 prepubertal boys treated with T cream 5% (10 mg/daily applied directly to the penis) for a duration of 30 days. The study documents the systemic absorption of transdermal T, although this does not appear to have significant negative effects on bone metabolism while Xu D et al. found that short term (6 months) and local application of DHT at low doses (0.1–0.3 mg/kg/day) in patients with micropenis could accelerate penile growth effectively without evident side effects [201, 202]; however, precautions still need be taken due to the paucity of long term data and the lack of ideal DHT dosage.

A recent randomized study on 49 patients compared the efficacy of transdermal DHT (5 mg/daily for 5 weeks with 1 or 2 repetitions if necessary) and T enanthate (50 mg i.m. monthly for 3 months, with 1 repetition if necessary) in treating isolated idiopathic micropenis in children and adolescents (mean age 9.7 ± 4.4 years). The study suggested a superiority of transdermal DHT compared to injectable exogenous T in the treatment of idiopathic micropenis, although both showed significant penis enlargement (mean penile size increment 2.82 vs. 1.87 cm, respectively) [203]. Based on the available evidence we thus suggest using T enanthate 25 mg i.m. in prepubertal children once a month for 3 months, repeatable only once after 3–6 months; or transdermal DHT (5 mg/daily for 6 weeks, repeatable 1–2 times). No evidence supports the use of hormonal therapies after puberty.

Remarks

Precautions still need be taken due to the paucity of long-term studies and the lack of ideal T and DHT dosage [189]. Moreover, T enanthate and DHT gel is off label for treatment during pediatric age (0–18 yrs.), at least in Italy, and an informed, signed consent need to be obtained by parents/legal representative of the minor. Furthermore, it should be considered that DHT is not available in all countries.

There is still uncertainty about the optimal age of therapy with androgens for maximal effect; given that tissue AR expression is high in early infancy, it would seem

appropriate to use androgens at that point, but it remains unclear whether such early use of androgens has any benefit on penile length in adulthood [204].

In addition to ongoing psychological support for the family and patient, the involvement of the adult endocrinologist/andrologist in the transitional period will help to explain the adolescent/young adult on the pro and cons of other future surgical options. Open doctor-patient communication regarding expectations, specific risks, benefits, and alternatives is of paramount importance in enabling the best possible results in this sensitive field [205].

During the transition process the adolescents/young adults must be counseled by expert personnel and advised with extreme caution about surgical options due to uncertain functional efficacy and the high rate of complications among patients with micropenis [190].

Hypospadias

Epidemiology

Hypospadias consists of an arrest in normal penile/urethral development with hypoplasia of the tissues forming the ventral side of the penis responsible of an ectopic meatus of the urethra. The earlier this process arrests, the more the form is proximal and severe [206]. Hypospadias is one of the most common congenital anomalies, although there is wide variation of prevalence according to countries and ethnicity (13.8 to 40/10,000), and there are conflicting data on the recent trends of prevalence [207, 208].

Pathophysiology

Male and female genital development diverges from the indifferent stage around 8 weeks' gestation under androgen influence. In normal males, the urethral groove fuses proximally to distally to form the tubular urethra, a process completed by ~17 weeks along with fusion of the ventral foreskin and straightening of the early penile curvature [209]. In hypospadias, disruption of urethral canalization or fusion results in an ectopic urethral meatus along the ventral shaft [206]. The pathogenesis is often unknown, but disrupted androgenic signaling, combined with genetic (Table 7) and environmental (Table 8) factors, is considered the main cause [210–215]. Proximal hypospadias, particularly when associated with cryptorchidism or micropenis, is more likely to have a genetic etiology. Commonly associated abnormalities include ventral penile curvature (chordee), hooded or incomplete prepuce, and hypoplastic corpora spongiosum [206].

Table 7 Genes potentially involved in hypospadias. Most of them are more frequently associated with other genital or multiorgan malformations [381]

Gene	MIM
Indifferent stage or early embryonic	
<i>WT1</i>	*607,102
<i>NR5A1 (SF1)</i>	*184,757
Early patterning	
<i>BMP4, BMP7</i>	*112,262, *112,267
<i>HOXA4, HOXB6, HOXA13</i>	*142,953, *142,961, *142,959
<i>FGF8, FGF10</i>	*600,483, *602,115
<i>FGFR2</i>	*176,943
<i>IRX5</i>	*606,195
<i>IRX6</i>	*606,196
<i>EYA1</i>	*601,653
<i>DGKK</i>	*300,837
<i>ZEB1</i>	*189,909
Masculinization	
<i>AR</i>	*313,700
<i>SRD5A2, SRD5A1</i>	*607,306, *184,753
<i>SRY</i>	*480,000
<i>SOX9</i>	*608,160
<i>FKBP4</i>	*600,611
<i>HSD3B2</i>	*613,890
<i>HSD17B3</i>	*605,573
Other	
<i>ESR1, ESR2</i>	*133,430, *601,663
<i>ATF3</i>	*603,148
<i>MAMLD1 (CXORF6)</i>	*300,120
<i>MID1</i>	*300,552
<i>INSL3</i>	*146,738
<i>BNC2</i>	*608,669

Clinical picture

Evaluation of hypospadias includes family history, detailed genital examination, and assessment of other congenital anomalies [216]. Examination should record SPL, penile curvature, foreskin development, and testicular position (Fig. 1). Preoperative classification is typically based on urethral meatus location: anterior (glandular/subcoronal), middle (distal/proximal/midshaft), and posterior (penoscrotal, scrotal, perineal) [217]. A more precise characterization also considers foreskin appearance and degree of curvature (Table 9) [206].

Quantitative scoring systems facilitate objective description of external genitalia in neonates and infants. A non-binary version of the External Masculinization Score (EMS) [117], the European External Genital Score (EGS), has been validated in multicenter studies (Table 4) [128]. The anogenital distance (AGD) correlates with prenatal androgen exposure [218], with males showing a higher AGDI/u ratio (0.49 ± 0.1) than females (0.39 ± 0.1), and EGS correlates with AGDI/u in males [128]. Such standardized metrics

improve clinical assessment, surgical planning, and multi-center research.

Diagnostic evaluation

Recommendations and suggestions

R4.1 We recommend evaluating hypospadias by detailed genital examination and classifying its severity by the physical findings reported in Table 3 (1, ⊕⊕⊕○).

R4.2 We suggest evaluating the commonly used anthropometric indicators such as external genitalia score (EGS), anogenital distance (AGD) and penoscrotal distance (PSD) in neonates and children carrying hypospadias (2, ⊕⊕○○).

R4.3 We recommend that in proximal and complex hypospadias, further diagnostic evaluation is performed, such as ultrasonography of the urinary tract and internal genital organs to detect other genito-urinary malformations (1, ⊕⊕⊕⊕).

R4.4 We recommend genetic testing in patients with scrotal or perineal hypospadias, especially if associated with cryptorchidism and micropenis because they are at high risk of Differences of Sex Development (DSD) (1, ⊕⊕⊕○).

Evidence

Several preoperative classification systems have been proposed to assess hypospadias severity based on position of the urethral meatus opening [218]. However, they lack precision and do not reliably consider the whole penile anatomical disruptions. From this aspect, the classification proposed by Baskin L, which we report in Table 3, considers the main anatomical aspects involved in the severity of hypospadias and offers relevant suggestions for the surgical or non-surgical management of this condition [206].

The current refined classification of hypospadias relies mainly on the quality of the urethral plate, the position of the urethral meatus, glans size and the degree of curvature. However, this can only be determined during surgery and there is still significant subjectivity between evaluators. If the postoperative classification can be described and predicted preoperatively based on local anatomical data, it will be of great significance for surgeons to design a proper surgical plan and preoperatively communicate with family members. He et al. has defined the anatomical abnormalities of hypospadias before puberty using current commonly used anthropometric index data to predict postoperative diagnostic classification. They found that anogenital distance (AGD) and penoscrotal distance (PSD) have a favorable predictive value for midshaft and proximal hypospadias [219]. Over 20% of boys with hypospadias are also known to have other extra-genital congenital conditions [220, 221].

Table 8 Environmental factors potentially interfering with male genitals formation.(Modified from [214])

Factors frequently investigated	Factors not frequently investigated
<i>Factors consistently associated with hypospadias</i>	
LBW/SGA	
Placental insufficiency	
Maternal Hypertension	
Pre-eclampsia	
Maternal intrauterine DES exposure	
<i>Factors associated with hypospadias in most studies</i>	
ICS	<i>Factors that seem to be associated with hypospadias</i>
Prolonged TTP	Paternal subfertility
High maternal BMI	Absence of nausea and vomiting in early pregnancy
Primiparity	Bleeding during pregnancy
Multiple pregnancy	Complications during labour
Pre-existing maternal diabetes	Maternal use of antihypertensive
Maternal use of antiepileptics	
<i>Factors showing inconsistent results</i>	
Preterm	Early age at menarche
Iron supplementation	Maternal thyroid disease
Maternal age	Fever in first trimester
Maternal vegetarian diet	Progestin/progestogens for threatened abortion
Maternal fish consumption	Paternal heavy metal exposure
Maternal and paternal exposure to pesticides	Living in rural or urban areas
Maternal occupational exposure to endocrine disruptors, heavy metals, phthalates	
Maternal serum levels of PCB's	
Seasonal trend	

In these cases, endocrinological evaluation is advised to exclude DSD (see DSD chapter), especially in case of concomitant unilateral or bilateral undescended testis [218].

Remarks

Severe hypospadias can masquerade as atypical genitalia warranting an evaluation for a disorder of sex development (see DSD chapter). The incidence of DSD is significantly increased in proximal or complex hypospadias and further assessment needs to be done to individuate patients with these conditions (pelvic ultrasound, karyotype, hormonal assessment and serum electrolytes) [222]. In cases where there are multiple major birth defects, further evaluation to detect an underlying syndrome or genetic defect must be performed [223, 224].

To evaluate the AGDs and the PSD measures, the TIDES method places the infant in a supine position with the lower half of the body exposed and the legs lifted in a froglike posture (with a 60–90° angle from the torso at the hip) and knees pulled back towards to shoulders [225]. AGDI (AGDlower), is measured from the center of the anus to the base of the labioscrotal border; AGDu (AGDupper), is measured from the center of the anus to the anterior base of the genital tubercle [128]. PSD measurement is based on skin fold boundaries to reflect the distance from the front, back,

left and right (12, 9, 3 o'clock) of the root of penis to the border of scrotum [219].

Therapeutic management

Recommendations and suggestions

R4.5 We recommend urologic referral and repair only for patients with hypospadias who are at-risk for voiding and/or sexual dysfunction (1, ⊕⊕○○).

R4.6 We recommend against performing circumcision in the newborn period for patients with asymmetric foreskin due to abnormal development on the ventral aspect of the penis (1, ⊕⊕⊕○).

R4.7 We suggest that hypospadias correction should preferably be performed between 6 and 18 months of age in full-term, healthy infants (2, ⊕⊕⊕○).

R4.8 We suggest tubularized incised plate urethroplasty (TIPU) or dorsal inlay graft urethroplasty (DIGU) as options for repairing of standard hypospadias, although, at present, no technique has demonstrated to be better (2, ⊕⊕○○).

R4.9 We suggest that a two-stage hypospadias repair should be used for the most severe forms of hypospadias, despite evidence on the best surgical technique is lacking (2, ⊕⊕○○).

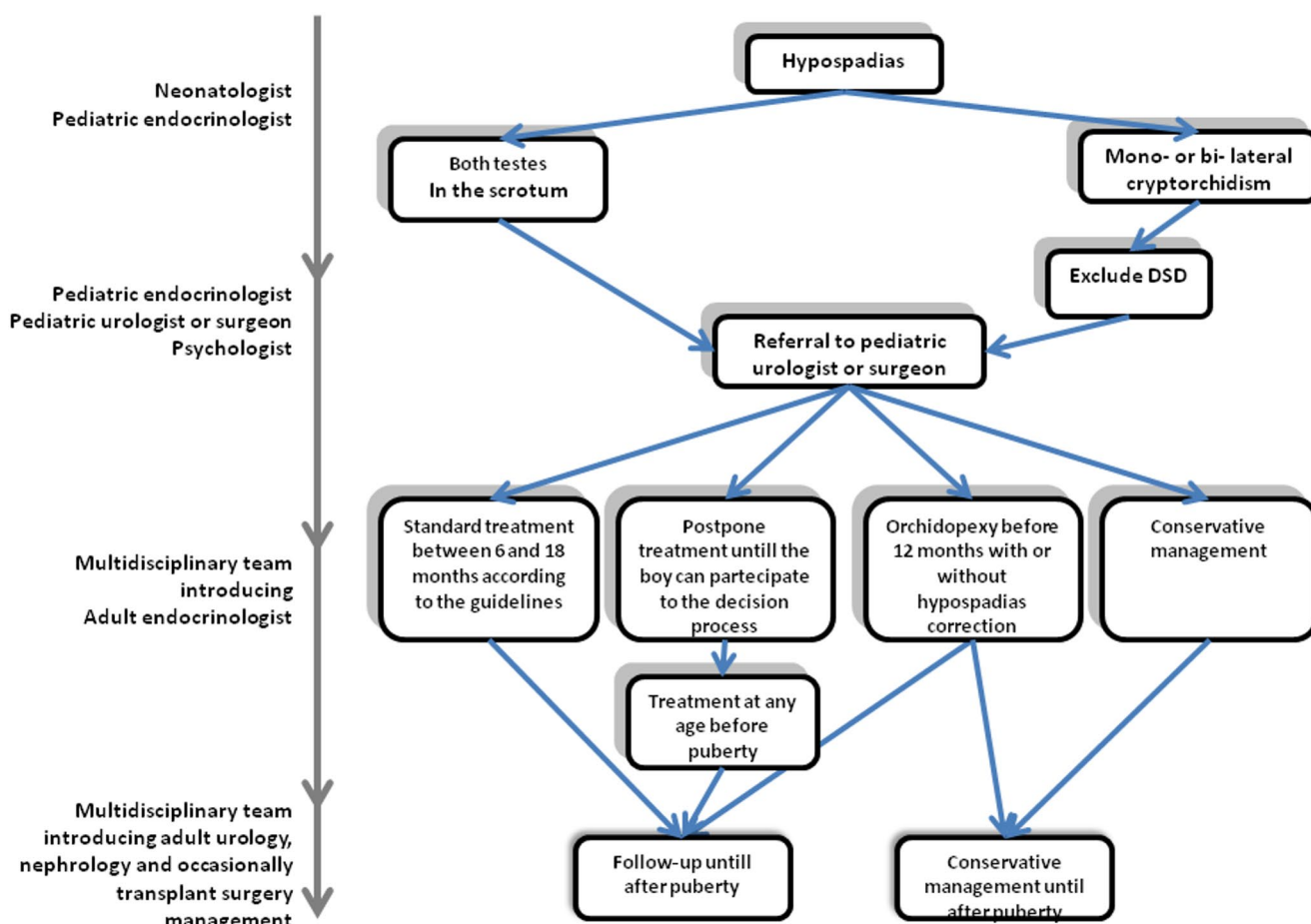


Fig. 1 Proposal algorithm for referral and treatment of hypospadias. (Modified from Ref. [235].

R4.10 We suggest preoperative hormonal therapy only for proximal hypospadias or hypospadias with microphallus (2, ⊕⊕○○).

R4.11 We recommend that the adolescents/young adults with hypospadias who underwent childhood repair should be adequately involved in a long-term follow-up to detect any physical/psychosocial problem that may emerge during the transition from adolescence to adulthood (1, ⊕⊕⊕○).

R4.12 We suggest using validated objective scoring systems to assist in evaluating the functional and cosmetic outcome of patients who underwent surgical repair (1, ⊕⊕⊕○).

Evidence

Surgical repair is not necessary for patients with mild defects (partial or standard distal forms of Table 3). This conservative approach is supported by a report of 56 adults with unrepaired hypospadias including 44 with a mild form who were satisfied with the appearance of their penis, voided in standing position and did not have infertility associated with the abnormal position of the urethral meatus [226].

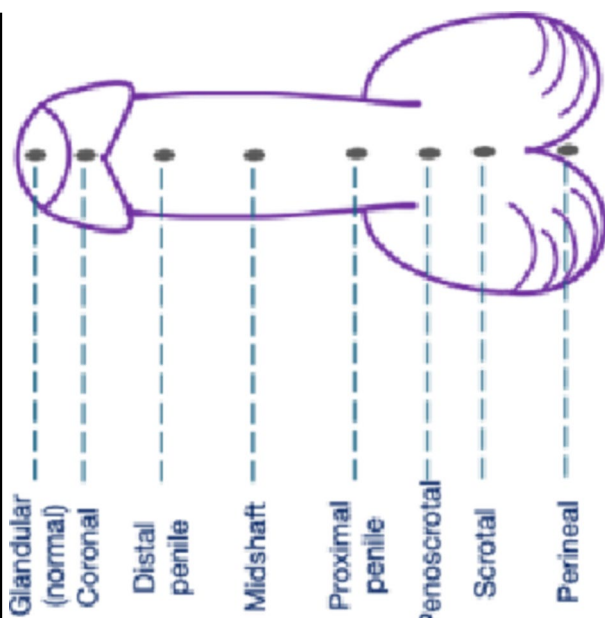
Urologic referral and potentially surgical repair are indicated for patients who are at risk of: (a) significant deflection of the urinary stream; (b) inability to urinate from a standing position; (c) erectile dysfunction due to penile curvature leading to intercourse difficulties; (d) fertility issues due to sperm deposition difficulties; (e) concern about developmental problems based on the appearance of the penis (Baskin L, 2025, www.UpToDate.com).

The choice of the age range 6–18 months for primary hypospadias repair arises from the awareness that emotional, cognitive, and body image development may be affected by both the genital anomaly and the reconstructive surgery. These psychological factors matter, as the child's reaction to both the surgery and the anesthetic trauma varies strongly with age. Postoperative behavioral problems such as aggressive or regressive behavior, night terrors, and anxiety may be more common at certain ages, particularly at 1 to 3 years of age. The period from 6 to 15 months is a relatively good time for surgery from the viewpoint of emotional development, provided parent/child separation is minimized [227]. However, age at surgery is not a risk factor for urethroplasty complication in pre-pubertal tubularised incised plate

Table 9 Classification of hypospadias based on physical findings (Modified from [206])

Type	Frequency (%)	Urethral location	Penile curvature [^]	Foreskin appearance	Management
Partial forms (incomplete presence of hypospadias)	10	Normal: with a urethral pit or ectopic urethra on the distal glans	Normal	Normal to mild asymmetric ventral deficiency	No surgical correction or evaluation needed
Standard (normal glans: MD* $\geq$ 1.4 cm)	40	Distal: on glans or coronal margin	Normal to mild	Ventral deficiency with a dorsal covered appearance	Surgical correction is an option; families may choose observation; no other evaluation
	25	Proximal: on penile shaft, at the penoscrotal connection, or within the scrotum	Moderate		Surgical correction is suggested; no further evaluation
Severe (abnormally small glans: MD* < 1.4 cm)	20	Scrotal or perineal	Severe	Ventral penile foreskin tethering (short urethral plate) or fusion of the foreskin to the scrotum	Surgical correction required (usually two stages procedure); endocrine/genetic evaluation to detect DSD
Us with intact prepuce		Large urethral opening at the coronal margin	Normal	Normal	Correction usually performed on family preference; no other evaluation

[^]Penile curvature is assessed when the penis is erect and classified as normal (0 to 15 degrees), mild (15 to 30 degrees), moderate (30 to 75 degrees), and severe (> 75 degrees); *MD: maximal diameter



urethroplasty (TIP) repair [228]. On the contrary, complication rate after primary TIP repair was 2.5 times higher in adults than in the pediatric group according to a prospective controlled study [229].

Two systematic reviews and metanalysis assessed whether dorsal inlay graft urethroplasty (DIGU) had any additional advantages over standard tubularised incised plate urethroplasty (TIPU) repair in children with primary hypospadias. There is not enough evidence to suggest that either technique offers more favorable outcomes. Until more robust randomized clinical trials (RCTs) exist, decisions regarding the appropriate repair should be based on the surgeon's experience and outcomes [230, 231].

The systematic review of Borkar et al., based on RCTs, provides evidence supporting the use of double dartos flap (DDF) over single dartos flap (SDF) in hypospadias repair, particularly in distal hypospadias using the TIPU procedure [232].

Hypospadias repair following neonatal circumcision in the absence of preputial skin is a challenging reconstruction. The reoperation rate in a paper of Erzberg et al. was 30%, like reoperative hypospadias surgery. Parents of newborns diagnosed with hypospadias should be encouraged to refrain from pre surgical routine neonatal circumcision [233].

There is no consensus on the best surgical approach to correcting severe hypospadias. Despite more than 250 different techniques for hypospadias repair, successful outcome depends mainly on the surgeon's skills and the availability of appropriate tissue [234, 235]. A recent meta-analysis comparing the outcomes of single stage (foreskin pedicled tube) versus two stage (foreskin free graft, FFG & foreskin pedicled flap, FPF) repair for proximal hypospadias in the last decade found that two-stage repair of proximal hypospadias had significantly less complications compared to single stage repair [236]. Among two-stage repairs specific complications were significantly less for FFG, although total complications were not significantly different from that seen with FPF. The results of two-stage repairs improved with higher case load supporting the concept of dedicated hypospadias centres [236].

Chua et al. systematically evaluated the effect of preoperative hormonal therapy (PHS) on postoperative complications rates following hypospadias repair. Pooled effect estimates with moderate quality of evidence from three RCTs suggested that significant lesser postoperative complications occurred among patients exposed to PHS (RR 0.36, 95% CI 0.20–0.65) [237].

Surgery for hypospadias remains challenging and complications may occur lifelong: early after surgery, during puberty due to fast growth, and in adulthood. The functional objectives of surgery are to create a normal urinary stream, normal erections, and normal coitus with a satisfactory

cosmesis as well [238]. However, these objectives can be elusive and multiple operations including free graft harvesting have been described for hypospadias repair. As boys reach puberty, despite initial successful surgical treatment of posterior hypospadias, penile development can result in new functional concerns such as fistula enlargement, worsening penile curvature, and poor cosmesis [239–241]. Patients with hypospadias should be routinely instructed to return for follow up after puberty. At that point, these teenagers should be reassessed with detailed physical examination and uroflowmetry. An open discussion of all available relevant medical data, including progressive information on any until now insufficiently communicated aspects of the condition, is crucial (Fig. 2) [106]. In addition, completing the Hypospadias Objective Penile Evaluation (HOPE) questionnaire or other validated scales to assess the functional, social and psychological outcomes will allow for a more objective and generalizable evaluation of the repair [242–245].

The network meeting “Lifelong Posterior Hypospadias” was organized to identify what is currently missing in the lifelong treatment of posterior hypospadias, to improve care, quality of life, and awareness for these patients. The participants concluded that patients should be more involved in defining desired treatment approach and outcome measures. For optimal outcome evaluation, standardization of data collection with cooperation between basic researchers from different centers, as well as involving clinicians and patients is necessary [246].

Remarks

Surgery of milder forms of hypospadias is an option for some families who want to pursue reconstruction to improve cosmetic appearance for religious (circumcision) or social reasons. When circumcision is desired by the family, it should be delayed until a freehand circumcision can be safely performed in the operating room later in life (Baskin L, 2024, www.UpToDate.com).

Repeated follow-up and psychological support during childhood/adolescence is of great importance especially for patients with more proximal hypospadias. In fact, a case-control study performed in Sweden to assess a possible negative influence on the psychosocial outcome in adult males with hypospadias showed that “distal” patients responded similarly to controls on several measures, and 70% did not desire additional medical follow-up; on the contrary, there were clear psychosocial differences in patients with mid-shaft and proximal hypospadias that warranted additional support [247].

Teenagers with hypospadias who will be transitioning into an adult urology practice need to have ownership of

Neonatal period	Age 4 years	Age 6 years	Early puberty (9–12 years)	Late pubertal age (14–18 years)	Adulthood (>18 years)
Information to family about diagnostic work-up and possible interventions according to hypospadias severity	Information to child about penile conformation and function	Continue information and answer questions; discuss puberty in case of hypogonadism; consider introducing privacy during PE	Privacy during PE; All information given; discuss transition	Organize transition; joint consultation(s); check knowledge and autonomy	Continued update of all information; discussion of recommended follow-up visits and screenings
-----Psychological and Peer support-----					
Involve pediatric urologist or surgeon in case of severe hypospadias	Words and vocabulary; Principles of body functioning, hormones and condition; shame and secrecy	Naming condition; Puberty (induction); fertility; bullying and isolation	Sexuality; testes surveillance; peer and relations	Fertility and gamete preservation; partnership and sexual functioning	Overall QoL; quality of sexual life; PS functioning; fertility issues.
Timing and topics might vary largely between individuals					
Information provided to parents			Information provided to affected individuals		

Fig. 2 The present figure reproduces the structure and partially modifies the content of fig. 2 in reference [106], after adaptation to hypospadias condition. Multidisciplinary care and data collection start at diagnosis and maintain throughout the individual's life. The focus of the information process gradually shifts from the parents to the affected child. Psychological and peer support are key elements at all ages.

their condition and be aware of what happened in the past in terms of number of previous surgeries and type of surgical technique used. This type of pediatric patient maturation will allow for a smoother transition to adult specialists, even when their medical records are not available [248, 249].

Epididymitis and orchitis

Epidemiology

Orchitis and epididymitis are inflammation of the testis and epididymis, respectively, with or without infection.

Isolated orchitis is rare and usually accompanied by epididymitis, hence the true incidence is unclear [250];

With appropriate individual adaptations, children should be informed of their conditions at an early age. Some of the suggested themes to be discussed by the team members are shown in boxes at the upper part of the figure and the lists of some important topics within these themes are represented in boxes at the lower part of the figure. PE, physical exam; PS, psychosexual; QoL, quality of life

moreover, current literature does not suggest any differential occurrence according to race or religion [250].

The annual incidence of acute epididymitis is approximately 1.2 per 1,000 pre-pubertal boys (mean age = 11 years), and it is approximately 1 per 144 from post-pubertal to transition; about 1/4 of patients in post-pubertal and transition age has recurrence within five years [251, 252].

Pathophysiology

Orchitis is the unilateral or bilateral inflammation of the testis, usually caused by viral or bacterial infections, while epididymitis is the inflammation of the epididymis and represents the most common intrascrotal inflammatory condition [250–252]. Both can be classified as acute (≤ 6 weeks) or chronic (> 6 weeks).

In children and adolescents, orchitis is usually viral, most commonly due to mumps or rubella, with mumps responsible for most isolated cases; rare cases have been reported after MMR vaccination [250].

Epididymitis typically results from retrograde pathogen ascent, most often bacterial [251, 252] although, non-infectious aetiologies such as vasculitis, autoimmune diseases, or medications (e.g., amiodarone) have been identified [253–258]. In prepubertal boys, epididymitis is generally a post-infectious inflammatory reaction to pathogens like mycoplasma, enteroviruses, or adenoviruses and usually follows a benign course. In postpubertal adolescents and young adults, sexually transmitted infections are the primary cause [251, 252].

Clinical picture

Epididymo-orchitis presents with gradual onset scrotal pain, sometimes radiating to the lower abdomen. Acute cases develop rapidly and last less than 6 weeks, whereas chronic cases progress more slowly and persist beyond 6 weeks. Symptoms may be associated with lower urinary tract infection or urethritis, including urethral discharge, dysuria, or penile irritation [259–267]. Testicular torsion is the main differential diagnosis and should be promptly considered as indicated below in the specific session.

Diagnostic evaluation

Recommendations and suggestions

R5.1 We recommend an andrological consultation in case of symptoms and signs of epididymo-orchitis to confirm the clinical diagnosis (1, ⊕⊕⊕⊕).

R5.2 We recommend performing bilateral scrotal ultrasonography (1, ⊕⊕⊕⊕).

R5.3 We suggest for *Neisseria Gonorrhoeae* and *Chlamydia Trachomatis* infections, acid amplification test (NAAT), in sexually active patients (2, ⊕⊕○○).

Evidence

In mumps, orchitis develops in both pre-pubertal and post-pubertal groups of patients, more rarely in pre-pubertal patients, and in 14–35% of patients in post-pubertal and transition age; the clinical syndrome develops 4 to 8 days after parotitis but can also occur in its absence [250]. Different viruses causing orchitis include varicella-zoster virus, coxsackievirus, echovirus, and cytomegalovirus [250]. Bacterial infections of the prostate and urinary tract infection can cause orchitis. Common causes of bacterial orchitis include *Escherichia coli*, *Klebsiella pneumoniae*,

Pseudomonas aeruginosa, and *Staphylococcus* and *Streptococcus*. Bacteria causing sexually transmitted infections can also determine orchitis in sexually active males; common pathogens include *Neisseria Gonorrhoeae* and *Chlamydia Trachomatis* [250]. Risk factors for epididymitis in pre-pubertal boys include recent urinary tract surgery or instrumentation procedures and anatomic abnormalities, such as posterior urethral valves or meatal stenosis, whereas risk factor in post-pubertal up to transition age include sexual activity, strenuous physical activity, bicycle or motorcycle riding, and prolonged periods of sitting, and epididymitis is most commonly caused by sexually transmitted infection by *Neisseria Gonorrhoeae* or *Chlamydia Trachomatis* [251, 252].

Remarks

Since sexually transmitted infections can rarely be found in pre-pubertal boys, diagnostic evaluation using NAAT must be guided by the presence of risk factors such as belonging to ethnic groups at risk, signs of abuse, presence of associated urethritis.

Therapeutic management

Recommendations and suggestions

R5.4 We recommend the use of anti-inflammatory treatment for symptomatic epididymo-orchitis (1, ⊕⊕⊕⊕).

R5.5 We suggest for *Neisseria Gonorrhoeae* infection: ceftriaxone i.m. once plus azithromycin orally once; if ceftriaxone is not available, we suggest cefixime orally once plus azithromycin orally once, or in case of azithromycin allergy, doxycycline (only for men > 12 years) orally 2 times a day for 7 days (2, ⊕⊕○○).

R5.6 We suggest for *Chlamydia Trachomatis* infection: Azithromycin orally once, or Doxycycline (only for men > 12 years) orally 2 times a day for 7 days (2, ⊕⊕○○).

Evidence

Complications of epididymo-orchitis are testicular atrophy, with up to 60% of cases demonstrating some degree of atrophy; impaired fertility or sterility; reactive hydrocele [250].

Remarks

Considering the possible complications, an appropriate therapy must be set up quickly, and if sequelae develop, adequate follow-up must be set up for the patient, both ultrasound and seminological.

Testicular tumors

Testicular tumors (TT) in pediatric age (0–18 years) require specific diagnostic and therapeutic management, as they share similarities with adult testicular GCTs but have key differences. It is crucial to distinguish between pre-pubertal and post-pubertal tumors, as biology and treatment protocols have developed differently [268–271]. Pre-pubertal tumors are very rare, generally benign, and rarely associated with distant metastases. In contrast, post-pubertal tumors are more common, malignant in 75% of cases, and often (20–30%) present with distant metastases at onset [272].

Evidence in childhood is limited due to the low incidence of TT. Studies focused on adolescence and the transition age are also lacking, as these age groups are often included in adolescent/young adult (AYA, 15–39 years) studies.

Epidemiology

Testicular tumors (TT) are the most common solid malignancy in individuals aged 20 to 45 [273], but are less prevalent in pediatric population, although the incidence is increasing in the last decades [274]. Overall, the incidence in pediatric age is approximately 3%. There are two peaks in GCT incidence: one in young children (0–4 years) and another beginning at puberty [275]. TT are rare in childhood (1–2%) [276], but increase to 15% during adolescence [274].

Pathophysiology

Most testicular tumors in children and adolescents are germ cell tumors (GCTs), with histologic subtypes varying by age. Prepubertal tumors are almost exclusively benign, including mature and immature teratomas and dermoid cysts, while yolk sac tumors are malignant [277]. Post-pubertal tumors are predominantly non-seminomas—choriocarcinoma, yolk sac tumor, embryonal carcinoma, and teratoma, in pure or mixed forms [278], whereas pure seminomas are rare in this age group [279].

Pubertal status is critical for management: post-pubertal teratomas are treated as malignant due to the risk of metastasis, while prepubertal teratomas are always benign. The latest WHO classification also includes certain neuroendocrine variants among prepubertal teratomas [280].

Post-pubertal GCTs, like adult tumors, are often associated with germ cell neoplasia in situ (GCNIS) [2, 281], and signs of testicular dysgenesis, which are absent in prepubertal tumors [282, 283], suggesting distinct developmental origins [284]. Nevertheless, the etiology of pre-pubertal tumors remains unknown.

Extragenital GCTs can occur in children, typically along the midline (intracranial, pineal, mediastinal) [284]. Sex cord-stromal tumors—including juvenile granulosa cell, Leydig, and Sertoli cell tumors—are rare (10–13%) and usually benign in children, though malignancy is occasionally observed in adolescents and young adults [285–287].

Clinical picture

Testicular tumors usually present as a firm, palpable mass, often painless but sometimes associated with discomfort or swelling. Detection often occurs via parents, caregivers, or pediatricians. Post-pubertal tumors are often diagnosed at more advanced stages than in children or adults [278, 288, 289]. In adolescents, diagnosis is frequently delayed due to reluctance to report symptoms [290], and the transition from pediatric to general care [291].

Diagnostic evaluation

Recommendations and suggestions

R6.1 We recommend performing scrotal ultrasound (US) in the presence of a palpable testicular mass, elevated serum tumor markers (STMs), or retroperitoneal/visceral masses (1, ⊕⊕⊕⊕).

R6.2 We recommend regular testicular examination and/or testicular US for individuals with risk factors, starting at the onset of puberty (Expert opinion).

R6.3 We suggest regular testicular US starting at puberty for patients with incidental finding of testicular microlithiasis and other known risk factors for testicular tumors (2, ⊕⊕○○).

R6.4 We recommend measuring serum tumor markers (α -FP, β hCG, LDH) at diagnosis, prior to any treatment, and during follow-up in post-pubertal patients. In pre-pubertal patients α -FP is sufficient to discriminate between malignant and benign tumors (1, ⊕⊕⊕⊕).

R6.5 In cases of small lesions with indeterminate findings at testicular US and normal STMs, we suggest repeating the US within 8 weeks. For selected indeterminate cases, consider second-level exams such as magnetic resonance imaging (MRI) or contrast-enhanced ultrasound (CEUS), (2, ⊕⊕○○).

Evidence

Risk factors for GCT are detailed in Table 10. Testicular cancer is etiologically linked to testicular dysgenesis syndrome (TDS), a condition characterized by poor testicular development, dysfunctional testicular somatic cells, and genital malformations such as cryptorchidism [292] and

Table 10 Risk factors for testicular germ cell tumors (GCT)

Risk factors for testicular GCT

Cryptorchidism
Hypospadia
Impaired germ cell production
First degree relative with GCT
Personal history of GCT
Down syndrome
Low birth weight
Premature birth
Testicular microlithiasis (if other risk factors)

hypospadias (see specific sections). TDS likely results from a combination of inherited genetic variation and environmental factors acting during early development, leading to impaired germ cell differentiation in adulthood [2, 281]. Additional risk factors include birth defects [293], a family history of GCT in a first-degree relative [294] and personal history of GCT [295]. Conflicting data exist in regard with testicular microlithiasis, which is associated with testicular tumor only in patients with additional risk factors [296–299]. A recent metanalysis including 595 children with testicular microlithiasis concluded that microlithiasis was not associated with testicular malignancy during childhood. However, the risk increases during transition age and adulthood follow-up, especially when other risk factors coexist [300].

A testicular physical examination is critical for detecting a mass and assessing its characteristics and should always be performed by primary health care providers in all patients, especially in those with risk factors. Testicular US is the diagnostic gold standard to identify testicular lesions (even the not palpable ones) [301, 302] and should be performed in all boys and adolescent with a palpable testicular mass, elevated serum tumor markers, retroperitoneal or visceral masses or in case of risk factors since puberty [303, 304].

STM assessment contribute to diagnosis, staging, and prognosis in testicular lesions. Common STM for GCT are alpha-fetoprotein (α -FP), human chorionic gonadotropin (β -hCG), and lactate dehydrogenase (LDH). α -FP is crucial in pre-pubertal patients as is secreted by yolk sac tumors, the only malignant form in childhood. Evidence suggests that no male over 6 months of age with a testicular lesion and α -FP < 100 ng/mL has a malignant tumor [305]. Therefore, its measurement is particularly useful for therapeutic decision.

In post-pubertal patients, α -FP, β -hCG, and LDH are crucial markers and should always be tested in cases of testicular lesions or suspected extragonadal GCT. β hCG is significantly elevated in choriocarcinoma, about 15–20% of seminomas, and less frequently in embryonal carcinoma. Nonetheless, a negative result for serum tumor markers does not exclude the diagnosis of GCT [306]. Although

LDH is a general inflammation marker, high levels indicate a large tumor burden. However, staging and decision-making should not be based on LDH alone [307].

In cases where testicular ultrasound results are indeterminate and serum tumor markers are normal; it is advisable to repeat imaging within 8 weeks for close monitoring. Additionally, in selected cases, more advanced imaging modalities such as CEUS and MRI can be employed to further refine the diagnosis [308–310].

Differential diagnosis should include orchitis, testicular trauma, adrenal rest tumors, extra-testicular tumors or testicular localization of leukemia or lymphoma [301].

After diagnosis confirmation, total body imaging and post-operative STMs are needed for staging. Staging is not required for benign prepubertal tumors [276]. Pre-pubertal staging is based on Children's Oncology Group (COG) staging system while post-pubertal staging uses American Joint Commission on Cancer (AJCC) TNMS system and IGCCCG system for metastatic disease [272, 311]. The different staging systems for pre-pubertal and post-pubertal patients complicate comparisons across trials and the development of risk-based treatment strategies.

Imaging of the chest, abdomen, and pelvis using cross-sectional techniques is essential for staging, with computed tomography (CT) being the primary approach. In certain cases, such as when there are neurological symptoms, known chest metastases, or elevated β hCG (> 5000 IU/L) or α FP (> 10,000 ng/ml) levels, MRI of the brain may also be indicated. Staging imaging can be conducted either before or after the primary tumor is surgically removed, depending on clinical context and suspicion of malignancy. At least abdomen MRI is recommended in sex cord-stromal tumors as 10% are malignant with poor prognosis [312].

Remarks

Educating patients, especially adolescents, on self-examination is a fundamental step for the early diagnosis of testicular tumors and should be the focus of widespread prevention campaigns. Early diagnosis is strongly associated with a better prognosis for these tumors, particularly in adolescents.

The medical history is crucial for accurately identifying risk factors and guiding the differential diagnosis. Pediatricians should inform patients with risk factors, especially those with present or previous cryptorchidism, as well as their parents/caregivers and the general practitioners responsible for their care post-pediatric discharge, about the importance of regular morpho-functional testicular examinations starting at puberty.

In cases where physical examination suggests a testicular mass, is imperative to perform a testicular US at specialized centers with expertise in pediatric and adolescent testicular

pathology. This ensures accurate diagnosis and staging and therefore appropriate therapeutic management.

Therapeutic management

Recommendations and suggestions

R6.6 We recommend a multidisciplinary management approach for TT in specialized centers with expertise in pediatric and adolescent care, paying particular attention to the transition to ensure long term follow-up in adulthood (1, ⊕⊕⊕⊕).

R6.7 We recommend determining pubertal status before any treatment (1, ⊕⊕⊕⊕).

R6.8 We recommend radical inguinal orchiectomy in case of suspicious malignant lesion and normal contralateral testis, in post-pubertal TT (1, ⊕⊕⊕⊕).

R6.9 We recommend performing testis-sparing surgery with intraoperative frozen-section examination in pre-pubertal tumors if STMs are negative (high suspicion of benign lesion) (1, ⊕⊕⊕⊕).

R6.10 We recommend discussing testis-sparing surgery with frozen section examination in patients with a high likelihood of having a benign TT which are suitable for enucleation (1, ⊕⊕⊕⊕).

R6.11 We recommend sperm banking in all post-pubertal patients, prior to adjuvant treatment, retroperitoneal lymph node dissection (RPLND). In patients without a normal contralateral testis or known subfertility, cryopreservation should be considered prior to orchiectomy (1, ⊕⊕⊕⊕).

R6.12 We recommend assessing pubertal stages until completion during follow-up, especially in patients who underwent adjuvant treatments or in patients with low-volume survival testicle, (1, ⊕⊕⊕⊕).

R6.13 We recommend oncology consultation after orchiectomy for proper treatment/follow-up, preferably in centers with expertise in pediatric and adolescent care (1, ⊕⊕⊕⊕).

R6.14 We suggest routine contralateral testicular US, even over a long period of time during adulthood (2, ⊕⊕⊕⊕).

Evidence

Pre-pubertal tumors Partial orchiectomy with intraoperative frozen section has become standard practice due to the high rate of benign lesions and should be performed whenever possible [313]. Orchiectomy should be considered for very large lesions occupying the entire testicle, or in case of α -FP > 100 ng/mL [305]. 80% of patients with yolk sac tumors present at clinical stage I. Approximately 80% of patients with yolk sac tumors present at clinical stage I, with the few metastatic cases usually confined to retroperitoneal

involvement. Platinum-based chemotherapy is highly effective for treating the disease and is typically administered before RPLND [276].

Post-pubertal tumors If clinical findings suggest unilateral testicular malignancy, radical orchiectomy is the treatment of choice. Partial orchiectomy may be considered for lesions highly likely to be benign based on first and/or second level exams [301, 308, 309], especially if the tumor is small, the patient has an anatomically or functionally solitary testicle, or in case of bilateral synchronous malignancies. An intraoperative frozen section of the mass with an experienced pathologist is recommended [314, 315]. If there is any concern for GCT, a radical orchiectomy should be performed. If the lesion is confirmed benign, the testicle will be preserved.

All post-pubertal boys should be offered sperm cryopreservation before initiating any adjuvant treatment [316–318], included RPLND [319] or prior to orchiectomy in case of abnormal contralateral testis or known subfertility [320, 321].

Treatment regimens for pediatric GCT have largely been based on clinical trial results from adult men. Whether these results apply equally to children with gonadal or extragonadal GCT remains to be established. To minimize side effects (lung toxicity, ototoxicity, nephrotoxicity) some pediatric protocols used reduced dose of bleomycin from once per week to once per cycle (PEb) [322–324] or carboplatin instead of cisplatin [325, 326]. However, there is some evidence that PEb use may have contributed to poorer outcomes in adolescents [278, 327]. Current ongoing trials will help to better understand the correct protocol for young patients.

About half of post-pubertal testicular GCTs present as clinical stage I. In these cases, active surveillance is the current recommendation [328, 329]. The relapse rate is 20–30%, with excellent survival after salvage therapy [330]. In patients with high-risk pathologic features on the orchiectomy specimen (large size, >44–50% embryonal carcinoma histology, lymphovascular invasion), adjuvant carboplatin-based chemotherapy can be suggested [331].

Radiotherapy has been used for many years, but current evidence demonstrates a non-inferiority of chemotherapy with less impact on fertility, hypogonadism, and onset of second malignancies [332–334]. RPLND is differently recommended based on guidelines: only after chemotherapy for persistent masses > 1 cm, or in stage II patients who have retroperitoneal-only disease and normal STMs [272].

Adolescent patients with non-seminomatous GCTs had slightly improved survival compared with adults, despite presenting with more advanced disease, achieving excellent

survival outcomes [288]. Therefore, follow-up can be long and should take into consideration several issues: (1) Leydig cell dysfunction and hypogonadism may prematurely develop in 10–15% of patients, increasing risk of hypogonadism in adulthood [335]. Monitoring gonadal function from the early stages of pubertal development is necessary in these patients [152]. (2) Adult male survivors who underwent chemotherapy face increased risks of early-onset cardiovascular disease and second malignancies. (3) Literature data reported a 1–3% lifetime risk of contralateral TGCT, even after many years especially if the first tumors occur in young age [295]. Routine testicular US is strongly suggested, even over the canonical oncological follow-up period. (4) An increased prevalence of depression is documented among patients with pediatric TT [336], especially in adolescents, who often need psychological support, due to the difficulty in expressing discomfort or addressing concerns with both family members and doctors [337].

Remarks

The assessment of pubertal stage is critical for a correct therapeutic management. TT management is decreed by stage, histology, and risk classification and differs between pre-pubertal and post-pubertal patients. The young age of the patient should not necessarily lead to a less aggressive approach, especially given the higher tendency for metastatic disease in post-pubertal young patients. However, the young age necessitates fertility preservation and a more attentive endocrinological follow-up aimed at achieving complete pubertal development. The psychological repercussions of this challenging chapter in the patients' lives should not be overlooked.

Testicular torsion

Epidemiology

Testicular torsion (TT) exhibits varying incidence rates depending on the age group considered, with a characteristic bimodal distribution. The two peak ages of occurrence are in infancy and early adolescence. TT accounts for 10–25% of acute scrotal events in the pediatric population [257, 338–340]. Its incidence is reported as 1 in 4000 males under 25 years and 1 in 1500 males under 18 years [339, 341, 342].

Pathophysiology

TT occurs when the testis twists around the spermatic cord, compressing the arterial inflow and causing ischemia and

acute scrotal pain [343]. Most affected individuals have no underlying conditions [344]. However, congenital anomalies can increase the risk by allowing excessive testicular mobility. The “bell-clapper” deformity, in which the tunica vaginalis wraps abnormally around the spermatic cord, leaves the testis inadequately attached to the posterior scrotum, facilitating torsion [343, 345].

Other risk factors include larger testicular volume, testicular tumors, horizontal testicular lie, previous cryptorchidism, a long intrascrotal spermatic cord, and less commonly trauma or cold exposure [343, 344, 346].

Clinical picture

TT typically presents with sudden, unilateral scrotal pain [347, 348], often accompanied by nausea, vomiting, a high-riding and firm testis, absent cremasteric reflex, and erythematous scrotal skin [260, 349, 350].

Diagnostic evaluation

Recommendations and suggestions

R7.1 We recommend prompt and accurate diagnosis of acute-onset scrotal pain through urological evaluation in the emergency department within 6 h, utilizing the TWIST (Testicular Workup for Ischemia and Suspected Torsion) score (1, ⊕⊕⊕⊕).

R7.2 We recommend direct surgical exploration for all cases of acute-onset scrotal pain without trauma and a high-risk TWIST score (1, ⊕⊕⊕⊕).

R7.3 We recommend the use of Power Doppler Ultrasonography of the scrotum as the initial imaging modality, to be performed by a specialist, in cases of acute-onset scrotal pain without trauma and an intermediate or low-risk TWIST score (1, ⊕⊕⊕⊕).

R7.4 We suggest, where available, the use of contrast-enhanced MRI to aid in diagnosis, provided it does not delay the diagnostic process. However, we do NOT recommend it in high-risk cases, which should be promptly referred for surgical exploration (2, ⊕⊕⊕⊕).

Evidence

TT is considered a surgical emergency that requires prompt identification and expert management, as it can threaten testicular viability, thus causing irreversible damage and potentially impacting future fertility if not promptly (within 6 h) and appropriately treated [347, 351–353]. TT must be differentiated from other potential causes of acute scrotal pain without trauma, though clinical presentation overlaps should be considered [354]. One such condition is

Table 11 TWIST (Testicular Workup for Ischemia and Suspected Torsion) score

Clinical variables	Points
Presence of testicular swelling	2 points
Presence of hard testicle	2 points
Absence of cremasteric reflex	1 point
Presence of high-riding testicle	1 point
Presence of nausea/vomiting	1 point
Score	Risk grading
6–7 points	High risk
1–5 points	Intermediate risk
0 points	Low risk

appendageal torsion (involving the appendix testis and appendix epididymis, both remnants of the Mullerian and Wolffian systems, respectively), which is more common than TT in prepubertal boys. Another is epididymitis (see also specific chapter), which typically presents with a more gradual onset of pain [347].

The primary goal during the initial evaluation of acute scrotal pain without trauma is to assess the clinical and physical presentation (as outlined in Table 11) to determine the TWIST (Testicular Workup for Ischemia and Suspected Torsion) score. This clinically validated scoring system for pediatric and transitional-age patients (3 months–18 years) has proven to have high positive predictive values [354, 355]. Cases with a high-risk TWIST score, strongly suggestive of TT, should proceed immediately to urological consultation and surgical exploration. Those with an intermediate or low-risk TWIST score and a questionable diagnosis may proceed to Power Doppler Ultrasonography (US) of the scrotum. Power Doppler has been shown to be more sensitive than color Doppler US in evaluating areas with slow blood flow, such as in the prepubertal testis, with a sensitivity of 63% to 86% and a specificity of 97% to 100% [356–358]. The presence of a twisted spermatic cord, often seen as the “whirlpool sign” (characterized by a sudden change in the course of the spermatic cord, forming a spiral twist at the level of the external inguinal ring or within the scrotal sac), and/or absent blood flow within the affected testicle confirms TT and necessitates immediate surgical exploration [354, 359–361]. Conversely, normal or increased blood flow, suggestive of inflammation or appendageal torsion, would guide appropriate treatment [354].

While MRI techniques were introduced in 2006, they are not typically employed for acute scrotum evaluation due to their time-consuming and costly nature compared to US. However, dynamic contrast-enhanced MRI, which assesses perfusion and apparent diffusion coefficient (ADC), may help differentiate TT from other acute scrotal conditions. A decrease or lack of perfusion in conjunction with lower mean ADC values has been shown to have a sensitivity of 93% and specificity of 100% for diagnosing TT [362–364].

Hematological parameters may assist in the differential diagnosis between TT and epididymo-orchitis, though large multicenter prospective studies are still needed to validate their clinical utility [357, 365].

Remarks

Time is critical in the diagnostic evaluation of acute-onset scrotal pain, and the workup should be completed within 6 h of pain onset, considering any potential delays before medical consultation. No diagnostic test should delay surgical exploration if TT is strongly suspected. While TT is less common than other causes of acute scrotal pain, early intervention is essential to preserve testicular viability.

Therapeutic management

Recommendations and suggestions

R7.5 We recommend rapid surgical treatment of TT, ideally within 6 h of acute scrotal pain onset, to maximize the chances of testicular tissue survival (1, ⊕⊕⊕⊕).

R7.6 We suggest performing manual detorsion, following the administration of analgesics and sedation, if surgery is not immediately available or while preparing for surgical exploration (2, ⊕⊕⊕○).

R7.7 We recommend against manual detorsion as a substitute for surgical exploration (1, ⊕⊕⊕○).

Evidence

There is a direct correlation between testicular survival and the duration of testicular torsion (TT), as well as the overall ischemic time. A prompt intervention within 6 h of acute scrotal pain onset is associated with a 97.5% testis salvage rate. However, after 25 to 48 h, only a small percentage of the affected testes will survive, depending on the degree of torsion [366–368].

Manual detorsion may be attempted if surgery is not immediately available or while preparing for surgical exploration. The return of blood flow to the affected testicle can be clinically assessed by pain relief and objectively confirmed with Doppler ultrasound. However, manual detorsion should not delay or replace surgical intervention [366, 369].

A transscrotal surgical approach is typically employed to access the affected testicle and perform detorsion of the spermatic cord [257, 370]. Once detorsion is performed, testicular viability must be assessed. If the testicle is deemed viable, it should be permanently fixed within the scrotum through orchidopexy. If the testicle is nonviable or necrotic, it should be removed [277, 371].

Contralateral orchidopexy should also be performed regardless of the affected testicle's viability, as some congenital malformations (e.g., “bell-clapper” deformity) are bilateral in up to 80% of patients, increasing the risk of TT in the other testicle [372–374].

Remarks

A rapid surgical approach is essential in cases of TT, and patients should be preoperatively counseled about the potential need for orchiectomy if the testicle is found to be necrotic or nonviable [375, 376].

Declarations

Conflict of interest Authors declare no competing interest.

Ethics standards This article does not contain any studies with human or animal subjects.

Research involving human participants and/or animals This article does not contain any studies with human participants or animals performed by any of the authors.

Informed consent For this type of study, formal consent is not required.

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References

- Juul A, Almstrup K, Andersson A, Jensen TK, Jorgensen N, Main KM, Rajpert-De Meyts E, Toppari J, Skakkebaek NE (2014) Possible fetal determinants of male infertility. *Nat Rev Endocrinol* 10:553–562. <https://doi.org/10.1038/nrendo.2014.97>
- Skakkebaek NE, Rajpert-De Meyts E, Main KM (2001) Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects. *Hum Reprod* 16:972–978. <https://doi.org/10.1093/humrep/16.5.972>
- Cangiano B, Sweet DS, Quinton R, Bonomi M (2021) Genetics of congenital hypogonadotropic hypogonadism: peculiarities and phenotype of an oligogenic disease. *Hum Genet* 140:77–111. <https://doi.org/10.1007/s00439-020-02147-1>
- Hadziselimovic F, Herzog B (1994) Andrological problems in adolescence. *Ther Umsch* 51:305–313
- Cavarzere P, Sulpasso M, Maines E, Vincenzi M, Gaudino R, Monti E, Chironi C, Tato L, Antoniazzi F (2011) Serum inhibin B levels before and after varicocelectomy in early adolescence. *J Endocrinol Invest* 34:265. <https://doi.org/10.3275/7796>
- Foresta C, Garolla A, Frigo AC, Carraro U, Isidori AM, Lenzi A, Ferlin A (2013) Anthropometric, penile and testis measures in post-pubertal Italian males. *J Endocrinol Invest* 36:287–292. <https://doi.org/10.3275/8514>
- Delfino M, Elia J, Imbrogno N, Argese N, Mazzilli R, Toscano V, Mazzilli F (2012) Testicular adrenal rest tumors in patients with congenital adrenal hyperplasia: prevalence and sonographic, hormonal, and seminal characteristics. *J Ultrasound Med* 31:383–388. <https://doi.org/10.7863/jum.2012.31.3.383>
- Serefoglu EC, Saitz TR, La Nasa JAJ, Hellstrom WJ (2013) Adolescent varicocele management controversies. *Andrology* 1:109–115. <https://doi.org/10.1111/j.2047-2927.2012.00004.x>
- Maggi M, Buvat J (2013) Standard operating procedures: pubertal tarda/delayed puberty–male. *J Sex Med* 10:285–293. <https://doi.org/10.1111/j.1743-6109.2012.02678.x>
- Fast AM, Deibert CM, Van Batavia JP, Nees SN, Glassberg KI (2014) Adolescent varicocelectomy: does artery sparing influence recurrence rate and/or catch-up growth? *Andrology* 2:159–164. <https://doi.org/10.1111/j.2047-2927.2013.00142.x>
- Foresta C, Garolla A, Zuccarello D, Pizzol D, Moretti A, Barzon L, Palu G (2010) Human papillomavirus found in sperm head of young adult males affects the progressive motility. *Fertil Steril* 93:802–806. <https://doi.org/10.1016/j.fertnstert.2008.10.050>
- Pereira NM (1999) Phimosis, hydrocele and varicocele. 3 frequent pathologies in the male child and adolescent. *Acta Med Port* 12:137–143
- Bonomi M, Cangiano B, Cianfarani S, Garolla A, Gianfrilli D, Lanfranco F, Rastrelli G, Sbardella E, Corona G, Isidori AM, Rochira V (2025) Management of andrological disorders from childhood and adolescence to transition age: guidelines from the Italian Society of Andrology and Sexual Medicine (SIAMS) in collaboration with the Italian Society for Pediatric Endocrinology and Diabetology (SIEDP)-Part-1. *J Endocrinol Invest* 48:1–22. <https://doi.org/10.1007/s40618-02435-x>
- Karara N, Corsello A, del Torso S, Hadjipanayis A, Magendie C, van Esso D, Grossman Z (2025) Paediatric Primary Care Across Europe: A Survey of 42 Countries. *Acta Paediatr*. <https://doi.org/10.1111/apa.70404>
- Ercan O, Alikasifoglu M, Erginoz E, Janda J, Kabicek P, Rubino A, Constantopoulos A, Ilter O, Vural M (2009) Demography of adolescent health care delivery and training in Europe. *Eur J Pediatr* 168:417–426. <https://doi.org/10.1007/s00431-008-0759-1>
- Swiglo BA, Murad MH, Schunemann HJ, Kunz R, Vigersky RA, Guyatt GH, Montori VM (2008) A case for clarity, consistency, and helpfulness: state-of-the-art clinical practice guidelines in endocrinology using the grading of recommendations, assessment, development, and evaluation system. *J Clin Endocrinol Metab* 93:666–673. <https://doi.org/10.1210/jc.2007-1907>
- Lucaccioni L, Trevisani V, Boncompagni A, Marrozzini L, Berardi A, Iughetti L (2021) Minipuberty: Looking Back to Understand Moving Forward. *Front Pediatr* 8:612235. <https://doi.org/10.3389/fped.2020.612235>
- Rohayem J, Alexander EC, Heger S, Nordenstrom A, Howard SR (2024) Mini-Puberty, Physiological and Disordered: Consequences, and Potential for Therapeutic Replacement. *Endocr Rev* 45:460–492. <https://doi.org/10.1210/edrv/bnae003>
- Becker M, Hesse V (2020) Minipuberty: Why Does it Happen? *Horm Res Paediatr* 93:76–84. <https://doi.org/10.1159/000508329>
- Boehm U, Bouloux P, Dattani MT, de Roux N, Dode C, Dunkel L, Dwyer AA, Giacobini P, Hardelin J, Juul A, Maghnie M, Pitteloud N, Prevot V, Raivio T, Tena-Sempere M, Quinton R, Young J (2015) Expert consensus document: European Consensus Statement on congenital hypogonadotropic

- hypogonadism—pathogenesis, diagnosis and treatment. *Nat Rev Endocrinol* 11:547–564. <https://doi.org/10.1038/nrendo.2015.112>
21. Davis SM, Howell S, Janusz J, Lahlou N, Reynolds R, Thompson T, Swenson K, Wilson R, Ross JL, Zeitler PS, Tartaglia NR (2025) Testosterone Effects on Short-Term Physical, Hormonal, and Neurodevelopmental Outcomes (TESTO) in Infants with 47,XXY. *J Clin Endocrinol Metab*. <https://doi.org/10.1210/clinem/dgaf217>
 22. Kohva E, Huopio H, Hietamaki J, Hero M, Miettinen PJ, Raivio T (2019) Treatment of gonadotropin deficiency during the first year of life: long-term observation and outcome in five boys. *Hum Reprod* 34:863–871. <https://doi.org/10.1093/humrep/dez040>
 23. Virtanen HE, Toppari J (2008) Epidemiology and pathogenesis of cryptorchidism. *Hum Reprod Update* 14:49–58. <https://doi.org/10.1093/humupd/dmm027>
 24. Acerini CL, Miles HL, Dunger DB, Ong KK, Hughes IA (2009) The descriptive epidemiology of congenital and acquired cryptorchidism in a UK infant cohort. *Arch Dis Child* 94:868–872. <https://doi.org/10.1136/adc.2008.150219>
 25. Berkowitz GS, Lapinski RH, Dolgin SE, Gazella JG, Bodian CA, Holzman IR (1993) Prevalence and natural history of cryptorchidism. *Pediatrics* 92:44–49
 26. SCORER CG (1964) The Descent of the Testis. *Arch Dis Child* 39:605–609. <https://doi.org/10.1136/adc.39.208.605>
 27. Barthold JS, Gonzalez R (2003) The epidemiology of congenital cryptorchidism, testicular ascent and orchiopexy. *J Urol* 170:2396–2401. <https://doi.org/10.1097/01.ju.0000095793.04232.d8>
 28. Anonymous (1992) Cryptorchidism: a prospective study of 7500 consecutive male births, 1984–8. John Radcliffe Hospital Cryptorchidism Study Group. *Arch Dis Child* 67:892–899. <https://doi.org/10.1136/adc.67.7.892>
 29. Hack WWM, Sijstermans K, van Dijk J, van der Voort-Doedens LM, de Kok ME, Hobbelt-Stoker MJ (2007) Prevalence of acquired undescended testis in 6-year, 9-year and 13-year-old Dutch schoolboys. *Arch Dis Child* 92:17–20. <https://doi.org/10.1136/adc.2005.076208>
 30. Abeyaratne MR, Aherne WA, Scott JE (1969) The vanishing testis. *Lancet* 2:822–824. [https://doi.org/10.1016/s0140-6736\(69\)92775-2](https://doi.org/10.1016/s0140-6736(69)92775-2)
 31. Lou CC, Lin JN, Tung TC, Wang KL (1994) Anatomical findings of the vanishing testis. *Changgeng Yi Xue Za Zhi* 17:121–124
 32. Merry C, Sweeney B, Puri P (1997) The vanishing testis: anatomical and histological findings. *Eur Urol* 31:65–67. <https://doi.org/10.1159/000474420>
 33. Borrow M, Gough MH (1970) Bilateral absence of testes. *Lancet* 1:366–361. [https://doi.org/10.1016/s0140-6736\(70\)90753-1](https://doi.org/10.1016/s0140-6736(70)90753-1)
 34. Smith NM, Byard RW, Bourne AJ (1991) Testicular regression syndrome—a pathological study of 77 cases. *Histopathology* 19:269–272. <https://doi.org/10.1111/j.1365-2559.1991.tb00033.x>
 35. Barteczko KJ, Jacob MI (2000) The testicular descent in human. Origin, development and fate of the gubernaculum Hunteri, processus vaginalis peritonei, and gonadal ligaments. *Adv Anat Embryol Cell Biol* 156:III–98
 36. Ramareddy RS, Alladi A, Siddappa OS (2013) Ectopic testis in children: experience with seven cases. *J Pediatr Surg* 48:538–541. <https://doi.org/10.1016/j.jpedsurg.2012.10.005>
 37. Rodprasert W, Virtanen HE, Makela J, Toppari J (2020) Hypogonadism and Cryptorchidism. *Front Endocrinol (Lausanne)* 10:906. <https://doi.org/10.3389/fendo.2019.00906>
 38. Bay K, Main KM, Toppari J, Skakkebaek NE (2011) Testicular descent: INSL3, testosterone, genes and the intrauterine milieu. *Nat Rev Urol* 8:187–196. <https://doi.org/10.1038/nrurol.2011.23>
 39. Hutson JM, Hasthorpe S (2005) Testicular descent and cryptorchidism: the state of the art in 2004. *J Pediatr Surg* 40:297–302. <https://doi.org/10.1016/j.jpedsurg.2004.10.033>
 40. Shono T, Ramm-Anderson S, Hutson JM (1994) Transabdominal testicular descent is really ovarian ascent. *J Urol* 152:781–784. [https://doi.org/10.1016/s0022-5347\(17\)32708-8](https://doi.org/10.1016/s0022-5347(17)32708-8)
 41. Zimmermann S, Steding G, Emmen JM, Brinkmann AO, Nayeria K, Holstein AF, Engel W, Adham IM (1999) Targeted disruption of the *Ins13* gene causes bilateral cryptorchidism. *Mol Endocrinol* 13:681–691. <https://doi.org/10.1210/mend.13.5.0272>
 42. Tomiyama H, Hutson JM, Truong A, AgoulNIK AI (2003) Transabdominal testicular descent is disrupted in mice with deletion of insulinlike factor 3 receptor. *J Pediatr Surg* 38:1793–1798. <https://doi.org/10.1016/j.jpedsurg.2003.08.047>
 43. Foresta C, Ferlin A (2004) Role of INSL3 and LGR8 in cryptorchidism and testicular functions. *Reprod Biomed Online* 9:294–298. [https://doi.org/10.1016/s1472-6483\(10\)62144-x](https://doi.org/10.1016/s1472-6483(10)62144-x)
 44. Beltran-Brown F, Villegas-Alvarez F (1988) Clinical classification for undescended testes: experience in 1,010 orchidopexies. *J Pediatr Surg* 23:444–447. [https://doi.org/10.1016/s0022-3468\(88\)80445-7](https://doi.org/10.1016/s0022-3468(88)80445-7)
 45. Boisen KA, Kaleva M, Main KM, Virtanen HE, Haavisto A, Schmidt IM, Chellakooty M, Damgaard IN, Mau C, Reunanen M, Skakkebaek NE, Toppari J (2004) Difference in prevalence of congenital cryptorchidism in infants between two Nordic countries. *Lancet* 363:1264–1269. [https://doi.org/10.1016/S0140-6736\(04\)15998-9](https://doi.org/10.1016/S0140-6736(04)15998-9)
 46. Preiksa RT, Zilaitiene B, Matulevicius V, Skakkebaek NE, Petersen JH, Jorgensen N, Toppari J (2005) Higher than expected prevalence of congenital cryptorchidism in Lithuania: a study of 1204 boys at birth and 1 year follow-up. *Hum Reprod* 20:1928–1932. <https://doi.org/10.1093/humrep/deh887>
 47. Heyns CF (1987) The gubernaculum during testicular descent in the human fetus. *J Anat* 153:93–112
 48. Hutson JM (1986) Testicular feminization: a model for testicular descent in mice and men. *J Pediatr Surg* 21:195–198. [https://doi.org/10.1016/s0022-3468\(86\)80830-2](https://doi.org/10.1016/s0022-3468(86)80830-2)
 49. Aschim EL, Nordenskjold A, Giwercman A, Lundin KB, Ruhayel Y, Haugen TB, Grotmol T, Giwercman YL (2004) Linkage between cryptorchidism, hypospadias, and GGN repeat length in the androgen receptor gene. *J Clin Endocrinol Metab* 89:5105–5109. <https://doi.org/10.1210/jc.2004-0293>
 50. Silva-Ramos M, Oliveira JM, Cabeda JM, Reis A, Soares J, Pimenta A (2006) The CAG repeat within the androgen receptor gene and its relationship to cryptorchidism. *Int Braz J Urol* 32:330–334 discussion 335. <https://doi.org/10.1590/s1677-55382006000300014>
 51. Wiener JS, Marcelli M, Gonzales ETJ, Roth DR, Lamb DJ (1998) Androgen receptor gene alterations are not associated with isolated cryptorchidism. *J Urol* 160:863–865. [https://doi.org/10.1016/s0022-5347\(01\)62823-4](https://doi.org/10.1016/s0022-5347(01)62823-4)
 52. Suzuki Y, Sasagawa I, Ashida J, Nakada T, Muroya K, Ogata T (2001) Screening for mutations of the androgen receptor gene in patients with isolated cryptorchidism. *Fertil Steril* 76:834–836. [https://doi.org/10.1016/s0015-0282\(01\)02016-7](https://doi.org/10.1016/s0015-0282(01)02016-7)
 53. Suzuki Y, Sasagawa I, Itoh K, Ashida J, Ogata T (2002) 5Alpha-reductase type 2 genes in Japanese males do not appear to be associated with cryptorchidism. *Fertil Steril* 78:330–334. [https://doi.org/10.1016/s0015-0282\(02\)03220-x](https://doi.org/10.1016/s0015-0282(02)03220-x)
 54. Elert A, Jahn K, Heidenreich A, Hofmann R (2003) The familial undescended testis. *Klin Padiatr* 215:40–45. <https://doi.org/10.1055/s-2003-36894>
 55. Barthold JS, Pugarelli J, MacDonald ML, Ren J, Adetunji MO, Polson SW, Mateson A, Wang Y, Sol-Church K, McCahan SM, Akins REJ, Devoto M, Robbins AK (2016) Polygenic inheritance of cryptorchidism susceptibility in the LE/orl rat. *Mol Hum Reprod* 22:18–34. <https://doi.org/10.1093/molehr/gav060>
 56. Clarnette TD, Sugita Y, Hutson JM (1997) Genital anomalies in human and animal models reveal the mechanisms and hormones

- governing testicular descent. *Br J Urol* 79:99–112. <https://doi.org/10.1046/j.1464-410x.1997.25622.x>
57. Quinton R, Duke VM, Robertson A, Kirk JM, Matfin G, de Zoysa PA, Azcona C, MacColl GS, Jacobs HS, Conway GS, Besser M, Stanhope RG, Bouloux PM (2001) Idiopathic gonadotrophin deficiency: genetic questions addressed through phenotypic characterization. *Clin Endocrinol (Oxf)* 55:163–174. <https://doi.org/10.1046/j.1365-2265.2001.01277.x>
 58. Bonomi M, Vezzoli V, Krausz C, Guizzardi F, Vezzani S, Simoni M, Bassi I, Duminuco P, Di Iorgi N, Giavoli C, Pizzocaro A, Russo G, Moro M, Fatti L, Ferlin A, Mazzanti L, Zatelli MC, Cannavo S, Isidori AM, Pincelli AI, Prodam F, Mancini A, Limone P, Tanda ML, Gaudino R, Salerno M, Francesca P, Maghnie M, Maggi M, Persani L, on Central Hypogonadism (NICe group) (2018) Italian Network on Central Hypogonadism, Italian Network Characteristics of a nationwide cohort of patients presenting with isolated hypogonadotropic hypogonadism (IHH). *Eur J Endocrinol* 178:23–32. <https://doi.org/10.1530/EJE-17-0065>
 59. Main KM, Schmidt IM, Skakkebaek NE (2000) A possible role for reproductive hormones in newborn boys: progressive hypogonadism without the postnatal testosterone peak. *J Clin Endocrinol Metab* 85:4905–4907. <https://doi.org/10.1210/jcem.85.12.7058>
 60. Radmayr C, Dogan HS, Hoebeke P, Kocvara R, Nijman R, Silay S, Stein R, Undre S, Tekgul S (2016) Management of undescended testes: European Association of Urology/European Society for Paediatric Urology Guidelines. *J Pediatr Urol* 12:335–343. <https://doi.org/10.1016/j.jpuro.2016.07.014>
 61. Smolko MJ, Kaplan GW, Brock WA (1983) Location and fate of the nonpalpable testis in children. *J Urol* 129:1204–1206. [https://doi.org/10.1016/s0022-5347\(17\)52643-9](https://doi.org/10.1016/s0022-5347(17)52643-9)
 62. Mau EE, Leonard MP (2017) Practical approach to evaluating testicular status in infants and children. *Can Fam Physician* 63:432–435
 63. Saylors S, Oyetunji TA (2024) Management of undescended testis. *Curr Opin Pediatr* 36:554–561. <https://doi.org/10.1097/MOP.0000000000001387>
 64. Cendron M, Huff DS, Keating MA, Snyder HM, Duckett JW (1993) Anatomical, morphological and volumetric analysis: a review of 759 cases of testicular maldescent. *J Urol* 149:570–573. [https://doi.org/10.1016/s0022-5347\(17\)36151-7](https://doi.org/10.1016/s0022-5347(17)36151-7)
 65. Tasian GE, Zaid H, Cabana MD, Baskin LS (2010) Proximal hypospadias and risk of acquired cryptorchidism. *J Urol* 184:715–720. <https://doi.org/10.1016/j.juro.2010.03.056>
 66. Stec AA, Thomas JC, DeMarco RT, Pope JC, Brock JW, Adams MC (2007) Incidence of testicular ascent in boys with retractile testes. *J Urol* 178:1722–1725. <https://doi.org/10.1016/j.juro.2007.05.091>
 67. Baird R, Gholoum S, Laberge J, Puligandla P (2011) Prematurity, not age at operation or incarceration, impacts complication rates of inguinal hernia repair. *J Pediatr Surg* 46:908–911. <https://doi.org/10.1016/j.jpedsurg.2011.02.059>
 68. Tasian GE, Copp HL (2011) Diagnostic performance of ultrasound in nonpalpable cryptorchidism: a systematic review and meta-analysis. *Pediatrics* 127:119–128. <https://doi.org/10.1542/peds.2010-1800>
 69. Kanemoto K, Hayashi Y, Kojima Y, Maruyama T, Ito M, Kohri K (2005) Accuracy of ultrasonography and magnetic resonance imaging in the diagnosis of non-palpable testis. *Int J Urol* 12:668–672. <https://doi.org/10.1111/j.1442-2042.2005.01102.x>
 70. Krishnaswami S, Fonnesbeck C, Penson D, McPheeters ML (2013) Magnetic resonance imaging for locating nonpalpable undescended testicles: a meta-analysis. *Pediatrics* 131:1908. <https://doi.org/10.1542/peds.2013-0073>
 71. Grumbach MM (2005) A window of opportunity: the diagnosis of gonadotropin deficiency in the male infant. *J Clin Endocrinol Metab* 90:3122–3127. <https://doi.org/10.1210/jc.2004-2465>
 72. Ibba A, Del Pistoia M, Balsamo A, Baronio F, Capalbo D, Russo G, DE Sanctis L, Bizzarri C (2021) Differences of sex development in the newborn: from clinical scenario to molecular diagnosis. *Minerva Pediatr (Torino)* 73:606–620. <https://doi.org/10.23736/S2724-5276.21.06512-5>
 73. White PC (2009) Neonatal screening for congenital adrenal hyperplasia. *Nat Rev Endocrinol* 5:490–498. <https://doi.org/10.1038/nrendo.2009.148>
 74. Brauner R, Neve M, Allali S, Trivin C, Lottmann H, Bashamboo A, McElreavey K (2011) Clinical, biological and genetic analysis of anorchia in 26 boys. *PLoS ONE* 6:e23292. <https://doi.org/10.1371/journal.pone.0023292>
 75. Kalfa N, Gaspari L, Ollivier M, Philibert P, Bergougnoux A, Paris F, Sultan C (2019) Molecular genetics of hypospadias and cryptorchidism recent developments. *Clin Genet* 95:122–131. <https://doi.org/10.1111/cge.13432>
 76. Moreno-Garcia M, Miranda EB (2002) Chromosomal anomalies in cryptorchidism and hypospadias. *J Urol* 168:2170–2172 discussion 2172. [https://doi.org/10.1016/S0022-5347\(05\)64346-7](https://doi.org/10.1016/S0022-5347(05)64346-7)
 77. Gates RL, Shelton J, Diefenbach KA, Arnold M, St Peter SD, Renaud EJ, Slidell MB, Somme S, Valusek P, Villalona GA, McAteer JP, Beres AL, Baerg J, Rentea RM, Kelley-Quon L, Kawaguchi AL, Hu Y, Miniati D, Ricca R, Baird R (2022) Management of the undescended testis in children: An American Pediatric Surgical Association Outcomes and Evidence Based Practice Committee Systematic Review. *J Pediatr Surg* 57:1293–1308. <https://doi.org/10.1016/j.jpedsurg.2022.01.003>
 78. Kuiri-Hanninen T, Sankilampi U, Dunkel L (2014) Activation of the hypothalamic-pituitary-gonadal axis in infancy: minipuberty. *Horm Res Paediatr* 82:73–80. <https://doi.org/10.1159/000362414>
 79. Bizzarri C, Cappa M (2020) Ontogeny of Hypothalamus-Pituitary Gonadal Axis and Minipuberty: An Ongoing Debate? *Front Endocrinol (Lausanne)* 11:187. <https://doi.org/10.3389/fendo.2020.00187>
 80. Winter JS, Taraska S, Faïman C (1972) The hormonal response to HCG stimulation in male children and adolescents. *J Clin Endocrinol Metab* 34:348–353. <https://doi.org/10.1210/jcem-34-2-348>
 81. Rey R (2003) How to evaluate gonadal function in the cryptorchid boy. Lessons from new testicular markers. *J Pediatr Endocrinol Metab* 16:357–364. <https://doi.org/10.1515/jpem.2003.16.3.357>
 82. Lee PA, Houk CP (2013) Cryptorchidism. *Curr Opin Endocrinol Diabetes Obes* 20:210–216. <https://doi.org/10.1097/MED.0b013e32835ffc7d>
 83. Allin BSR, Dumann E, Fawcner-Corbett D, Kwok C, Skerrett C, Paediatric Surgery Trainees Research Network (2018) Systematic review and meta-analysis comparing outcomes following orchidopexy for cryptorchidism before or after 1 year of age. *BJS Open* 2:1–12. <https://doi.org/10.1002/bjs5.36>
 84. Kollin C, Stukenborg JB, Nurmio M, Sundqvist E, Gustafsson T, Soder O, Toppari J, Nordenskjold A, Ritzen EM (2012) Boys with undescended testes: endocrine, volumetric and morphometric studies on testicular function before and after orchidopexy at nine months or three years of age. *J Clin Endocrinol Metab* 97:4588–4595. <https://doi.org/10.1210/jc.2012-2325>
 85. Rodprasert W, Virtanen HE, Toppari J (2024) Cryptorchidism and puberty. *Front Endocrinol (Lausanne)* 15:1347435. <https://doi.org/10.3389/fendo.2024.1347435>
 86. Tasian GE, Hittelman AB, Kim GE, DiSandro MJ, Baskin LS (2009) Age at orchiopexy and testis palpability predict germ and Leydig cell loss: clinical predictors of adverse histological features of cryptorchidism. *J Urol* 182:704–709. <https://doi.org/10.1016/j.juro.2009.04.032>
 87. Rusnack SL, Wu H, Huff DS, Snyder HM, Zderic SA, Carr MC, Canning DA (2002) The ascending testis and the testis undescended since birth share the same histopathology. *J Urol*

- 168:2590–2591. [https://doi.org/10.1016/S0022-5347\(05\)64223-1](https://doi.org/10.1016/S0022-5347(05)64223-1)
88. Elder JS (2016) Surgical Management of the Undescended Testis: Recent Advances and Controversies. *Eur J Pediatr Surg* 26:418–426. <https://doi.org/10.1055/s-0036-1592197>
 89. Johnson DE, Woodhead DM, Pohl DR, Robison JR (1968) Cryptorchidism and testicular tumorigenesis. *Surgery* 63:919–922
 90. Lip SZL, Murchison LED, Cullis PS, Govan L, Carachi R (2013) A meta-analysis of the risk of boys with isolated cryptorchidism developing testicular cancer in later life. *Arch Dis Child* 98:20–26. <https://doi.org/10.1136/archdischild-2012-302051>
 91. Cools M, Drop SLS, Wolffenbuttel KP, Oosterhuis JW, Looijenga LHJ (2006) Germ Cell Tumors in the Intersex Gonad: Old Paths, New Directions, Moving Frontiers. *Endocr Rev* 27:468–484. <http://doi.org/10.1210/er.2006-0005>
 92. Elsherbeny MS, Abdelhay S (2019) Human chorionic gonadotropin hormone for treatment of congenital undescended testis: Anatomical barriers to its success. *J Pediatr Surg* 54:2413–2415. <https://doi.org/10.1016/j.jpedsurg.2019.01.067>
 93. Bogaert G, Vanhoyland M, Hadziselimovic F (2023) Low-dose every-second-day LHRH treatment following bilateral orchidopexy in children with bilateral cryptorchidism may improve their fertility outcome. *J Pediatr Urol* 19:128e1–128e7. <https://doi.org/10.1016/j.jpuro.2022.10.017>
 94. Kolon TF, Herndon CDA, Baker LA, Baskin LS, Baxter CG, Cheng EY, Diaz M, Lee PA, Seashore CJ, Tasian GE, Barthold JS, American Urological Association (2014) Evaluation and treatment of cryptorchidism: AUA guideline. *J Urol* 192:337–345. <http://doi.org/10.1016/j.juro.2014.05.005>
 95. Wei Y, Wang Y, Tang X, Liu B, Shen L, Long C, Lin T, He D, Wu S, Wei G (2018) Efficacy and safety of human chorionic gonadotropin for treatment of cryptorchidism: A meta-analysis of randomised controlled trials. *J Paediatr Child Health* 54:900–906. <https://doi.org/10.1111/jpc.13920>
 96. Pitteloud N, Hayes FJ, Dwyer A, Boepple PA, Lee H, Crowley WFJ (2002) Predictors of outcome of long-term GnRH therapy in men with idiopathic hypogonadotropic hypogonadism. *J Clin Endocrinol Metab* 87:4128–4136. <https://doi.org/10.1210/jc.2002-020518>
 97. Cangiano B, Goggi G, Federici S, Bresesti C, Cotellessa L, Guizzardi F, Vezzoli V, Duminuco P, Persani L, Bonomi M (2021) Predictors of reproductive and non-reproductive outcomes of gonadotropin mediated pubertal induction in male patients with congenital hypogonadotropic hypogonadism (CHH). *J Endocrinol Invest* 44:2445–2454. <https://doi.org/10.1007/s40618-021-01556-x>
 98. Lambert A, Bougneres P (2016) Growth and descent of the testes in infants with hypogonadotropic hypogonadism receiving subcutaneous gonadotropin infusion. *Int J Pediatr Endocrinol* 2016:13–19 Epub 2016 Jul 4. <https://doi.org/10.1186/s13633-016-0031-9>
 99. Papadimitriou DT, Chrysis D, Nyktari G, Zoupanos G, Liakou E, Papadimitriou A, Mastorakos G (2019) Replacement of Male Mini-Puberty. *J Endocr Soc* 3:1275–1282. <https://doi.org/10.1210/je.2019-00083>
 100. Wei Y, Wang Y, Tang X, Liu B, Shen L, Long C, Lin T, He D, Wu S, Wei G (2018) Efficacy and safety of human chorionic gonadotropin for treatment of cryptorchidism: A meta-analysis of randomised controlled trials. *J Paediatr Child Health* 54:900–906. <https://doi.org/10.1111/jpc.13920>
 101. Bu Q, Pan Z, Jiang S, Wang A, Cheng H (2016) The Effectiveness of hCG and LHRH in Boys with Cryptorchidism: A Meta-Analysis of Randomized Controlled Trials. *Horm Metab Res* 48:318–324. <https://doi.org/10.1055/s-0042-104059>
 102. Florou M, Tsilidis KK, Siomou E, Koletsis T, Symioti A, Spyridakis I, Kaselas C, Ntzani EE (2023) Orchidopexy for congenital cryptorchidism in childhood and adolescence and testicular cancer in adults: an updated systematic review and meta-analysis of observational studies. *Eur J Pediatr* 182:2499–2507. <https://doi.org/10.1007/s00431-023-04947-9>
 103. Hensle TW, Deibert CM (2012) Adult male health risks associated with congenital abnormalities. *Urol Clin North Am* 39:109–114. <https://doi.org/10.1016/j.ucl.2011.09.002>
 104. Woodhouse CRJ, Lipshultz L, Hwang K, Mouriquand P, Creighton S (2012) Adult care of children from pediatric urology: part 2. *J Urol* 188:717–723. <https://doi.org/10.1016/j.juro.2012.05.001>
 105. Pakkasjarvi N, Taskinen S (2024) Surgical treatment of cryptorchidism: current insights and future directions. *Front Endocrinol (Lausanne)* 15:1327957. <https://doi.org/10.3389/fendo.2024.1327957>
 106. Cools M, Nordenstrom A, Robeva R, Hall J, Westerveld P, Fluck C, Kohler B, Berra M, Springer A, Schweizer K, Pastorski V, COST Action BM1303 working group 1 (2018) Caring for individuals with a difference of sex development (DSD): a Consensus Statement. *Nat Rev Endocrinol* 14:415–429. <https://doi.org/10.1038/s41574-018-0010-8>
 107. Kregel S, Eckoldt F, Richter-Unruh A, Kohler B, Leuschner I, Mentzel H, Moss A, Schweizer K, Stein R, Werner-Rosen K, Wieacker P, Wiesemann C, Wunsch L, Richter-Appelt H (2019) Variations of sex development: The first German interdisciplinary consensus paper. *J Pediatr Urol* 15:114–123. <https://doi.org/10.1016/j.jpuro.2018.10.008>
 108. Bever Yv, Bruggenwirth HT, Wolffenbuttel KP, Dessens AB, Groenenberg IAL, Knapen MFCM, De Baere E, Cools M, van Ravenswaaij-Arts CMA, Sikkema-Raddatz B, van der Laahsen-H, Kempers M, Rinne T, Hersmus R, Looijenga L, Hannema SE (2020) Under-reported aspects of diagnosis and treatment addressed in the Dutch-Flemish guideline for comprehensive diagnostics in disorders/differences of sex development. *J Med Genet* 57:581–589. <https://doi.org/10.1136/jmedgenet-2019-106354>
 109. Bennecke E, Bernstein S, Lee P, van de Grift TC, Nordenskjold A, Rapp M, Simmonds M, Streuli JC, Thyen U, Wiesemann C, dsd-LIFE Group (2021) Early Genital Surgery in Disorders/Differences of Sex Development: Patients' Perspectives. *Arch Sex Behav* 50:913–923. <https://doi.org/10.1007/s10508-021-01953-6>
 110. Ahmed SF, Achermann J, Alderson J, Crouch NS, Elford S, Hughes IA, Krone N, McGowan R, Mushtaq T, O'Toole S, Perry L, Rodie ME, Skae M, Turner HE (2021) Society for Endocrinology UK Guidance on the initial evaluation of a suspected difference or disorder of sex development (Revised 2021). *Clin Endocrinol (Oxf)* 95:818–840. <https://doi.org/10.1111/cen.14528>
 111. Jurgensen M, Rapp M, Dohnert U, Frielitz F, Ahmed F, Cools M, Thyen U, Hiort O (2021) Assessing the health-related management of people with differences of sex development. *Endocrine* 71:675–680. <https://doi.org/10.1007/s12020-021-02627-y>
 112. Kavanaugh GL, Mohnach L, Youngblom J, Kellison JG, Sandberg DE, Accord Alliance (2021) Good practices in pediatric clinical care for disorders/differences of sex development. *Endocrine* 73:723–733. <https://doi.org/10.1007/s12020-021-02748-4>
 113. Wechsung K, Marshall L, Jurgensen M, Neumann U, On Behalf Of The Empower-Dsd Study Group (2022) Diagnosis of DSD in Children-Development of New Tools for a Structured Diagnostic and Information Management Program within the Empower-DSD Study. *J Clin Med* 11:3859. <https://doi.org/10.3390/jcm1133859>
 114. Gardner M, Khorashad BS, Lee PA, Kogan BA, Sandberg DE (2024) Recommendations for 46,XX Congenital Adrenal Hyperplasia Across Two Decades: Insights from the North American Differences of Sex Development Clinician Survey. *Arch Sex Behav* 53:1695–1711. <https://doi.org/10.1007/s10508-024-02853-1>

115. Khorashad BS, Gardner M, Lee PA, Kogan BA, Sandberg DE (2024) Recommendations for 46,XY Disorders/Differences of Sex Development Across Two Decades: Insights from North American Pediatric Endocrinologists and Urologists. *Arch Sex Behav* 53:2939–2956. <https://doi.org/10.1007/s10508-024-02942-1>
116. Lee PA, Houk CP, Ahmed SF, Hughes IA, International Consensus Conference on Intersex organized by the Lawson Wilkins Pediatric Endocrine Society and the European Society for Paediatric Endocrinology (2006) Consensus statement on management of intersex disorders. *International Consensus Conference on Intersex. Pediatrics* 118:488. <https://doi.org/10.1542/peds.2006-0738>
117. Ahmed SF, Khwaja O, Hughes IA (2000) The role of a clinical score in the assessment of ambiguous genitalia. *BJU Int* 85:120–124. <https://doi.org/10.1046/j.1464-410x.2000.00354.x>
118. Hiort O, Birnbaum W, Marshall L, Wunsch L, Werner R, Schroder T, Dohnert U, Holterhus P (2014) Management of disorders of sex development. *Nat Rev Endocrinol* 10:520–529. <https://doi.org/10.1038/nrendo.2014.108>
119. Del Valle I, Buonocore F, Duncan AJ, Lin L, Barenco M, Parinaik R, Shah S, Hubank M, Gerrelli D, Achermann JC (2017) A genomic atlas of human adrenal and gonad development. *Wellcome Open Res* 2:25. <https://doi.org/10.12688/wellcomeopenres.11253.2>
120. Haqq CM, Donahoe PK (1998) Regulation of sexual dimorphism in mammals. *Physiol Rev* 78:1–33. <https://doi.org/10.1152/physrev.1998.78.1.1>
121. MacLaughlin DT, Donahoe PK (2004) Sex determination and differentiation. *N Engl J Med* 350:367–378. <https://doi.org/10.1056/NEJMr022784>
122. Harley VR, Clarkson MJ, Argentaro A (2003) The molecular action and regulation of the testis-determining factors, SRY (sex-determining region on the Y chromosome) and SOX9 [SRY-related high-mobility group (HMG) box 9]. *Endocr Rev* 24:466–487. <https://doi.org/10.1210/er.2002-0025>
123. Vetro A, Dehghani MR, Kraoua L, Giorda R, Beri S, Cardarelli L, Merico M, Manolagos E, Parada-Bustamante A, Castro A, Radi O, Camerino G, Brusco A, Sabaghian M, Sofocleous C, Forzano F, Palumbo P, Palumbo O, Calvano S, Zelante L, Grammatico P, Giglio S, Basly M, Chaabouni M, Carella M, Russo G, Bonaglia MC, Zuffardi O (2015) Testis development in the absence of SRY: chromosomal rearrangements at SOX9 and SOX3. *Eur J Hum Genet* 23:1025–1032. <https://doi.org/10.1038/ejhg.2014.237>
124. Joss N, Rey RA (2020) What Does AMH Tell Us in Pediatric Disorders of Sex Development? *Front Endocrinol (Lausanne)* 11:619. <https://doi.org/10.3389/fendo.2020.00619>
125. Batista RL, Mendonca BB (2022) The Molecular Basis of 5alpha-Reductase Type 2 Deficiency. *Sex Dev* 16:171–183. <https://doi.org/10.1159/000525119>
126. Crouch NS, Creighton SM (2014) Transition of care for adolescents with disorders of sex development. *Nat Rev Endocrinol* 10:436–442. <https://doi.org/10.1038/nrendo.2014.62>
127. Hullmann SE, Chalmers LJ, Wisniewski AB (2012) Transition from pediatric to adult care for adolescents and young adults with a disorder of sex development. *J Pediatr Adolesc Gynecol* 25:155–157. <https://doi.org/10.1016/j.jpag.2011.11.003>
128. van der Straaten S, Springer A, Zecic A, Hebenstreit D, Tonnhofer U, Gawlik A, Baumert M, Szeliga K, Debulpaep S, Desloovere A, Tack L, Smets K, Wasniewska M, Corica D, Calafiore M, Ljubicic ML, Busch AS, Juul A, Nordenstrom A, Sigurdsson J, Fluck CE, Haamberg T, Graf S, Hannema SE, Wolfenbutter KP, Hiort O, Ahmed SF, Cools M (2020) The External Genitalia Score (EGS): A European Multicenter Validation Study. *J Clin Endocrinol Metab* 105:dgz142. <https://doi.org/10.1210/clinem/dgz142>
129. Thankamony A, Pasterski V, Ong KK, Acerini CL, Hughes IA (2016) Anogenital distance as a marker of androgen exposure in humans. *Andrology* 4:616–625. <https://doi.org/10.1111/andr.12156>
130. Kulle A, Krone N, Holterhus PM, Schuler G, Greaves RF, Juul A, de Rijke YB, Hartmann MF, Saba A, Hiort O, Wudy SA, EU COST Action (2017) Steroid hormone analysis in diagnosis and treatment of DSD: position paper of EU COST Action BM 1303 ‘DSDnet’. *Eur J Endocrinol* 176:P1–P9. <https://doi.org/10.1530/EJE-16-0953>
131. Rosner W, Vesper H, Pediatric Endocrine Society (2010) Toward excellence in testosterone testing: a consensus statement. *J Clin Endocrinol Metab* 95:4542–4548. <https://doi.org/10.1210/jc.2010-1314>
132. Brunello FG, Rey RA (2022) AMH and AMHR2 Involvement in Congenital Disorders of Sex Development. *Sex Dev* 16:138–146. <https://doi.org/10.1159/000518273>
133. Johannsen TH, Main KM, Ljubicic ML, Jensen TK, Andersen HR, Andersen MS, Petersen JH, Andersson A, Juul A (2018) Sex Differences in Reproductive Hormones During Mini-Puberty in Infants With Normal and Disordered Sex Development. *J Clin Endocrinol Metab* 103:3028–3037. <https://doi.org/10.1210/jc.2018-00482>
134. Alaniz VI, Kobernik EK, Dillman J, Quint EH (2016) Utility of Ultrasound and Magnetic Resonance Imaging in Patients with Disorders of Sex Development Who Undergo Prophylactic Gonadectomy. *J Pediatr Adolesc Gynecol* 29:577–581. <https://doi.org/10.1016/j.jpag.2016.03.007>
135. Riccabona M, Darge K, Lobo M, Ording-Muller L, Augdal TA, Avni FE, Blickman J, Damasio BM, Ntoulia A, Papadopoulou F, Vivier P, Willi U (2015) ESPR Uroradiology Taskforce—imaging recommendations in paediatric uroradiology, part VIII: retrograde urethrography, imaging disorder of sexual development and imaging childhood testicular torsion. *Pediatr Radiol* 45:2023–2028. <https://doi.org/10.1007/s00247-015-3452-3>
136. Audi L, Ahmed SF, Krone N, Cools M, McElreavey K, Holterhus PM, Greenfield A, Bashamboo A, Hiort O, Wudy SA, McGowan R, The EU COST Action (2018) GENETICS IN ENDOCRINOLOGY: Approaches to molecular genetic diagnosis in the management of differences/disorders of sex development (DSD): position paper of EU COST Action BM 1303 ‘DSDnet’. *Eur J Endocrinol* 179:R197–R206. <https://doi.org/10.1530/EJE-18-0256>
137. Persani L, Cools M, Ioakim S, Faisal Ahmed S, Andonova S, Avbelj-Stefanija M, Baronio F, Bouligand J, Bruggenwirth HT, Davies JH, De Baere E, Dzivite-Krisane I, Fernandez-Alvarez P, Gheldof A, Giavoli C, Gravholt CH, Hiort O, Holterhus P, Juul A, Krausz C, Lagerstedt-Robinson K, McGowan R, Neumann U, Novelli A, Peyrassol X, Phylactou LA, Rohayem J, Touraine P, Westra D, Vezzoli V, Rossetti R (2022) The genetic diagnosis of rare endocrine disorders of sex development and maturation: a survey among Endo-ERN centres. *Endocr Connect* 11:e220367. doi: 10.1530/EC-0367. Print 2022 Dec 1. <https://doi.org/10.1530/EC-22-0367>
138. O’Connell MA, Atlas G, Ayers K, Sinclair A (2023) Establishing a Molecular Genetic Diagnosis in Children with Differences of Sex Development: A Clinical Approach. *Horm Res Paediatr* 96:128–143. <https://doi.org/10.1159/000520926>
139. Ahmed SF, Alimusina M, Batista RL, Domenice S, Lisboa Gomes N, McGowan R, Patjamontri S, Mendonca BB (2022) The Use of Genetics for Reaching a Diagnosis in XY DSD. *Sex Dev* 16:207–224. <https://doi.org/10.1159/000524881>
140. D’Alborton F, Assante MT, Foresti M, Balsamo A, Bertelloni S, Dati E, Nardi L, Bacchi ML, Mazzanti L (2015) Quality of Life and Psychological Adjustment of Women Living with 46,XY Differences of Sex Development. *J Sex Med* 12:1440–1449. <https://doi.org/10.1111/jsm.12884>

141. Jurgensen M, Kleinemeier E, Lux A, Steensma TD, Cohen-Kettenis PT, Hiort O, Thyen U, Kohler B, DSD Network Working Group (2013) Psychosexual development in adolescents and adults with disorders of sex development—results from the German Clinical Evaluation Study. *J Sex Med* 10:2703–2714. <https://doi.org/10.1111/j.1743-6109.2012.02751.x>
142. Schweizer K, Brunner F, Gedrose B, Handford C, Richter-Appelt H (2017) Coping With Diverse Sex Development: Treatment Experiences and Psychosocial Support During Childhood and Adolescence and Adult Well-Being. *J Pediatr Psychol* 42:504–519. <https://doi.org/10.1093/jpepsy/jsw058>
143. Nordenstrom A, Thyen U (2014) Improving the communication of healthcare professionals with affected children and adolescents. *Endocr Dev* 27:113–127. <https://doi.org/10.1159/000363636>
144. Lee PA, Houk CP (2010) The role of support groups, advocacy groups, and other interested parties in improving the care of patients with congenital adrenal hyperplasia: pleas and warnings. *Int J Pediatr Endocrinol* 2010:563640. <https://doi.org/10.1155/2010/563640>
145. Baratz AB, Sharp MK, Sandberg DE (2014) Disorders of sex development peer support. *Endocr Dev* 27:99–112. <https://doi.org/10.1159/000363634>
146. Alimussina M, Diver LA, McGowan R, Ahmed SF (2018) Genetic testing of XY newborns with a suspected disorder of sex development. *Curr Opin Pediatr* 30:548–557. <https://doi.org/10.1097/MOP.0000000000000644>
147. Leon NY, Reyes AP, Harley VR (2019) A clinical algorithm to diagnose differences of sex development. *Lancet Diabetes Endocrinol* 7:560–574. [https://doi.org/10.1016/S2213-8587\(18\)30339-5](https://doi.org/10.1016/S2213-8587(18)30339-5)
148. Fukami M, Ogata T (2014) Cytochrome P450 oxidoreductase deficiency: rare congenital disorder leading to skeletal malformations and steroidogenic defects. *Pediatr Int* 56:805–808. <https://doi.org/10.1111/ped.12518>
149. Siminoff LA, Sandberg DE (2015) Promoting Shared Decision Making in Disorders of Sex Development (DSD): Decision Aids and Support Tools. *Horm Metab Res* 47:335–339. <https://doi.org/10.1055/s-0035-1545302>
150. Finkelstein GP, Vieites A, Bergada I, Rey RA (2021) Disorders of Sex Development of Adrenal Origin. *Front Endocrinol (Lausanne)* 12:770782. <https://doi.org/10.3389/fendo.2021.770782>
151. Birnbaum W, Bertelloni S (2014) Sex hormone replacement in disorders of sex development. *Endocr Dev* 27:149–159. <https://doi.org/10.1159/000363640>
152. Nordenstrom A, Ahmed SF, van den Akker E, Blair J, Bonomi M, Brachet C, Broersen LHA, Claahsen-van der Grinten HL, Dessens AB, Gawlik A, Gravholt CH, Juul A, Krausz C, Raivio T, Smyth A, Touraine P, Vitali D, Dekkers OM (2022) Pubertal induction and transition to adult sex hormone replacement in patients with congenital pituitary or gonadal reproductive hormone deficiency: an Endo-ERN clinical practice guideline. *Eur J Endocrinol* 186:G9–G49. <https://doi.org/10.1530/EJE-22-0073>
153. Hembree WC, Cohen-Kettenis PT, Gooren L, Hannema SE, Meyer WJ, Murad MH, Rosenthal SM, Safer JD, Tangpricha V, T'Sjoen GG (2017) Endocrine Treatment of Gender-Dysphoric/Gender-Incongruent Persons: An Endocrine Society Clinical Practice Guideline. *J Clin Endocrinol Metab* 102:3869–3903. <https://doi.org/10.1210/jc.2017-01658>
154. Rosenthal SM (2021) Challenges in the care of transgender and gender-diverse youth: an endocrinologist's view. *Nat Rev Endocrinol* 17:581–591. <https://doi.org/10.1038/s41574-021-00535-9>
155. Coleman E, Radix AE, Bouman WP, Brown GR, de Vries ALC, Deutsch MB, Ettner R, Fraser L, Goodman M, Green J, Hancock AB, Johnson TW, Karasic DH, Knudson GA, Leibowitz SF, Meyer-Bahlburg HFL, Monstrey SJ, Motmans J, Nahata L, Nieder TO, Reisner SL, Richards C, Schechter LS, Tangpricha V, Tishelman AC, Van Trotsenburg MAA, Winter S, Ducheny K, Adams NJ, Adrian TM, Allen LR, Azul D, Bagga H, Basar K, Bathory DS, Belinky JJ, Berg DR, Berli JU, Bluebond-Langner RO, Bouman M, Bowers ML, Brassard PJ, Byrne J, Capitan L, Cargill CJ, Carswell JM, Chang SC, Chelvakumar G, Corneil T, Dalke KB, De Cuypere G, de Vries E, Den Heijer M, Devor AH, Dhejne C, D'Marco A, Edmiston EK, Edwards-Leeper L, Ehrbar R, Ehrensaft D, Einfeld J, Elaut E, Erickson-Schroth L, Feldman JL, Fisher AD, Garcia MM, Gijs L, Green SE, Hall BP, Hardy TLD, Irwig MS, Jacobs LA, Janssen AC, Johnson K, Klink DT, Kreukels BPC, Kuper LE, Kvach EJ, Malouf MA, Massey R, Mazur T, McLachlan C, Morrison SD, Mosser SW, Neira PM, Nygren U, Oates JM, Obedin-Maliver J, Pagkalos G, Patton J, Phanuphak N, Rachlin K, Reed T, Rider GN, Ristori J, Robbins-Cherry S, Roberts SA, Rodriguez-Wallberg KA, Rosenthal SM, Sabir K, Safer JD, Scheim AI, Seal LJ, Schoole TJ, Spencer K, St Amand C, Steensma TD, Strang JF, Taylor GB, Tilleman K, T'Sjoen GG, Vala LN, Van Mello NM, Veale JF, Vercill JA, Vincent B, Wesp LM, West MA, Arcelus J (2022) Standards of Care for the Health of Transgender and Gender Diverse People, Version 8. *Int J Transgend Health* 23:S1–S259. <https://doi.org/10.1080/26895269.2022.2100644>
156. Ludvigsson JF, Adolfsson J, Hoistad M, Rydelius P, Kristrom B, Landen M (2023) A systematic review of hormone treatment for children with gender dysphoria and recommendations for research. *Acta Paediatr* 112:2279–2292. <https://doi.org/10.1111/apa.16791>
157. Calcaterra V, Tornese G, Zuccotti G, Staiano A, Cherubini V, Gaudino R, Fazzi EM, Barbi E, Chiarelli F, Corsello G, Esposito SMR, Ferrara P, Iughetti L, Laforgia N, Maghnie M, Marseglia G, Perilongo G, Pettoello-Mantovani M, Ruggieri M, Russo G, Salerno M, Striano P, Valerio G, Wasniewska M, Italian Academy of Pediatrics, Italian Society of Pediatrics (2024) Adolescent gender dysphoria management: position paper from the Italian Academy of Pediatrics, the Italian Society of Pediatrics, the Italian Society for Pediatric Endocrinology and Diabetes, the Italian Society of Adolescent Medicine and the Italian Society of Child and Adolescent Neuropsychiatry. *Ital J Pediatr* 50:73–77. <https://doi.org/10.1186/52-024-01644-7>
158. Kohler B, Kleinemeier E, Lux A, Hiort O, Gruters A, Thyen U, DSD Network Working Group (2012) Satisfaction with genital surgery and sexual life of adults with XY disorders of sex development: results from the German clinical evaluation study. *J Clin Endocrinol Metab* 97:577–588. <https://doi.org/10.1210/jc.2011-1441>
159. Sircili MHP, e Silva FAdQ, Costa EMF, Brito VN, Arnhold IJP, Denes FT, Inacio M, de Mendonca BB (2010) Long-term surgical outcome of masculinizing genitoplasty in large cohort of patients with disorders of sex development. *J Urol* 184:1122–1127. <https://doi.org/10.1016/j.juro.2010.05.022>
160. Minto CL, Liao L, Woodhouse CRJ, Ransley PG, Creighton SM (2003) The effect of clitoral surgery on sexual outcome in individuals who have intersex conditions with ambiguous genitalia: a cross-sectional study. *Lancet* 361:1252–1257. [https://doi.org/10.1016/S0140-6736\(03\)12980-7](https://doi.org/10.1016/S0140-6736(03)12980-7)
161. Cools M, Verhagen E, Hoebeke P, Van Hoecke E, Cannoot P (2024) Working towards convergence of the clinical management of differences of sex development/intersex conditions and the human rights framework: A case study. *Clin Endocrinol (Oxf)* 101:499–506. <https://doi.org/10.1111/cen.15002>
162. Magritte E, Williams J, Amyot E, Usipuk M, Sanders C (2023) Listening to Individuals with Differences in Sex Development or Intersex and Their Families: Not Doing Surgery Does Not Mean Doing Nothing. *Horm Res Paediatr* 96:228–237. <https://doi.org/10.1159/000525452>

163. Meyer-Bahlburg HFL (2022) The Timing of Genital Surgery in Somatic Intersexuality: Surveys of Patients' Preferences. *Horm Res Paediatr* 95:12–20. <https://doi.org/10.1159/000521958>
164. Krege S, Falhammar H, Lax H, Roehle R, Claahsen-van der Grinten H, Kortmann B, Duranteau L, Nordenskjöld A, dsd-LIFE group (2022) Long-Term Results of Surgical Treatment and Patient-Reported Outcomes in Congenital Adrenal Hyperplasia-A Multicenter European Registry Study. *J Clin Med* 11:4629. <https://doi.org/10.3390/jcm11154629>
165. Mouriquand PDE, Gorduz DB, Gay C, Meyer-Bahlburg HFL, Baker L, Baskin LS, Bouvattier C, Braga LH, Caldamone AC, Duranteau L, El Ghoneimi A, Hensle TW, Hoebeke P, Kaefer M, Kalfa N, Kolon TF, Manzoni G, Mure P, Nordenskjöld A, Pippi Salle JL, Poppas DP, Ransley PG, Rink RC, Rodrigo R, Sann L, Schober J, Sibai H, Wisniewski A, Wolfenbutter KP, Lee P (2016) Surgery in disorders of sex development (DSD) with a gender issue: If (why), when, and how? *J Pediatr Urol* 12:139–149. <https://doi.org/10.1016/j.jpuro.2016.04.001>
166. Mouriquand P, Caldamone A, Malone P, Frank JD, Hoebeke P (2014) The ESPU/SPU standpoint on the surgical management of Disorders of Sex Development (DSD). *J Pediatr Urol* 10:8–10. <https://doi.org/10.1016/j.jpuro.2013.10.023>
167. Wolfenbutter KP, Crouch NS (2014) Timing of feminising surgery in disorders of sex development. *Endocr Dev* 27:210–221. <https://doi.org/10.1159/000363665>
168. Looijenga LHJ, Hersmus R, de Leeuw, Bertie H, C G M, Stoop H, Cools M, Oosterhuis JW, Drop SLS, Wolfenbutter KP (2010) Gonadal tumours and DSD. *Best Pract Res Clin Endocrinol Metab* 24:291–310. <https://doi.org/10.1016/j.beem.2009.10.002>
169. Hersmus R, van Bever Y, Wolfenbutter KP, Biermann K, Cools M, Looijenga LHJ (2017) The biology of germ cell tumors in disorders of sex development. *Clin Genet* 91:292–301. <https://doi.org/10.1111/cge.12882>
170. Cools M, Looijenga L (2017) Update on the Pathophysiology and Risk Factors for the Development of Malignant Testicular Germ Cell Tumors in Complete Androgen Insensitivity Syndrome. *Sex Dev* 11:175–181. <https://doi.org/10.1159/000477921>
171. van der Zwan YG, Biermann K, Wolfenbutter KP, Cools M, Looijenga LHJ (2015) Gonadal maldevelopment as risk factor for germ cell cancer: towards a clinical decision model. *Eur Urol* 67:692–701. <https://doi.org/10.1016/j.eururo.2014.07.011>
172. Cools M, Looijenga LHJ, Wolfenbutter KP, T'Sjoen G (2014) Managing the risk of germ cell tumourigenesis in disorders of sex development patients. *Endocr Dev* 27:185–196. <https://doi.org/10.1159/000363642>
173. Looijenga LHJ, Kao C, Idrees MT (2019) Predicting Gonadal Germ Cell Cancer in People with Disorders of Sex Development; Insights from Developmental Biology. *Int J Mol Sci* 20:5017. <https://doi.org/10.3390/ijms20205017>
174. Johnson EK, Rosoklija I, Shurba A, D'Oro A, Gordon EJ, Chen D, Finlayson C, Holl JL (2017) Future fertility for individuals with differences of sex development: Parent attitudes and perspectives about decision-making. *J Pediatr Urol* 13:402–413. <https://doi.org/10.1016/j.jpuro.2017.06.002>
175. Finlayson C, Fritsch MK, Johnson EK, Rosoklija I, Gosiengfiao Y, Yerkes E, Madonna MB, Woodruff TK, Cheng E (2017) Presence of Germ Cells in Disorders of Sex Development: Implications for Fertility Potential and Preservation. *J Urol* 197:937–943. <https://doi.org/10.1016/j.juro.2016.08.108>
176. Nowotny HF, Reisch N (2023) Challenges Waiting for an Adult with DSD. *Horm Res Paediatr* 96:207–221. <https://doi.org/10.1159/000527433>
177. Aaronson IA (1994) Micropenis: medical and surgical implications. *J Urol* 152:4–14
178. Veale D, Miles S, Bramley S, Muir G, Hodson J (2015) Am I normal? A systematic review and construction of nomograms for flaccid and erect penis length and circumference in up to 15 521 men. *BJU Int* 115:978–986. <https://doi.org/10.1111/bju.13010>
179. Cheng PS, Chanoine J (2001) Should the Definition of Micropenis Vary According to Ethnicity? *Hormone Res Paediatrics* 55:278–281. <https://doi.org/10.1159/000050013>
180. Analia Tomova F, Deepinder R, Robeva H, Lalabonova P, Kumanov A Agarwal (2010) Growth and Development of Male External Genitalia. *Arch Pediatr Adolesc Med* 164:1152. <https://doi.org/10.1001/archpediatrics.2010.223>
181. Wang Y, Zeng Q, Xiong F, Zeng Y (2018) Male external genitalia growth curves and charts for children and adolescents aged 0 to 17 years in Chongqing, China. *Asian J Androl* 20:567–571. https://doi.org/10.4103/aja.aja_51_18
182. Bergeson PS, Hopkin RJ, Bailey RB, MCGill LC, Piatt JP (1993) The Inconspicuous Penis. *Pediatr (Evanston)* 92:794–799. <https://doi.org/10.1542/peds.92.6.794>
183. Zalel Y, Pinhas-Hamiel O, Lipitz S, Mashiach S, Achiron R (2001) The development of the fetal penis—an in utero sonographic evaluation. *Ultrasound Obstet Gynecol* 17:129–131. <https://doi.org/10.1046/j.1469-0705.2001.00216.x>
184. NELSON CP, PARK JM, WAN J, BLOOM DA, DUNN RL, WEI JT, THE INCREASING INCIDENCE OF CONGENITAL PENILE ANOMALIES IN THE UNITED STATES (2005) *J Urol* 174:1573–1576. <https://doi.org/10.1097/01.ju.0000179249.21944.7e>
185. Douglas A, Husmann (2002) Micropenis: An Animal Model and its Human Correlates. *Adv Exp Med Biol* 511:41–56. https://doi.org/10.1007/978-1-4615-0621-8_4
186. Baskin L, Sinclair A, Derpinghaus A, Cao M, Li Y, Overland M, Aksel S, Cunha GR (2021) Estrogens and development of the mouse and human external genitalia. *Differ (London)* 118:82–106. <https://doi.org/10.1016/j.diff.2020.09.004>
187. Baskin LS, Ridgewood (2020) *N J* 144:192–193. doi: <https://doi.org/10.1016/j.urology.2020.04.140>
188. Wiygul J, Palmer LS (2011) Micropenis. *Sci World J* 11:1462–1469. <https://doi.org/10.1100/tsw.2011.135>
189. Stancampiano MR, Suzuki K, O'Toole S, Russo G, Yamada G, Faisal Ahmed S (2022) Congenital Micropenis: Etiology And Management. *J Endocr Soc* 6:bvab172. <https://doi.org/10.1210/jendso/bvab172>
190. Khadilkar V, Mondkar SA (2023) Micropenis. *Indian J Pediatr* 90:598–604. <https://doi.org/10.1007/s12098-023-04540-w>
191. Cakir OO, Pozzi E, Castiglione F, Alnajjar HM, Salonia A, Muneer A (2021) Penile Length Measurement: Methodological Challenges and Recommendations, a Systematic Review. *J Sex Med* 18:433–439. <https://doi.org/10.1016/j.jsxm.2020.11.012>
192. Andersson A, Toppari J, Haavisto A, Petersen JH, Simell T, Simell O, Skakkebaek NE (1998) Longitudinal Reproductive Hormone Profiles in Infants: Peak of Inhibin B Levels in Infant Boys Exceeds Levels in Adult Men1. *J Clin Endocrinol Metab* 83:675–681. <https://doi.org/10.1210/jcem.83.2.4603>
193. Lanciotti L, Cofini M, Leonardi A, Penta L, Esposito S (2018) Up-To-Date Review About Minipuberty and Overview on Hypothalamic-Pituitary-Gonadal Axis Activation in Fetal and Neonatal Life. *Front Endocrinol* 9:410. <https://doi.org/10.3389/fendo.2018.00410>
194. Nina Callens G, De Cuypere E, Van Hoecke, Guy T, Sjoen S, Monstrey M, Cools P Hoebeke (2013) Sexual Quality of Life After Hormonal and Surgical Treatment, Including Phalloplasty, in Men with Micropenis: A Review. *J Sex Med* 10:2890–2903. <https://doi.org/10.1111/jsm.12298>
195. Arianne Dessens G, Guaragna-Filho A, Kyriakou J, Bryce C, Sanders A, Nordenskjöld M, Rozas V, Iotova A, Ediat A, Juul M, Krawczynski O, Hiort SF, Ahmed (2017) Understanding the needs of professionals who provide psychosocial care for children

- and adults with disorders of sex development. *BMJ Paediatrics Open* 1:e. <https://doi.org/10.1136/bmjpo-2017-000132>
196. Narinder Teckchandani M Bajpai (2014) Penile length nomogram for Asian Indian prepubertal boys. *J Pediatr Urol* 10:352–354. <https://doi.org/10.1016/j.jpuro.2013.09.017>
 197. Boas M, Boisen KA, Virtanen HE, Kaleva M, Suomi A, Schmidt IM, Damgaard IN, Kai CM, Chellakooty M, Skakkebaek NE, Toppari J, Main KM (2006) Postnatal penile length and growth rate correlate to serum testosterone levels: a longitudinal study of 1962 normal boys. *Eur J Endocrinol* 154:125–129. <https://doi.org/10.1530/eje.1.02066>
 198. Garcia-Quevedo L, Blanco J, Sarrate Z, Català V, Bassas L, Vidal F (2011) Hidden mosaicism in patients with Klinefelter's syndrome: implications for genetic reproductive counselling. *Hum Reprod (Oxford)* 26:3486–3493. <https://doi.org/10.1093/humrep/der351>
 199. Nadar R, Phadke N, Khatod K, Khadilkar V, Khadilkar AV (2014) Clinical applicability of rapid detection of SRY and DYS14 genes in patients with disorders of sex development using an indigenously developed 5' exonuclease based assay. *J Pediatr Endocrinol Metabolism* 27:869–872. <https://doi.org/10.1515/jpem-2013-0416>
 200. RajendraB Nerli AK, Guntaka PB, Patne MB Hiremath (2013) Penile growth in response to hormone treatment in children with micropenis. *Indian J Urol* 29:288. <https://doi.org/10.4103/0970-1591.120107>
 201. Xu D, Lu L, Xi L, Cheng R, Pei Z, Bi Y, Ruan S, Feihong Luo (2017) Efficacy and safety of percutaneous administration of dihydrotestosterone in children of different genetic backgrounds with micropenis. *J Pediatr Endocrinol Metab* 30:1285. <https://doi.org/10.1515/jpem-2016-0400>
 202. Arisaka O, Hoshi M, Kanazawa S, Nakajima D, Numata M, Nishikura K, Oyama M, Nitta A, Kuribayashi T, Kano K, Nakayama Y, Yamashiro Y (2001) Systemic effects of transdermal testosterone for the treatment of micropallus in children. *Pediatr Int* 43:134–136. <https://doi.org/10.1046/j.1442-200x.2001.01353.x>
 203. Karrou M, Messaoudi N, Assarrar I, Alla A, Rouf S, Latrech H (2023) Efficacy of Transdermal Dihydrotestosterone and Testosterone Enanthate for Penile Augmentation in Patients With Idiopathic Micropenis: A Comparative Randomized Study. *Clinical medicine insights. Endocrinol diabetes* 16:11795514231208328. <https://doi.org/10.1177/11795514231208328>
 204. McMahon DR, Kramer SA, Husmann DA (1995) Micropenis: Does Early Treatment with Testosterone Do More Harm Than Good? *J Urol* 154:825–829. [https://doi.org/10.1016/S0022-5347\(01\)67175-1](https://doi.org/10.1016/S0022-5347(01)67175-1)
 205. Falcone M, Bettocchi C, Carvalho J, Ricou M, Boeri L, Capogrosso P, Cocci A, Corona G, Gül M, Hatzichristodoulou G, Jones TH, Kadioğlu A, Kalkanli A, Martinez-Salamanca JI, Milenkovic U, Morgado LA, Russo GI, Serefoglu EC, Tharakan T, Verze P, Minhas S, Salonia A (2024) European Association of Urology Guidelines on Penile Size Abnormalities and Dysmorphophobia: Summary of the 2023 Guidelines. *Eur Urol focus* 10:432–441. <https://doi.org/10.1016/j.euf.2023.08.012>
 206. Laurence Baskin (2017) What Is Hypospadias? *Clin Pediatr* 56:409–418. <https://doi.org/10.1177/0009922816684613>
 207. Springer A, van den Heijkant M, Baumann S (2016) Worldwide prevalence of hypospadias. *J Pediatr Urol* 12:152e1–1527. <https://doi.org/10.1016/j.jpuro.2015.12.002>
 208. Loane M, Dolk H, Kelly A, Teljeur C, Greenlees R, Densem J (2011) Paper 4: EUROCAT statistical monitoring: Identification and investigation of ten year trends of congenital anomalies in Europe. *Birth Defects Res Part A: Clin Mol Teratology* 91:S31–S43. <https://doi.org/10.1002/bdra.20778>
 209. Li Y, Sinclair A, Cao M, Shen J, Choudhry S, Botta S, Cunha G, Baskin L (2015) Canalization of the Urethral Plate Precedes Fusion of the Urethral Folds during Male Penile Urethral Development: The Double Zipper Hypothesis. *J Urol* 193:1353–1360. <https://doi.org/10.1016/j.juro.2014.09.108>
 210. Porter MP, Faizan MK, Grady RW, Mueller BA (2005) Hypospadias in Washington State: Maternal Risk Factors and Prevalence Trends. *Pediatr (Evanston)* 115:e495–e499. <https://doi.org/10.1542/peds.2004-1552>
 211. Marijn M, Brouwers, Wouter FJ, Feitz LAJ, Roelofs, Lambertus ALM, Kiemeney, Robert PE, de Gier, Nel Roeleveld (2006) Risk factors for hypospadias. *Eur J Pediatrics* 166:671–678. <https://doi.org/10.1007/s00431-006-0304-z>
 212. Baskin LS, M.D (2008) Can we prevent hypospadias? *Fertil Steril* 89:e39. <https://doi.org/10.1016/j.fertnstert.2007.12.024>
 213. Carmichael SL, Mohammed N, Ma C, Iovannisci D, Choudhry S, Baskin LS, Witte JS, Shaw GM, Lammer EJ (2013) Diacylglycerol Kinase K Variants Impact Hypospadias in a California Study Population. *J Urol* 189:305–311. <https://doi.org/10.1016/j.juro.2012.09.002>
 214. George M, Schneuer FJ, Jamieson SE, Holland AJA (2015) Genetic and environmental factors in the aetiology of hypospadias. *Pediatr Surg Int* 31:519–527. <https://doi.org/10.1007/s00383-015-3686-z>
 215. van der Zanden LFM, van Rooij IALM, Feitz WFJ, Franke B, Knoers NVAM, Roeleveld N (2012) Aetiology of hypospadias: a systematic review of genes and environment. *Hum Reprod Update* 18:260–283. <https://doi.org/10.1093/humupd/dms002>
 216. Khuri FJ, Hardy BE, Churchill BM (1981) Urologic anomalies associated with hypospadias. *Urol Clin North Am* 8:565–571
 217. Duckett JWJ (1989) Hypospadias *Pediatr Rev* 11:37–42. <https://doi.org/10.1542/pir.11-2-37>
 218. van der Horst HJR, de Wall LL (2017) Hypospadias, all there is to know. *Eur J Pediatrics* 176:435–441. <https://doi.org/10.1007/s00431-017-2864-5>
 219. He Z, Yang B, Tang Y, Wang X (2024) Development and verification of machine learning model based on anogenital distance, penoscrotal distance, and 2D:4D finger ratio before puberty to predict hypospadias classification. *Front Pediatr* 12:1297642. <https://doi.org/10.3389/fped.2024.1297642>
 220. Cox K, Bryce J, Jiang J, Rodie M, Sinnott R, Alkhawari M, Arlt W, Audi L, Balsamo A, Bertelloni S, Cools M, Darendeliler F, Drop S, Ellaithi M, Guran T, Hiort O, Holterhus P, Hughes I, Krone N, Lisa L, Morel Y, Soder O, Wieacker P, Ahmed SF (2014) Novel Associations in Disorders of Sex Development: Findings From the I-DSD Registry. *J Clin Endocrinol Metab* 99:E348–E355. <https://doi.org/10.1210/jc.2013-2918>
 221. Ahmed SF, Dobbie R, Finlayson AR, Gilbert J, Youngson G, Chalmers J, Stone D (2004) Prevalence of hypospadias and other genital anomalies among singleton births, 1988–1997, in Scotland. *Archives Disease Child - Fetal Neonatal Ed* 89:149. <https://doi.org/10.1136/adc.2002.024034>
 222. Manzoni G, Bracka A, Palminteri E, Marrocco G (2004) Hypospadias surgery: when, what and by whom? *BJU Int* 94:1188–1195. <https://doi.org/10.1046/j.1464-410x.2004.05128.x>
 223. Tong SY, Donaldson K, Hutson JM (1996) When is hypospadias not hypospadias? *Med J Aust* 164:153–154. <https://doi.org/10.5694/j.1326-5377.1996.tb122014.x>
 224. Scougall K, Bryce J, Baronio F, Boal RL, Castera JR, Castro S, Cheetham T, Costa EC, Darendeliler F, Davies JH, Dirlwanger M, Gazdag G, Globa E, Guerra-Junior G, Guran T, Herrmann G, Holterhus P, Akgul AK, Markosyan R, McElreavey K, Miranda ML, Nordenstrom A, O'Toole S, Poyrazoglu S, Russo G, Schwitzgebel V, Stancampiano M, Steigert M, Ahmed SF, Lucas-Herald AK (2023) Predictors of surgical complications in boys with hypospadias: data from an international registry. *World J Pediatr Surg* 6:e000599–e000599 eCollection 2023. <https://doi.org/10.1136/wjps-2023-000599>

225. Sathyanarayana S, Grady R, Redmon JB, Ivceck K, Barrett E, Janssen S, Nguyen R, Swan SH, TIDES Study Team (2015) Anogenital distance and penile width measurements in The Infant Development and the Environment Study (TIDES): methods and predictors. *J Pediatr Urol* 11:76e1–76e6. <https://doi.org/10.1016/j.jpuro.2014.11.018>
226. Dodds PR, Batter SJ, Shield DE, Serels SR, Garafalo FA, Maloney PK (2008) Adaptation of adults to uncorrected hypospadias. *Urology* 71:682–685 discussion 685. <https://doi.org/10.1016/j.urology.2007.07.078>
227. Anonymous (1996) Timing of elective surgery on the genitalia of male children with particular reference to the risks, benefits, and psychological effects of surgery and anesthesia. *American Academy of Pediatrics. Pediatrics* 97:590–594
228. Bush NC, Holzer M, Zhang S, Snodgrass W (2013) Age does not impact risk for urethroplasty complications after tubularized incised plate repair of hypospadias in prepubertal boys. *J Pediatr Urol* 9:252–256. <https://doi.org/10.1016/j.jpuro.2012.03.014>
229. Bhat A, Bhat M, Kumar V, Kumar R, Mittal R, Saksena G (2016) Comparison of variables affecting the surgical outcomes of tubularized incised plate urethroplasty in adult and pediatric hypospadias. *J Pediatr Urol* 12:108e1–108e7. <https://doi.org/10.1016/j.jpuro.2015.09.005>
230. Silay MS, 't Hoen L, Bhatt N, Quaedackers J, Bogaert G, Dogan HS, Nijman RJM, Rawashdeh Y, Stein R, Tekgul S, Radmayr C (2021) Are there any benefits of using an inlay graft in the treatment of primary hypospadias in children? A systematic review and meta-analysis. *J Pediatr Urol* 17:303–315. <https://doi.org/10.1016/j.jpuro.2021.02.013>
231. Alshafei A, Cascio S, Boland F, O'Shea N, Hickey A, Quinn F (2020) Comparing the outcomes of tubularized incised plate urethroplasty and dorsal inlay graft urethroplasty in children with hypospadias: a systematic review and meta-analysis. *J Pediatr Urol* 16:154–161. <https://doi.org/10.1016/j.jpuro.2020.01.009>
232. Borkar N, Tiwari C, Nair A, Sinha CK, Ratan SK, Naredi BK (2024) Single dartos flap versus double dartos flap in hypospadias repair: A systematic review and meta-analysis with trial sequential analysis and fragility index. *Urologia* 91:439–447. <https://doi.org/10.1177/03915603241231058>
233. Herzberg H, Dubi-Sobol A, Mendelson T, Ben-David R, Bar-Yaakov N, Savin Z, Ben-Chaim J, Bar-Yosef Y (2022) Operative techniques and long-term outcomes of hypospadias repair in the absence of preputial skin after neonatal circumcision. *J Pediatr Surg* 57:676–680. <https://doi.org/10.1016/j.jpedsurg.2022.06.021>
234. Dunkel L, Quinton R (2014) Transition in endocrinology: induction of puberty. *Eur J Endocrinol* 170:229. <https://doi.org/10.1530/EJE-13-0894>
235. van der Horst HJR, de Wall LL (2017) Hypospadias, all there is to know. *Eur J Pediatr* 176:435–441. <https://doi.org/10.1007/s00431-017-2864-5>
236. Babu R, Chandrasekharam VVS (2021) Meta-analysis comparing the outcomes of single stage (foreskin pedicled tube) versus two stage (foreskin free graft & foreskin pedicled flap) repair for proximal hypospadias in the last decade. *J Pediatr Urol* 17:681–689. <https://doi.org/10.1016/j.jpuro.2021.05.014>
237. Chua ME, Gnech M, Ming JM, Silangeruz JM, Sanger S, Lopes RI, Lorenzo AJ, Braga LH (2017) Preoperative hormonal stimulation effect on hypospadias repair complications: Meta-analysis of observational versus randomized controlled studies. *J Pediatr Urol* 13:470–480. <https://doi.org/10.1016/j.jpuro.2017.06.019>
238. Shukla AR, Patel RP, Canning DA (2004) Hypospadias. *Urol Clin North Am* 31:445–60, viii. <https://doi.org/10.1016/j.ucl.2004.04.020>
239. Adams J, Bracka A (2016) Reconstructive surgery for hypospadias: A systematic review of long-term patient satisfaction with cosmetic outcomes. *Indian J Urol* 32:93–102. <https://doi.org/10.4103/0970-1591.179178>
240. Long CJ, Chu DI, Tenney RW, Morris AR, Weiss DA, Shukla AR, Srinivasan AK, Zderic SA, Kolon TF, Canning DA (2017) Intermediate-Term Followup of Proximal Hypospadias Repair Reveals High Complication Rate. *J Urol* 197:852–858. <https://doi.org/10.1016/j.juro.2016.11.054>
241. Spinoit A, Poelaert F, Van Praet C, Groen L, Van Laecke E, Hoebeke P (2015) Grade of hypospadias is the only factor predicting for re-intervention after primary hypospadias repair: a multivariate analysis from a cohort of 474 patients. *J Pediatr Urol* 11:70e1–70e6. <https://doi.org/10.1016/j.jpuro.2014.11.014>
242. Sullivan KJ, Hunter Z, Andrioli V, Guerra L, Leonard M, Klassen A, Keays MA (2017) Assessing quality of life of patients with hypospadias: A systematic review of validated patient-reported outcome instruments. *J Pediatr Urol* 13:19–27. <https://doi.org/10.1016/j.jpuro.2016.11.010>
243. van der Toorn F, de Jong TPVM, de Gier RPE, Callewaert PRH, van der Horst EHJR, Steffens MG, Hoebeke P, Nijman RJM, Bush NC, Wolfenbuttel KP, van den Heijkant MMC, van Capelle J, Wildhagen M, Timman R, van Busschbach JJV (2013) Introducing the HOPE (Hypospadias Objective Penile Evaluation)-score: a validation study of an objective scoring system for evaluating cosmetic appearance in hypospadias patients. *J Pediatr Urol* 9:1006–1016. <https://doi.org/10.1016/j.jpuro.2013.01.015>
244. Krull S, Rissmann A, Krause H, Mohnike K, Roehl F, Koehn A, Hass H (2018) Outcome after Hypospadias Repair: Evaluation Using the Hypospadias Objective Penile Evaluation Score. *Eur J Pediatr Surg* 28:268–272. <https://doi.org/10.1055/s-0037-1602252>
245. Ramnarine SD, Parente A, Vargas V, Escassi A, Paredes RM (2024) Long term follow-up on young adults that underwent hypospadias repair through urethral advancement in childhood. *Rev Int Androl* 22:23–28. <https://doi.org/10.22514/j.androl.2024.004>
246. Lammers RJM, Tsachouridis G, Andersson MK, Dormeus S, Ekerhult TO, Frankiewicz M, Gunn CJ, Matuszewski M, de Mooij KL, Schroeder RPJ, Wyndaele MIA, Xing Z, De Kort LMO, de Graaf P (2024) What should be next in lifelong posterior hypospadias: Conclusions from the 2023 ERN eUROGEN and EJP-RD networking meeting. *Neurourol Urodyn* 43:1097–1103. <https://doi.org/10.1002/nau.25305>
247. Ortqvist L, Andersson M, Strandqvist A, Nordenstrom A, Frisen L, Holmdahl G, Nordenskjold A (2017) Psychosocial outcome in adult men born with hypospadias. *J Pediatr Urol* 13:79e1–79e7. <https://doi.org/10.1016/j.jpuro.2016.08.008>
248. Rourke K, Braga LH (2018) Transitioning patients with hypospadias and other penile abnormalities to adulthood: What to expect? *Can Urol Assoc J* 12:S27–S33. <https://doi.org/10.5489/auaj.5227>
249. Wong NC, Braga LH (2015) The influence of pre-operative hormonal stimulation on hypospadias repair. *Front Pediatr* 3:31. <https://doi.org/10.3389/fped.2015.00031>
250. Azmat CE, Vaitla P (2025) Orchitis. Anonymous StatPearls. StatPearls Publishing LLC, Treasure Island (FL)
251. McConaghy JR, Panchal B (2016) Epididymitis: An Overview. *Am Fam Physician* 94:723–726
252. Trojan TH, Lishnak TS, Heiman D (2009) Epididymitis and orchitis: an overview. *Am Fam Physician* 79:583–587
253. Hviid A, Rubin S, Muhlemann K (2008) Mumps *Lancet* 371:932–944. [https://doi.org/10.1016/S0140-6736\(08\)60419-5](https://doi.org/10.1016/S0140-6736(08)60419-5)
254. Cavusoglu YH, Karaman A, Karaman I, Erdogan D, Aslan MK, Varlikli O, Cakmak O (2005) Acute scrotum -- etiology and management. *Indian J Pediatr* 72:201–203
255. Klin B, Zlotkevich L, Horne T, Efrati Y, Serour F, Lotan G (2001) Epididymitis in childhood: a clinical retrospective study over 5 years. *Isr Med Assoc J* 3:833–835

256. Makela E, Lahdes-Vasama T, Rajakorpi H, Wikstrom S (2007) A 19-year review of paediatric patients with acute scrotum. *Scand J Surg* 96:62–66. <https://doi.org/10.1177/145749690709600112>
257. McAndrew HF, Pemberton R, Kikiros CS, Gollow I (2002) The incidence and investigation of acute scrotal problems in children. *Pediatr Surg Int* 18:435–437. <https://doi.org/10.1007/s00383-002-0806-3>
258. Sakellaris GS, Charissis GC (2008) Acute epididymitis in Greek children: a 3-year retrospective study. *Eur J Pediatr* 167:765–769. <https://doi.org/10.1007/s00431-007-0584-y>
259. Somekh E, Gorenstein A, Serour F (2004) Acute epididymitis in boys: evidence of a post-infectious etiology. *J Urol* 171:391–394 discussion 394. <https://doi.org/10.1097/01.ju.0000102160.55494.1f>
260. Kadish HA, Bolte RG (1998) A retrospective review of pediatric patients with epididymitis, testicular torsion, and torsion of testicular appendages. *Pediatrics* 102:73–76. <https://doi.org/10.1542/peds.102.1.73>
261. Sauvat F, Hennequin S, Ait Ali Slimane M, Gauthier F (2002) Age for testicular torsion? *Arch Pediatr* 9:1226–1229. [https://doi.org/10.1016/s0929-693x\(02\)00112-4](https://doi.org/10.1016/s0929-693x(02)00112-4)
262. Gamoudi D, Flew S, Benardon S, Poder A, Radcliffe K (2019) 2018 European guideline on the organization of a consultation for sexually transmitted infections. *J Eur Acad Dermatol Venereol* 33:1452–1458. <https://doi.org/10.1111/jdv.15577>
263. Garthwaite MAE, Johnson G, Lloyd S, Eardley I (2007) The implementation of European Association of Urology guidelines in the management of acute epididymo-orchitis. *Ann R Coll Surg Engl* 89:799–803. <https://doi.org/10.1308/003588407X232026>
264. Hagley M (2003) Epididymo-orchitis and epididymitis: a review of causes and management of unusual forms. *Int J STD AIDS* 14:372–377 quiz 378. <https://doi.org/10.1258/095646203765371240>
265. Street EJ, Justice ED, Kopa Z, Portman MD, Ross JD, Skerlev M, Wilson JD, Patel R (2017) The 2016 European guideline on the management of epididymo-orchitis. *Int J STD AIDS* 28:744–749. <https://doi.org/10.1177/0956462417699356>
266. Pilatz A, Hossain H, Kaiser R, Mankertz A, Schuttler CG, Domann E, Schuppe H, Chakraborty T, Weidner W, Wagenlehner F (2015) Acute epididymitis revisited: impact of molecular diagnostics on etiology and contemporary guideline recommendations. *Eur Urol* 68:428–435. <https://doi.org/10.1016/j.eururo.2014.12.005>
267. Tracy CR, Steers WD, Costabile R (2008) Diagnosis and management of epididymitis. *Urol Clin North Am* 35:101–108. <https://doi.org/10.1016/j.ucl.2007.09.013>. vii
268. Collinson K, Murray MJ, Orsi NM, Cummings M, Shipley J, Joffe JK, Coleman N, Stark D (2014) Age-related biological features of germ cell tumors. *Genes Chromosomes Cancer* 53:215–227. <https://doi.org/10.1002/gcc.22131>
269. Rodriguez S, Jafer O, Goker H, Summersgill BM, Zafarana G, Gillis AJM, van Gurp, R J H L M, Oosterhuis JW, Lu Y, Huddart R, Cooper CS, Clark J, Looijenga LHJ, Shipley JM (2003) Expression profile of genes from 12p in testicular germ cell tumors of adolescents and adults associated with i(12p) and amplification at 12p11.2-p12.1. *Oncogene* 22:1880–1891. <https://doi.org/10.1038/sj.onc.1206302>
270. Palmer RD, Foster NA, Vowler SL, Roberts I, Thornton CM, Hale JP, Schneider DT, Nicholson JC, Coleman N (2007) Malignant germ cell tumours of childhood: new associations of genomic imbalance. *Br J Cancer* 96:667–676. <https://doi.org/10.1038/sj.bjc.6603602>
271. Harms D, Zahn S, Gobel U, Schneider DT (2006) Pathology and molecular biology of teratomas in childhood and adolescence. *Klin Padiatr* 218:296–302. <https://doi.org/10.1055/s-2006-942271>
272. Jarvis H, Cost NG, Saltzman AF (2021) Testicular tumors in the pediatric patient. *Semin Pediatr Surg* 30:151079. <https://doi.org/10.1016/j.sempedsurg.2021.151079>
273. Znaor A, Skakkebaek NE, Rajpert-De Meyts E, Kulis T, Laversanne M, Gurney J, Sarfati D, McGlynn KA, Bray F (2022) Global patterns in testicular cancer incidence and mortality in 2020. *Int J Cancer* 151:692–698. <https://doi.org/10.1002/ijc.33999>
274. Ward E, DeSantis C, Robbins A, Kohler B, Jemal A (2014) Childhood and adolescent cancer statistics, 2014. *CA Cancer J Clin* 64:83–103. <https://doi.org/10.3322/caac.21219>
275. Pierce JL, Frazier AL, Amatruda JF (2018) Pediatric Germ Cell Tumors: A Developmental Perspective. *Adv Urol* 2018:9059382. <https://doi.org/10.1155/2018/9059382>
276. Stein R, Quaedackers J, Bhat NR, Dogan HS, Nijman RJM, Rawashdeh YF, Silay MS, 't Hoen LA, Tekgul S, Radmayr C, Bogaert G (2021) EAU-ESPU pediatric urology guidelines on testicular tumors in prepubertal boys. *J Pediatr Urol* 17:529–533. <https://doi.org/10.1016/j.jpuro.2021.06.006>
277. Taskinen S, Fagerholm R, Aronniemi J, Rintala R, Taskinen M (2008) Testicular tumors in children and adolescents. *J Pediatr Urol* 4:134–137. <https://doi.org/10.1016/j.jpuro.2007.10.002>
278. Cost NG, Lubahn JD, Adibi M, Romman A, Wickiser JE, Raj GV, Sagalowsky AI, Margulis V (2014) A comparison of pediatric, adolescent, and adult testicular germ cell malignancy. *Pediatr Blood Cancer* 61:446–451. <https://doi.org/10.1002/pbc.24773>
279. Williamson SR, Delahunt B, Magi-Galluzzi C, Algaba F, Egevad L, Ulbright TM, Tickoo SK, Srigley JR, Epstein JI, Berney DM, Members of the ISUP Testicular Tumour Panel (2017) The World Health Organization 2016 classification of testicular germ cell tumours: a review and update from the International Society of Urological Pathology Testis Consultation Panel. *Histopathology* 70:335–346. <https://doi.org/10.1111/his.13102>
280. Berney DM, Looijenga LHJ, Idrees M, Oosterhuis JW, Rajpert-De Meyts E, Ulbright TM, Skakkebaek NE (2016) Germ cell neoplasia in situ (GCNIS): evolution of the current nomenclature for testicular pre-invasive germ cell malignancy. *Histopathology* 69:7–10. <https://doi.org/10.1111/his.12958>
281. Rajpert-De Meyts E (2006) Developmental model for the pathogenesis of testicular carcinoma in situ: genetic and environmental aspects. *Hum Reprod Update* 12:303–323. <https://doi.org/10.1093/humupd/dmk006>
282. Moch H, Cubilla AL, Humphrey PA, Reuter VE, Ulbright TM (2016) The 2016 WHO Classification of Tumours of the Urinary System and Male Genital Organs-Part A: Renal, Penile, and Testicular Tumours. *Eur Urol* 70:93–105. <https://doi.org/10.1016/j.eururo.2016.02.029>
283. Mosbech CH, Rechnitzer C, Brok JS, Rajpert-De Meyts E, Hoei-Hansen CE (2014) Recent advances in understanding the etiology and pathogenesis of pediatric germ cell tumors. *J Pediatr Hematol Oncol* 36:263–270. <https://doi.org/10.1097/MPH.0000000000000125>
284. Oosterhuis JW, Looijenga LHJ (2019) Human germ cell tumours from a developmental perspective. *Nat Rev Cancer* 19:522–537. <https://doi.org/10.1038/s41568-019-0178-9>
285. Luckie TM, Danzig M, Zhou S, Wu H, Cost NG, Karaviti L, Venkatraman R (2019) A Multicenter Retrospective Review of Pediatric Leydig Cell Tumor of the Testis. *J Pediatr Hematol Oncol* 41:74–76. <https://doi.org/10.1097/MPH.0000000000001124>
286. Grogg JB, Schneider K, Bode P, Kranzbuhler B, Eberli D, Sulser T, Beyer J, Lorch A, Hermanns T, Fankhauser CD (2020) Risk factors and treatment outcomes of 239 patients with testicular granulosa cell tumors: a systematic review of published case series data. *J Cancer Res Clin Oncol* 146:2829–2841. <https://doi.org/10.1007/s00432-020-03326-3>

287. Talon I, Moog R, Kauffmann I, Grandadam S, Becmeur F (2005) Sertoli cell tumor of the testis in children: reevaluation of a rarely encountered tumor. *J Pediatr Hematol Oncol* 27:491–494. <https://doi.org/10.1097/01.mph.0000179960.65265.a2>
288. Amini A, Waxweiler TV, Maroni PD, Kessler ER, Cost CR, Greffe BS, Garrington TP, Liu AK, Cost NG (2016) Survival outcomes of adolescent and adult patients with non-seminomatous testicular germ-cell tumors: A population-based study. *J Pediatr Urol* 12:405e1–405e9. <https://doi.org/10.1016/j.jpuro.2016.06.014>
289. Veneroni L, Mariani L, Lo Vullo S, Favini F, Catania S, Vajna de Pava M, Massimino M, Ferrari A (2013) Symptom interval in pediatric patients with solid tumors: adolescents are at greater risk of late diagnosis. *Pediatr Blood Cancer* 60:605–610. <https://doi.org/10.1002/pbc.24312>
290. Gianfrilli D, Ferlin A, Isidori AM, Garolla A, Maggi M, Pivonello R, Santi D, Sansone A, Balercia G, Granata ARM, Sinisi A, Lanfranco F, Pasqualetti P, Foresta C, Lenzi A, ‘Amico-Andrologo’ Study Group (2019) Risk behaviours and alcohol in adolescence are negatively associated with testicular volume: results from the Amico-Andrologo survey. *Andrology* 7:769–777. <https://doi.org/10.1111/andr.12659>
291. Bleyer A, Barr R, Hayes-Lattin B, Thomas D, Ellis C, Anderson B, Biology and Clinical Trials Subgroups of the US National Cancer Institute Progress Review Group in Adolescent and Young Adult Oncology (2008) The distinctive biology of cancer in adolescents and young adults. *Nat Rev Cancer* 8:288–298. <https://doi.org/10.1038/nrc2349>
292. Chan E, Wayne C, Nasr A, FRCSC for Canadian Association of Pediatric Surgeon Evidence-Based Resource (2014) Ideal timing of orchiopexy: a systematic review. *Pediatr Surg Int* 30:87–97. <https://doi.org/10.1007/s00383-013-3429-y>
293. Schraw JM, Sok P, Desrosiers TA, Janitz AE, Langlois PH, Canfield MA, Frazier AL, Plon SE, Lupo PJ, Poynter JN (2023) Associations between birth defects and childhood and adolescent germ cell tumors according to sex, histologic subtype, and site. *Cancer* 129:3300–3308. <https://doi.org/10.1002/cncr.34906>
294. Chia VM, Li Y, Goldin LR, Graubard BI, Greene MH, Korde L, Rubertone MV, Erickson RL, McGlynn KA (2009) Risk of cancer in first- and second-degree relatives of testicular germ cell tumor cases and controls. *Int J Cancer* 124:952–957. <https://doi.org/10.1002/ijc.23971>
295. Zequi SC, da Costa WH, Santana TBM, Favaretto RL, Sacomani CAR, Guimaraes GC (2012) Bilateral testicular germ cell tumours: a systematic review. *BJU Int* 110:1102–1109. <https://doi.org/10.1111/j.1464-410X.2012.11056.x>
296. Marte A, Pintozzi L, Creti G, Chiesa PL, Renzo DD, Gasparella M, Maggio GD, Bagnara V, Merlini E, Tadini B, Caldaruolo E, Sangiorgio L, Battaglini G, Nappo SG, Caione P (2017) Long-Term Follow-Up of Testicular Microlithiasis in Children and Adolescents: Multicenter Prospective Cohort Study of the Italian Society of Pediatric Urology. *Eur J Pediatr Surg* 27:155–160. <https://doi.org/10.1055/s-0036-1572552>
297. Volokhina YV, Oyoyo UE, Miller JH (2014) Ultrasound demonstration of testicular microlithiasis in pediatric patients: is there an association with testicular germ cell tumors? *Pediatr Radiol* 44:50–55. <https://doi.org/10.1007/s00247-013-2778-y>
298. Trout AT, Chow J, McNamara ER, Darge K, Ramirez Grueso R, Munden M, Rothan SM, Navarro OM, Tijerin Bueno M, Bove KE, Chikwava KR, Heider A, Hicks MJ, Somers GR, Zhang B, Dillman JR (2017) Association between Testicular Microlithiasis and Testicular Neoplasia: Large Multicenter Study in a Pediatric Population. *Radiology* 285:576–583. <https://doi.org/10.1148/radiol.2017162625>
299. Cooper ML, Kaefer M, Fan R, Rink RC, Jennings SG, Karmazyn B (2014) Testicular microlithiasis in children and associated testicular cancer. *Radiology* 270:857–863. <https://doi.org/10.1148/radiol.13130394>
300. 't Hoen LA, Bhatt NR, Radmayr C, Dogan HS, Nijman RJM, Quaedackers J, Rawashdeh YF, Silay MS, Tekgul S, Stein R, Bogaert G (2021) The prognostic value of testicular microlithiasis as an incidental finding for the risk of testicular malignancy in children and the adult population: A systematic review. On behalf of the EAU pediatric urology guidelines panel. *J Pediatr Urol* 17:815–831. <https://doi.org/10.1016/j.jpuro.2021.06.013>
301. Pozza C, Tenuta M, Sesti F, Bertolotto M, Huang DY, Sidhu PS, Maggi M, Isidori AM, Lotti F (2023) Multiparametric Ultrasound for Diagnosing Testicular Lesions: Everything You Need to Know in Daily Clinical Practice. *Cancers (Basel)* 15:5332. <https://doi.org/10.3390/cancers15225332>
302. Spaziani M, Lecis C, Tarantino C, Sbardella E, Pozza C, Gianfrilli D (2021) The role of scrotal ultrasonography from infancy to puberty. *Andrology* 9:1306–1321. <https://doi.org/10.1111/andr.13056>
303. Tallen G, Hernaiz Driever P, Degenhardt P, Henze G, Riebel T (2011) High reliability of scrotal ultrasonography in the management of childhood primary testicular neoplasms. *Klin Padiatr* 223:131–137. <https://doi.org/10.1055/s-0031-1271813>
304. Wunsch R, von Rohden L, Cleaveland R, Aumann V (2014) Small part ultrasound in childhood and adolescence. *Eur J Radiol* 83:1549–1559. <https://doi.org/10.1016/j.ejrad.2014.04.028>
305. Ross JH, Rybicki L, Kay R (2002) Clinical behavior and a contemporary management algorithm for prepubertal testis tumors: a summary of the Prepubertal Testis Tumor Registry. *J Urol* 168:1675–1679. <https://doi.org/10.1097/01.ju.0000030749.27823.f5>
306. Murray MJ, Huddart RA, Coleman N (2016) The present and future of serum diagnostic tests for testicular germ cell tumours. *Nat Rev Urol* 13:715–725. <https://doi.org/10.1038/nrurol.2016.170>
307. Stokes W, Amini A, Maroni PD, Kessler ER, Stokes C, Cost CR, Greffe BS, Garrington TP, Liu AK, Cost NG (2017) Patterns of care and survival outcomes for adolescent and young adult patients with testicular seminoma in the United States: A National Cancer Database analysis. *J Pediatr Urol* 13:386e1–386e7. <https://doi.org/10.1016/j.jpuro.2016.12.009>
308. Manganaro L, Saldari M, Pozza C, Vinci V, Gianfrilli D, Greco E, Franco G, Sergi ME, Scialpi M, Catalano C, Isidori AM (2018) Dynamic contrast-enhanced and diffusion-weighted MR imaging in the characterisation of small, non-palpable solid testicular tumours. *Eur Radiol* 28:554–564. <https://doi.org/10.1007/s00330-017-5013-7>
309. Tenuta M, Sesti F, Bonaventura I, Mazzotta P, Pofi R, Gianfrilli D, Pozza C (2021) Use of contrast enhanced ultrasound in testicular diseases: A comprehensive review. *Andrology* 9:1369–1382. <https://doi.org/10.1111/andr.13057>
310. Isidori AM, Pozza C, Gianfrilli D, Giannetta E, Lemma A, Pofi R, Barbagallo F, Manganaro L, Martino G, Lombardo F, Cantisani V, Franco G, Lenzi A (2014) Differential diagnosis of nonpalpable testicular lesions: qualitative and quantitative contrast-enhanced US of benign and malignant testicular tumors. *Radiology* 273:606–618. <https://doi.org/10.1148/radiol.14132718>
311. Frazier AL, Hale JP, Rodriguez-Galindo C, Dang H, Olson T, Murray MJ, Amatruda JF, Thornton C, Arul GS, Billmire D, Shaikh F, Pashankar F, Stoneham S, Krailo M, Nicholson JC (2015) Revised risk classification for pediatric extracranial germ cell tumors based on 25 years of clinical trial data from the United Kingdom and United States. *J Clin Oncol* 33:195–201. <https://doi.org/10.1200/JCO.2014.58.3369>
312. Fankhauser CD, Grogg JB, Hayoz S, Wettstein MS, Dieckmann K, Sulser T, Bode P, Clarke NW, Beyer J, Hermanns T (2020)

- Risk Factors and Treatment Outcomes of 1,375 Patients with Testicular Leydig Cell Tumors: Analysis of Published Case Series Data. *J Urol* 203:949–956. <https://doi.org/10.1097/JU.0000000000000705>
313. Bujons A, Sfulcini JC, Pascual M, Feu OA, Garat JM, Villavicencio H (2011) Prepubertal testicular tumours and efficacy of testicular preserving surgery. *BJU Int* 107:1812–1816. <https://doi.org/10.1111/j.1464-410X.2010.09796.x>
 314. Silverio PC, Schoofs F, Iselin CE, Tille J (2015) Fourteen-year experience with the intraoperative frozen section examination of testicular lesion in a tertiary university center. *Ann Diagn Pathol* 19:99–102. <https://doi.org/10.1016/j.anndiagpath.2014.12.006>
 315. Matei DV, Vartolomei MD, Renne G, Tringali VML, Russo A, Bianchi R, Cozzi G, Bottero D, Musi G, Mazzarol G, Ferro M, de Cobelli O (2017) Reliability of Frozen Section Examination in a Large Cohort of Testicular Masses: What Did We Learn? *Clin Genitourin Cancer* 15:e689–e696. <https://doi.org/10.1016/j.clgc.2017.01.012>
 316. Anderson RA, Mitchell RT, Kelsey TW, Spears N, Telfer EE, Wallace WHB (2015) Cancer treatment and gonadal function: experimental and established strategies for fertility preservation in children and young adults. *Lancet Diabetes Endocrinol* 3:556–567. [https://doi.org/10.1016/S2213-8587\(15\)00039-X](https://doi.org/10.1016/S2213-8587(15)00039-X)
 317. Wallace WHB, Anderson RA, Irvine DS (2005) Fertility preservation for young patients with cancer: who is at risk and what can be offered? *Lancet Oncol* 6:209–218. [https://doi.org/10.1016/S1470-2045\(05\)70092-9](https://doi.org/10.1016/S1470-2045(05)70092-9)
 318. Gandini L, Sgro P, Lombardo F, Paoli D, Culasso F, Toselli L, Tsamatopoulos P, Lenzi A (2006) Effect of chemo- or radiotherapy on sperm parameters of testicular cancer patients. *Hum Reprod* 21:2882–2889. <https://doi.org/10.1093/humrep/del167>
 319. Mistretta FA, de Cobelli O, Verze P, Botticelli F, Jannello L, Luzzagio S, Cozzi G, Bianchi R, Di Trapani E, Ferro M, Cordima G, Bottero D, Matei DV, Mirone V, Musi G (2022) A comprehensive evaluation of sexual and reproductive outcomes following robot-assisted retroperitoneal lymph node dissection for nonseminomatous germ cell tumor. *Asian J Androl* 24:579–583. <https://doi.org/10.4103/aja2021132>
 320. Long CJ, Ginsberg JP, Kolon TF (2016) Fertility Preservation in Children and Adolescents With Cancer. *Urology* 91:190–196. <https://doi.org/10.1016/j.urology.2015.10.047>
 321. Ginsberg JP, Ogle SK, Tuchman LK, Carlson CA, Reilly MM, Hobbie WL, Rourke M, Zhao H, Meadows AT (2008) Sperm banking for adolescent and young adult cancer patients: sperm quality, patient, and parent perspectives. *Pediatr Blood Cancer* 50:594–598. <https://doi.org/10.1002/pbc.21257>
 322. Fonseca A, Frazier AL, Shaikh F (2019) Germ Cell Tumors in Adolescents and Young Adults. *J Oncol Pract* 15:433–441. <https://doi.org/10.1200/JOP.19.00190>
 323. Rogers PC, Olson TA, Cullen JW, Billmire DF, Marina N, Rescorla F, Davis MM, London WB, Lauer SJ, Giller RH, Cushing B, Pediatric Oncology Group 9048, Children's Cancer Group 8891 (2004) Treatment of children and adolescents with stage II testicular and stages I and II ovarian malignant germ cell tumors: A Pediatric Intergroup Study–Pediatric Oncology Group 9048 and Children's Cancer Group 8891. *J Clin Oncol* 22:3563–3569. <https://doi.org/10.1200/JCO.2004.01.006>
 324. de Wit R, Roberts JT, Wilkinson PM, de Mulder PH, Mead GM, Fossa SD, Cook P, de Prieck L, Stenning S, Collette L (2001) Equivalence of three or four cycles of bleomycin, etoposide, and cisplatin chemotherapy and of a 3- or 5-day schedule in good-prognosis germ cell cancer: a randomized study of the European Organization for Research and Treatment of Cancer Genitourinary Tract Cancer Cooperative Group and the Medical Research Council. *PG* 629–40. *J Clin Oncol* 19:1629–1640. <https://doi.org/10.1200/JCO.2001.19.6.1629>
 325. Frazier AL, Stoneham S, Rodriguez-Galindo C, Dang H, Xia C, Olson TA, Murray MJ, Amatruda JF, Shaikh F, Pashankar F, Billmire D, Krailo M, Stark D, Brougham MFH, Nicholson JC, Hale JP (2018) Comparison of carboplatin versus cisplatin in the treatment of paediatric extracranial malignant germ cell tumours: A report of the Malignant Germ Cell International Consortium. *Eur J Cancer* 98:30–37. <https://doi.org/10.1016/j.ejca.2018.03.004>
 326. Shaikh F, Nathan PC, Hale J, Uleryk E, Frazier L (2013) Is there a role for carboplatin in the treatment of malignant germ cell tumors? A systematic review of adult and pediatric trials. *Pediatr Blood Cancer* 60:587–592. <https://doi.org/10.1002/pbc.24288>
 327. Cost NG, Cost CR, Geller JI, Defoor WRJ (2014) Adolescent urologic oncology: current issues and future directions. *Urol Oncol* 32:59–69. <https://doi.org/10.1016/j.urolonc.2012.08.002>
 328. Cafferty FH, Gabe R, Huddart RA, Rustin G, Stenning SP, Bara A, Bathia R, Freeman SC, Alder L, Joffe JK (2012) UK management practices in stage I seminoma and the Medical Research Council Trial of Imaging and Schedule in Seminoma Testis managed with surveillance. *Clin Oncol (R Coll Radiol)* 24:25–29. <https://doi.org/10.1016/j.clon.2011.09.005>
 329. Schlatter M, Rescorla F, Giller R, Cushing B, Vinocur C, Colombani P, Cullen J, London W, Davis M, Lauer S, Olson T, Children's Cancer Group, Pediatric Oncology Group (2003) Excellent outcome in patients with stage I germ cell tumors of the testes: a study of the Children's Cancer Group/Pediatric Oncology Group. *J Pediatr Surg* 38:319–324. <https://doi.org/10.1053/jpsu.2003.50101>
 330. Kollmannsberger C, Tandstad T, Bedard PL, Cohn-Cedermark G, Chung PW, Jewett MA, Powles T, Warde PR, Daneshmand S, Protheroe A, Tyldesley S, Black PC, Chi K, So AI, Moore MJ, Nichols CR (2015) Patterns of relapse in patients with clinical stage I testicular cancer managed with active surveillance. *J Clin Oncol* 33:51–57. <https://doi.org/10.1200/JCO.2014.56.2116>
 331. Cost NG, Lubahn JD, Adibi M, Romman A, Wickiser JE, Raj GV, Sagalowsky AI, Margulis V (2014) Risk stratification of pubertal children and postpubertal adolescents with clinical stage I testicular nonseminomatous germ cell tumors. *J Urol* 191:1485–1490. <https://doi.org/10.1016/j.juro.2013.08.047>
 332. Kenney LB, Cohen LE, Shnorhavorian M, Metzger ML, Lockart B, Hijiya N, Duffey-Lind E, Constine L, Green D, Meacham L (2012) Male reproductive health after childhood, adolescent, and young adult cancers: a report from the Children's Oncology Group. *J Clin Oncol* 30:3408–3416. <https://doi.org/10.1200/JCO.2011.38.6938>
 333. Giwercman A, von der Maase H, Berthelsen JG, Rorth M, Bertelsen A, Skakkebaek NE (1991) Localized irradiation of testes with carcinoma in situ: effects on Leydig cell function and eradication of malignant germ cells in 20 patients. *J Clin Endocrinol Metab* 73:596–603. <https://doi.org/10.1210/jcem-73-3-596>
 334. Oliver RTD, Mead GM, Rustin GJS, Joffe JK, Aass N, Coleman R, Gabe R, Pollock P, Stenning SP (2011) Randomized trial of carboplatin versus radiotherapy for stage I seminoma: mature results on relapse and contralateral testis cancer rates in MRC TE19/EORTC 30982 study (ISRCTN27163214). *J Clin Oncol* 29:957–962. <https://doi.org/10.1200/JCO.2009.26.4655>
 335. Jacobs LA, Vaughn DJ (2012) Hypogonadism and infertility in testicular cancer survivors. *J Natl Compr Canc Netw* 10:558–563. <https://doi.org/10.6004/jncn.2012.0053>
 336. Johannsdottir IM, Karlstad O, Loge JH, Fossa SD, Kiserud C, Skurtveit S (2017) Prescriptions of Antidepressants to Survivors of Cancer in Childhood, Adolescence, and Young Adulthood: A Population-Based Study. *J Adolesc Young Adult Oncol* 6:120–126. <https://doi.org/10.1089/jayao.2016.0041>
 337. Darabos K, Hoyt MA (2017) Cancer-Related Worry and Physical Well-Being in the Context of Perceived Stress in Young Adults

- with Testicular Cancer. *J Adolesc Young Adult Oncol* 6:363–366. <https://doi.org/10.1089/jayao.2016.0069>
338. Waldert M, Klatte T, Schmidbauer J, Remzi M, Lackner J, Marberger M (2010) Color Doppler sonography reliably identifies testicular torsion in boys. *Urology* 75:1170–1174. <https://doi.org/10.1016/j.urology.2009.07.1298>
 339. Drlik M, Kocvara R (2013) Torsion of spermatic cord in children: a review. *J Pediatr Urol* 9:259–266. <https://doi.org/10.1016/j.jpuro.2012.05.016>
 340. Bandarkar AN, Blask AR (2018) Testicular torsion with preserved flow: key sonographic features and value-added approach to diagnosis. *Pediatr Radiol* 48:735–744. <https://doi.org/10.1007/s00247-018-4093-0>
 341. Zhao LC, Lautz TB, Meeks JJ, Maizels M (2011) Pediatric testicular torsion epidemiology using a national database: incidence, risk of orchiectomy and possible measures toward improving the quality of care. *J Urol* 186:2009–2013. <https://doi.org/10.1016/j.juro.2011.07.024>
 342. Grimsby GM, Schlomer BJ, Menon VS, Ostrov L, Keays M, Sheth KR, Villanueva C, Granberg C, Dajusta D, Hill M, Sanchez E, Harrison CB, Jacobs MA, Burgu B, Hennes H, Baker LA (2018) Prospective Evaluation of Predictors of Testis Atrophy After Surgery for Testis Torsion in Children. *Urology* 116:150–155. <https://doi.org/10.1016/j.urology.2018.03.009>
 343. Wampler SM, Llanes M (2010) Common scrotal and testicular problems. *Prim Care* 37:613–26, x. <https://doi.org/10.1016/j.pop.2010.04.009>
 344. Ludvigson AE, Beaulé LT (2016) Urologic Emergencies. *Surg Clin North Am* 96:407–424. <https://doi.org/10.1016/j.suc.2016.02.001>
 345. Ringdahl E, Teague L (2006) Testicular torsion. *Am Fam Physician* 74:1739–1743
 346. Ekici M, Ozgur BC, Senturk AB, Nalbant I (2018) Relationship of Low Temperature with Testicular Torsion. *J Coll Physicians Surg Pak* 28:378–380. <https://doi.org/10.29271/jcpsp.2018.05.378>
 347. Davis JE, Silverman M (2011) Scrotal emergencies. *Emerg Med Clin North Am* 29:469–484. <https://doi.org/10.1016/j.emc.2011.04.011>
 348. Dogra VS, Rubens DJ, Gottlieb RH, Bhatt S (2004) Torsion and beyond: new twists in spectral Doppler evaluation of the scrotum. *J Ultrasound Med* 23:1077–1085. <https://doi.org/10.7863/jum.2004.23.8.1077>
 349. Rabinowitz R (1984) The importance of the cremasteric reflex in acute scrotal swelling in children. *J Urol* 132:89–90. [https://doi.org/10.1016/s0022-5347\(17\)49476-6](https://doi.org/10.1016/s0022-5347(17)49476-6)
 350. Sheth KR, Keays M, Grimsby GM, Granberg CF, Menon VS, Dajusta DG, Ostrov L, Hill M, Sanchez E, Kuppermann D, Harrison CB, Jacobs MA, Huang R, Burgu B, Hennes H, Schlomer BJ, Baker LA (2016) Diagnosing Testicular Torsion before Urological Consultation and Imaging: Validation of the TWIST Score. *J Urol* 195:1870–1876. <https://doi.org/10.1016/j.juro.2016.01.101>
 351. Barada JH, Weingarten JL, Cromie WJ (1989) Testicular salvage and age-related delay in the presentation of testicular torsion. *J Urol* 142:746–748. [https://doi.org/10.1016/s0022-5347\(17\)38875-4](https://doi.org/10.1016/s0022-5347(17)38875-4)
 352. Howe AS, Vasudevan V, Kongnyuy M, Rychik K, Thomas LA, Matuskova M, Friedman SC, Gitlin JS, Reda EF, Palmer LS (2017) Degree of twisting and duration of symptoms are prognostic factors of testis salvage during episodes of testicular torsion. *Transl Androl Urol* 6:1159–1166. <https://doi.org/10.21037/tau.2017.09.10>
 353. Samson P, Hartman C, Palmerola R, Rahman Z, Siev M, Palmer LS, Ghorayeb SR (2017) Ultrasonographic Assessment of Testicular Viability Using Heterogeneity Levels in Torsed Testicles. *J Urol* 197:925–930. <https://doi.org/10.1016/j.juro.2016.09.112>
 354. Expert Panel on Urological Imaging; Wang CL, Aryal B, Oto A, Allen BC, Akin O, Alexander LF, Bardo DME, Chong J, Froemming AT, Fulgham PF, Heller MT, Maranchie JK, Mody RN, Patel BN, Schieda N, Turkbey IB, Venkatesan AM, Yoo DC, Lockhart ME (2019) ACR Appropriateness Criteria((R)) Acute Onset of Scrotal Pain-Without Trauma, Without Antecedent Mass. *J Am Coll Radiol* 16:S38–S43. <https://doi.org/10.1016/j.jacr.2019.02.016>
 355. Sharp VJ, Kieran K, Arlen AM (2013) Testicular torsion: diagnosis, evaluation, and management. *Am Fam Physician* 88:835–840
 356. Sparano A, Acampora C, Scaglione M, Romano L (2008) Using color power Doppler ultrasound imaging to diagnose the acute scrotum. A pictorial essay. *Emerg Radiol* 15:289–294. <https://doi.org/10.1007/s10140-008-0710-9>
 357. Bitkin A, Aydin M, Ozgur BC, Irkilata L, Akgunes E, Keles M, Sarici H, Atilla MK (2018) Can haematologic parameters be used for differential diagnosis of testicular torsion and epididymitis? *Andrologia* 50:https://doi.org/10.1111/and.12819. Epub 2017 May 12. doi: 10.1111/and.12819
 358. Yu K, Wang T, Chen H, Wang H (2012) The dilemma in the diagnosis of acute scrotum: clinical clues for differentiating between testicular torsion and epididymo-orchitis. *Chang Gung Med J* 35:38–45. <https://doi.org/10.4103/2319-4170.106168>
 359. Munden MM, Williams JL, Zhang W, Crowe JE, Munden RF, Cisek LJ (2013) Intermittent testicular torsion in the pediatric patient: sonographic indicators of a difficult diagnosis. *AJR Am J Roentgenol* 201:912–918. <https://doi.org/10.2214/AJR.12.9448>
 360. Vijayaraghavan SB (2006) Sonographic differential diagnosis of acute scrotum: real-time whirlpool sign, a key sign of torsion. *J Ultrasound Med* 25:563–574. <https://doi.org/10.7863/jum.2006.25.5.563>
 361. Avery LL, Scheinfeld MH (2013) Imaging of penile and scrotal emergencies. *Radiographics* 33:721–740. <https://doi.org/10.1148/r.g.333125158>
 362. Terai A, Yoshimura K, Ichioka K, Ueda N, Utsunomiya N, Kohei N, Arai Y, Watanabe Y (2006) Dynamic contrast-enhanced subtraction magnetic resonance imaging in diagnostics of testicular torsion. *Urology* 67:1278–1282. <https://doi.org/10.1016/j.urology.2005.12.021>
 363. Maki D, Watanabe Y, Nagayama M, Ishimori T, Okumura A, Amoh Y, Nakashita S, Terai A, Dodo Y (2011) Diffusion-weighted magnetic resonance imaging in the detection of testicular torsion: feasibility study. *J Magn Reson Imaging* 34:1137–1142. <https://doi.org/10.1002/jmri.22698>
 364. Watanabe Y, Nagayama M, Okumura A, Amoh Y, Suga T, Terai A, Dodo Y (2007) MR imaging of testicular torsion: features of testicular hemorrhagic necrosis and clinical outcomes. *J Magn Reson Imaging* 26:100–108. <https://doi.org/10.1002/jmri.20946>
 365. Yucel C, Ozlem Ilbey Y (2019) Predictive value of hematological parameters in testicular torsion: retrospective investigation of data from a high-volume tertiary care center. *J Int Med Res* 47:730–737. <https://doi.org/10.1177/0300060518809778>
 366. Kapoor S (2008) Testicular torsion: a race against time. *Int J Clin Pract* 62:821–827. <https://doi.org/10.1111/j.1742-1241.2008.01727.x>
 367. Mellick LB, Sinex J (2019) Testicular Torsion Pain Honeymoons. *Pediatr Emerg Care* 35:e241–e244. <https://doi.org/10.1097/PEC.0000000000001286>
 368. Mellick LB, Sinex JE, Gibson RW, Mears K (2019) A Systematic Review of Testicle Survival Time After a Torsion Event. *Pediatr Emerg Care* 35:821–825. <https://doi.org/10.1097/PEC.0000000000001287>
 369. Yagil Y, Naroditsky I, Milhem J, Leiba R, Leiderman M, Badaan S, Gaitini D (2010) Role of Doppler ultrasonography in the triage of acute scrotum in the emergency department. *J Ultrasound Med* 29:11–21. <https://doi.org/10.7863/jum.2010.29.1.11>

370. Corbett HJ, Simpson ET (2002) Management of the acute scrotum in children. *ANZ J Surg* 72:226–228. <https://doi.org/10.1046/j.1445-2197.2002.02355.x>
371. Bayne AP, Madden-Fuentes RJ, Jones EA, Cisek LJ, Gonzales ETJ, Reavis KM, Roth DR, Hsieh MH (2010) Factors associated with delayed treatment of acute testicular torsion—do demographics or interhospital transfer matter? *J Urol* 184:1743–1747. <https://doi.org/10.1016/j.juro.2010.03.073>
372. Favorito LA, Cardinot TM, Morais ARM, Sampaio FJB (2004) Urogenital anomalies in human male fetuses. *Early Hum Dev* 79:41–47. <https://doi.org/10.1016/j.earlhumdev.2004.02.004>
373. Favorito LA, Cavalcante AG, Costa WS (2004) Anatomic aspects of epididymis and tunica vaginalis in patients with testicular torsion. *Int Braz J Urol* 30:420–424. <https://doi.org/10.1590/s1677-55382004000500014>
374. Bolln C, Driver CP, Youngson GG (2006) Operative management of testicular torsion: current practice within the UK and Ireland. *J Pediatr Urol* 2:190–193. <https://doi.org/10.1016/j.jpuro.2005.07.006>
375. Molokwu CN, Somani BK, Goodman CM (2011) Outcomes of scrotal exploration for acute scrotal pain suspicious of testicular torsion: a consecutive case series of 173 patients. *BJU Int* 107:990–993. <https://doi.org/10.1111/j.1464-410X.2010.09557.x>
376. Yang C, Song B, Tan J, Liu X, Wei G (2011) Testicular torsion in children: a 20-year retrospective study in a single institution. *ScientificWorldJournal* 11:362–368. <https://doi.org/10.1100/tsw.2011.39>
377. Jaiswal V, Khadilkar V, Khadilkar A, Lohiya N (2019) Stretched Penile Length and Testicular Size from Birth to 18 Years in Boys from Western Maharashtra. *Indian J Endocrinol Metabol* 23:3–8. https://doi.org/10.4103/ijem.IJEM_242_18
378. Analia Tomova F, Deepinder R, Robeva H, Lalabonova P, Kumanov A, Agarwal (2010) Growth and Development of Male External Genitalia. *Arch Pediatr Adolesc Med* 164:1152. <https://doi.org/10.1001/archpediatrics.2010.223>
379. Schonfeld WA, Beebe GW (1942) Normal Growth and Variation in the Male Genitalia from Birth to Maturity. *J Urol* 48:759–777. [https://doi.org/10.1016/S0022-5347\(17\)70767-7](https://doi.org/10.1016/S0022-5347(17)70767-7)
380. Feldman KW, Smith DW (1975) Fetal phallic growth and penile standards for newborn male infants. *J Pediatr* 86:395–398. [https://doi.org/10.1016/S0022-3476\(75\)80969-3](https://doi.org/10.1016/S0022-3476(75)80969-3)
381. van der Zanden LFM, van Rooij IALM, Feitz WFJ, Franke B, Knoers NVAM, Roeleveld N (2012) Aetiology of hypospadias: a systematic review of genes and environment. *Hum Reprod Update* 18:260–283. <https://doi.org/10.1093/humupd/dms002>

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